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(54) Title: VERTEBRATE PATCHED-2 PROTEIN

(57) Abstract

The present invention relates to nucleotide sequences, including expressed sequence tags (ESTs), oligonucleotide probes, polypeptides, antibodies, vectors and host cells expressing, immunoadhesins, agonists and antagonists to *patched*-2.

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VERTEBRATE PATCHED-2 PROTEIN

FIELD OF THE INVENTION

The present invention relates generally to signaling molecules, specifically to signaling and mediator molecules in the hedgehog (*hh*) cascade which are involved in cell proliferation and differentiation.

BACKGROUND OF THE INVENTION

Development of multicellular organisms depends, at least in part, on mechanisms which specify, direct or maintain positional information to pattern cells, tissues, or organs. Various secreted signaling molecules, such as members of the transforming growth factor-beta (TGF-β), Wnt, fibroblast growth factors and hedgehog families have been associated with patterning activity of different cells and structures in *Drosophila* as well as in vertebrates. Perrimon, *Cell*: 80: 517-520 (1995).

Segment polarity genes were first discovered in *Drosophila*, which when mutated caused a change in the pattern of structures of the body segments. These changes affected the pattern along the head to tail axis. Hedgehog (*Hh*) was first identified as a segment-polarity gene by a genetic screen *in Drosophila melanogaster*, Nusslein-Volhard *et al., Roux. Arch. Dev. Biol.* 193: 267-282 (1984), that plays a wide variety of developmental functions. . Perrimon, *supra*. Although only one Drosophila *Hh* gene has been identified, three mammalian *Hh* homologues have been isolated: Sonic *Hh (Shh)*, Desert *Hh (Dhh)* and *Indian Hh (Ihh)*, Echelard *et al., Cell* 75: 1417-30 (1993); Riddle *et al., Cell* 75: 1401-16 (1993). *Shh* is expressed at high level in the notochord and floor plate of developing vertebrate embryos, and acts to establish cell fate in the developing limb, somites and neural tube. *In vitro* explant assays as well as ectopic expression of *Shh* in transgenic animals show that *SHh* plays a key role in neural tube patterning, Echelard *et al.* (1993), *supra.*; Ericson *et al., Cell* 81: 747-56 (1995); Marti *et al., Nature* 375: 322-5 (1995); Roelink et al. (1995), *supra*; Hynes *et al., Neuron* 19: 15-26 (1997). *Hh* also plays a role in the development of limbs (Krauss *et al., Cell* 75: 1431-44 (1993); Laufer *et al., Cell* 79, 993-1003 (1994)), somites (Fan and Tessier-Lavigne, *Cell* 79, 1175-86 (1994); Johnson *et al., Cell* 79: 1165-73 (1994)), lungs (Bellusci *et al., Develop.* 124: 53-63 (1997) and skin (Oro *et al., Science* 276: 817-21 (1997). Likewise, *Ihh* and *Dhh* are involved in bone, gut and germinal cell development, Apelqvist *et al., Curr. Biol.* 7: 801-4 (1997); Bellusci *et al., Development* 124: 53-63 (1997); Bitgood *et al., Curr. Biol.* 6: 298-304 (1996); Roberts *et al., Development* 121: 3163-74 (1995). Specifically, *Ihh* has been implicated in chondrocyte development [Vortkamp, A. *et al., Science* 273: 613-22 (1996)] while *Dhh* plays a key role in testis development. Bitgood *et al., supra*. With the exception of the gut, in which both *Ihh* and *Shh* are expressed, the expression patterns of the hedgehog family members do not overlap. Bitgood *et al., supra*.

At the cell surface, Hh function appears to be mediated by a multicomponent receptor complex involving patched (*ptch*) and smoothened (*smo*), two multi-transmembrane proteins initially identified as segment polarity genes in *Drosophila* and later characterized in vertebrates. Nakano *et al., Nature* 341: 508-513 (1989); Goodrich *et al., Genes Dev.* 10: 301-312 (1996); Marigo *et al., Develop.* 122: 1225-1233 (1996); van den Heuvel, M. & Ingham, P.W., *Nature* 382: 547-551 (1996); Alcedo, J. *et al., Cell* 86: 221-232 (1996); Stone, D.M. *et al., Nature* 384: 129-34 (1996). Upon binding of *Hh* to *Ptch*, the normal inhibitory effect of *Ptch* on *Smo* is relieved, allowing *Smo* to transduce the *Hh* signal across the plasma membrane. It

remains to be established if the *Ptch/Smo* receptor complex mediates the action of all 3 mammalian hedgehogs or if specific components exist. Interestingly, a second murine *Ptch* gene, *Ptch-2* was recently isolated [Motoyama, J. *et al., Nature Genetics* 18: 104-106 (1998)], but its function as a *Hh* receptor has not been established. In order to characterize *Ptch*-2 and compare it to *Ptch* with respect to the biological function of the various *Hh* family members, Applicants have isolated the human *Ptch*-2 gene. Biochemical analysis of *Ptch* and *Ptch*-2 show that both bind to all members of the *Hh* family with similar affinity and that both molecules can form a complex with *Smo*. However, the expression patterns of *Ptch*-2 and *Ptch* do not overlap. While *Ptch* is expressed throughout the mouse embryo, *Ptch*-2 is found mainly in spermatocytes which require Desert Hedgehog (*Dhh*) for proper development suggesting that *Ptch*-2 mediates *Dhh's* activity in the testis. Chromosomal localization of *Ptch*-2 places it on chromosome 1p33-34, a region deleted in some germ cell tumors, raising the possibility that *Ptch*-2 may be a tumor suppressor in *Dhh* target cells.

SUMMARY OF THE INVENTION

In one embodiment, the invention provides an isolated nucleic acid molecule having at least about 80% sequence identity to (a) a DNA molecule encoding a patched-2 polypeptide comprising the sequence of amino acids 1 to 1203 of Fig. 1, or (b) the complement of the DNA molecule of (a); and encoding a polypeptide having *patched*-2 biological activity. The sequence identity preferably is > 91%, more preferably about 92%, most preferably about 95%. In one aspect, the isolated nucleic acid has at least > 91%, preferably at least about 92%, and even more preferably at least about 95% sequence identity with a polypeptide having amino acid residues 1 to about 1203 of Fig. 1. In a further aspect, the isolated nucleic acid molecule comprises DNA encoding a human *patched-2* polypeptide having amino acid residues 1 to about 1203 of Fig. 1. In yet another aspect, the invention provides for an isolated nucleic acid comprising DNA having at least a 95% sequence identity to (a) a DNA molecule encoding the same mature polypeptide encoded by the cDNA in ATCC Deposit No. 209778 (designation: pRK7.hptc2.Flag-1405), alternatively the coding sequence of clone pRK7.hptc2.Flag-1405, deposited under accession number ATCC 209778. In a still further aspect, the invention provides for a nucleic acid comprising human patched-2 encoding sequence of the cDNA in ATCC deposit No. 209778 (designation: pRK7.hptc2.Flag-1405) or a sequence which hybridizes thereto under stringent conditions.

In another embodiment, the invention provides a vector comprising DNA encoding a human *patched-2* polypeptide. A host cell comprising such a vector is also provided. By way of example, the host cells may be mammalian cells, (e.g. CHO cells), prokaryotic cells (e.g., *E. coli*) or yeast cells (e.g., *Saccharomyces cerevisiae*). A process for producing *patched-2* polypeptides is further provided and comprises culturing host cells under conditions suitable for expression of *patched-2* and recovering the same from the cell culture.

In yet another embodiment, the invention provides an isolated *patched*-2 polypeptide. In particular, the invention provides isolated native sequence *patched*-2 polypeptide, which in one embodiment is a human patched-2 including an amino acid sequence comprising residues 1 to about 1203 of Figure 1. Human *patched*-2 polypeptides with or without the initiating methionine are specifically included. Alternatively, the invention provides a human *patched-2* polypeptide encoded by the nucleic acid deposited under accession

number ATCC Deposit No. 209778.

In yet another embodiment, the invention provides chimeric molecules comprising a *patched-2* polypeptide patched-2 to a heterologous polypeptide or amino acid sequence. An example of such a chimeric molecule comprises a *patched-2* polypeptide *patched*-2 to an epitope tag sequence or a constant region of an immunoglobulin.

In yet another embodiment, the invention provides expressed sequence tag (EST) comprising the nucleotide sequences identified in Fig. 2A (905531) (SEQ ID NO:3) and Fig. 2B (1326258) (SEQ ID NO:5).

In yet another embodiment, the invention provides for alternatively spliced variants of human *patched-2* having patched-2 biological activity.

In yet another embodiment, the invention provides for method of using *patched*-2 for the treatment of disorders which are mediated at least in part by Hedgehog (Hh), especially Desert hedgehog (*Dhh*). In particular, testicular cancer. In yet another embodiment, the invention provides a method of using antagonists or agonists of *patched*-2 for treating disorders or creating a desirable physiological condition effected by blocking *Hh* signaling, especially *Dhh* signaling. (*E.g*, contraception).

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the nucleotide (SEQ ID NO:1) and derived amino acid (SEQ ID NO:2) sequence of a native sequence of human P*tch*-2.

Figure 2A shows EST 905531 (SEQ ID NO:3) and Fig. 2B shows EST 1326258 (SEQ ID NO:5) in alignment with human *Ptch* (SEQ ID NO:18). These ESTs were used in the cloning of human full-length *Ptch*-2 (SEQ ID NO:1).

Figure 3 shows a comparison between human *Ptch* (SEQ ID NO:4) and *Ptch*-2 (SEQ ID NO:2). Gaps introduced for optimal alignment are indicated by dashes. Identical amino acids are boxed. The 12 transmembrane domains are indicated by the gray boxes, all of which are conserved between the two sequences. Alignment results between the two sequences indicate 53% identity. The most significant difference is a shorter C-terminal intracellular domain in human *Ptch*-2 (SEQ ID NO:2) in comparison with human *Ptch* (SEQ ID NO:4).

Figure 4 shows a northern blot of *Ptch*-2 (SEQ ID NO:2) which indicates expression is limited to the testis. Multiple human fetal and adult tissue northern blots were probe fragments corresponding to the 3'-untranslated region of murine Ptch-2.

Figure 5 shows a chromosomal localization of two BAC clones which were isolated by PCR screening with human patched-2 derived probes. Both probes were mapped by FISH to human chromosome 1p33-34.

Figure 6 is an *in situ* hybridization comparing *Ptch* (SEQ ID NO:4), *Ptch*-2 (SEQ ID NO:2) and *Fused* (FuRK) (SEQ ID NO:10) expression. High magnification of mouse testis showing expression of (a) *Ptch*, *Ptch*-2 (SEQ ID NO:2) (b) and FuRK (SEQ ID NO:10) (c). Low magnification of testis section hybridized with *Ptch*-2 sense (SEQ ID NO:11) (d) and anti-sense probe (SEQ ID NO:12) (e) respectively. Fig. 6(f) shows low magnification of testis section hybridized with FuRK (SEQ ID NO:10 encoding nucleic acid). Scale bar: a, b, c: 0.05 mm; d, e, f: 0.33 mm.

Figure 7A is logarithmic plot comparing the binding *Ptch*-2 (SEQ ID NO:2) to *Dhh* (SEQ ID

NO:13) and *Shh* (SEQ ID NO:14). Competitive binding of recombinant murine ^{125}I-*Shh* to 293 cells overexpressing *Ptch* (SEQ ID NO:4) or *Ptch-2* (SEQ ID NO:2). There was no detectable binding to mock transfected cells (data not shown). Figure 7B is a western blot illustrating co-immunoprecipitation of epitope tagged *Ptch* (SEQ ID NO:4) or *Ptch*-2 (SEQ ID NO:2) with epitope tagged *Smo* (SEQ ID NO:15). Immunoprecipitation was performed with antibodies to the Flag tagged *Ptch* (SEQ ID NO:4) and analyzed on a 6% acrylamide gel with antibodies to the Myc tagged *Smo* (SEQ ID NO:15). Protein complexes can be detected for both *Ptch* (SEQ ID NO:4) and *Ptch*-2 (SEQ ID NO:2) with *Smo* (SEQ ID NO:15) *Ptch* (SEQ ID NO:4) and *Ptch*-2 (SEQ ID NO:2) express at similar levels as shown by immunoprecipitation using antibodies to the Flag-tag and western blot using the same anti-Flag antibody.

Figure 8 is a sequence comparison between human *Ptch*-2 (SEQ ID NO:2) and murine *Ptch*-2 (SEQ ID NO:7). which indicates that there is about 91% identity between the two sequences.

Figure 9 is an *in situ* hybridization which demonstrates the accumulation of *Ptch* (SEQ ID NO:4) and

Ptch-2 (SEQ ID NO:2) mRNA detected by in situ hybridization in basal cells of E18 transgenic mice overexpressing *SMO*-M2 (SEQ ID NO:16) (Xie *et al.*, *Nature* 391: 90-92 (1998).

Figure 10 is a partial sequence representing clone 3A (SEQ ID NO:8). a partial *patched*-2 fragment which was initially isolated from a fetal brain library.

Figure 11 is a partial sequence representing clone 16.1 (SEQ ID NO:9). a partial *patched*-2 fragment which isolated from a testis library.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

I. Definitions

The terms "*patched-2*" and "*patched-2* polypeptide" when used herein encompass native sequence *patched-2* and *patched-2* variants (which are further defined herein) having *patched-2* biological activity. *Patched-2* may be isolated from a variety of sources, such as from testes tissue types or from another source, or prepared by recombinant or synthetic methods.

A "native sequence *patched-2*" comprises a polypeptide having the same amino acid sequence as a human *patched-2* derived from nature. Such native sequence *patched-2* can be isolated from nature or can be produced by recombinant and/or synthetic means. The term "native sequence vertebrate *patched-2*" specifically encompasses naturally occurring truncated forms of human *patched-2*, naturally occurring variant forms (*e.g.*, alternatively spliced forms) and naturally-occurring allelic variants of human *patched-2*. Thus, one embodiment of the invention, the native sequence *patched-2* is a mature or full-length native *Ptch-2* comprising amino acids 1 to 1203 of Fig. 1 (SEQ ID NO:2) with or without the initiating methionine at position 1.

"*Patched-2* variant" means an active human *patched-2* as defined below having at least > 91% amino acid sequence identity to (a) a DNA molecule encoding a *patched-2* polypeptide. or (b) the complement of the DNA molecule of (a). In a particular embodiment, the *patched-2* variant has at least > 91% amino acid sequence homology with the human *Ptch-2* (SEQ ID NO:2) having the deduced amino acid sequence shown in Fig. 1 for a full-length native sequence human *patched-2*. Such *patched-2* variants

include, without limitation, *patched-2* polypeptides wherein one or more amino acid residues are added, or deleted, at the N- or C-terminus of the sequence of Fig. 1 (SEQ ID NO:2). Preferably, the nucleic acid or amino acid sequence identity is at least about 92%, more preferably at least about 93%, and even more preferably at least about 95%.

"Percent (%) amino acid sequence identity" with respect to the *patched-2* sequences identified herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the *patched-2* sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST-2 software that are set to their default parameters. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared.

"Percent (%) nucleic acid sequence identity" with respect to the *patched-2* sequences identified herein is defined as the percentage of nucleotides in a candidate sequence that are identical with the nucleotides in the *patched-2* sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining percent nucleic acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST-2 software that are set to their default parameters. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared.

The term "epitope tagged" when used herein refers to a chimeric polypeptide comprising *patched-2* polypeptide, or a portion thereof, *patched*-2 to a "tag polypeptide". The tag polypeptide has enough residues to provide an epitope against which an antibody can be made, yet is short enough such that it does not interfere with activity of the *patched-2* polypeptide. The tag polypeptide preferably also is fairly unique so that the antibody does not substantially cross-react with other epitopes. Suitable tag polypeptides generally have at least six amino acid residues and usually between about 8 to about 50 amino acid residues (preferably, between about 10 to about 20 residues).

As used herein, the term "immunoadhesin" designates antibody-like molecules which combine the binding specificity of a heterologous protein (an "adhesin") with the effector functions of immunoglobulin constant domains. Structurally, the immunoadhesin comprise a fusion of an amino acid sequence with the desired binding specificity which is other than the antigen recognition and binding site of an antibody (i.e., is "heterologous"), and an immunoglobulin constant domain sequence. The adhesin part of an immunoadhesin molecule typically is a contiguous amino acid sequence comprising at least the binding site of a receptor or a ligand. The immunoglobulin constant domain sequence in the immunoadhesins may be obtained from any immunoglobulin, such as IgG-1, IgG-2, IgG-3 or IgG-4 subtypes, IgA (including IgA-1 and IgA-2, IgE, IgD or IgM. Immunoadhesion reported in the literature include fusions of the T cell receptor* [Gascoigne *et al.*, *Proc. Natl. Acad. Sci. USA* 84: 2936-2940 (1987)]; CD4* [Capron *et al.*, *Nature* 337: 525-531 (1989); Traunecker *et al.*, *Nature* 339: 68-70 (1989); Zettmeissl *et al.*, *DNA Cell Biol. USA* 9: 347-353 (1990); Byrn

et al.. *Nature* 344, 667-670 (1990)]; L-selectin (homing receptor) [Watson *et al.*. *J. Cell. Biol.* 110, 2221-2229 (1990); Watson *et al.*, *Nature* 349, 164-167 (1991)]; CD44* [Aruffo *et al.*, *Cell* 61, 1303-1313 (1990)]; CD28* and B7* [Linsley *et al.*, *J. Exp. Med.* 173, 721-730 (1991)]; CTLA-4* [Lisley *et al.*. *J. Exp. Med.* 174, 561-569 (1991)]; CD22* [Stamenkovic *et al.*, *Cell* 66. 1133-1144 (1991)]; TNF receptor [Ashkenazi *et al.*, *Proc. Natl. Acad. Sci. USA* 88, 10535-10539 (1991); Lesslauer *et al.*, *Eur. J. Immunol.* 27, 2883-2886 (1991); Peppel *et al.*, *J. Exp. Med.* 174, 1483-1489 (1991)]; NP receptors [Bennett *et al.*. J. Biol. Chem. 266, 23060-23067 (1991)]; IgE receptor α-chain* [Ridgway and Gorman, J. Cell. Biol. 115, abstr. 1448 (1991)]; HGF receptor [Mark, M.R. *et al.*, *J. Biol. Chem.*, 267(36): 26166-26171 (1992)], where the asterisk (*) indicates that the receptor is a member of the immunoglobulin superfamily.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends upon the ability of denatured DNA to reanneal when complementary strands are present in an environment near but below their T^m (melting temperature). The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature which can be used. As a result, it follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. Moreover, stringency is also inversely proportional to salt concentrations. For additional details and explanation of stringency of hybridization reactions, see Ausubel *et al.*. *Current Protocols in Molecular Biology* (1995).

"Stringent conditions," as defined herein may be identified by those that: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42°C; (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 μg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) and 50% formamide at 55°C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C.

"Isolated," when used to describe the various polypeptides disclosed herein, means polypeptide that has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous solutes. In preferred embodiments, the polypeptide will be purified (1) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (2) to homogeneity by SDS-PAGE under non-reducing or reducing conditions using Coomassie blue or, preferably, silver stain. Isolated polypeptide includes polypeptide *in situ* within recombinant cells, since at least one component of the vertebrate *patched-2* natural environment will not be present. Ordinarily, however, isolated polypeptide will be prepared by at least one purification step.

An "isolated" *patched-2* nucleic acid molecule is a nucleic acid molecule that is identified and separated from at least one contaminant nucleic acid molecule with which it is ordinarily associated in the natural source of the *patched-2* nucleic acid. An isolated *patched-2* nucleic acid molecule is other than in the form or setting in which it is found in nature. Isolated *patched-2* nucleic acid molecules therefore are distinguished from the corresponding native *patched-2* nucleic acid molecule as it exists in natural cells.

The term "control sequences" refers to DNA sequences necessary for the expression of an operably linked coding sequence in a particular host organism. The control sequences that are suitable for prokaryotes, for example, include a promoter, optionally an operator sequence, and a ribosome binding site. Eukaryotic cells are known to utilize promoters, polyadenylation signals, and enhancers.

Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading frame. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

The term "antibody" is used in the broadest sense and specifically covers single monoclonal antibodies (including agonist and antagonist antibodies), antibody compositions with polyepitopic specificity, as well as antibody fragments (e.g., Fab, $F(ab')_2$ and Fv), so long as they exhibit the desired biological activity.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations that typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by the hybridoma culture, uncontaminated by other immunoglobulins. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler & Milstein, Nature 256:495 (1975), or may be made by recombinant DNA methods [see, e.g. U.S. Patent No. 4,816,567 (Cabilly *et al.*)].

The monoclonal antibodies herein specifically include "chimeric" antibodies (immunoglobulins) in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while

the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass. as well as fragments of such antibodies, so long as they exhibit the desired biological activity [U.S. Patent No. 4,816,567; Cabilly *et al.*; Morrison *et al.*, Proc. Natl. Acad. Sci. USA 81, 6851-6855 (1984)].

"Humanized" forms of non-human (e.g. murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', $F(ab')_2$ or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, corresponding non-human residues replace Fv framework residues of the human immunoglobulin. Furthermore, humanized antibody may comprise residues that are found neither in the recipient antibody nor in the imported CDR or framework sequences. These modifications are made to further refine and optimize antibody performance. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. For further details see: Jones *et al.*, Nature 321, 522-525 (1986); Riechmann *et al.*, Nature 332, 323-327 (1988); Presta, Curr. Op. Struct. Biol. 2 593-596 (1992) and U.S. Patent No. 5,225,539 (Winter) issued July 6, 1993.

"Active" or "activity" for the purposes herein refers to form(s) of *patched-2* which retain the biologic and/or immunologic activities of native or naturally occurring *patched-2*. A preferred activity is the ability to bind to and affect, *e.g.*, block or otherwise modulate, *hedgehog (Hh)*, especially *desert hedgehog (Dhh)* signaling. For example, the regulation of the pathogenesis of testicular cancer, male spermatocyte formation and basal cell carcinoma.

The term "antagonist" is used herein in the broadest sense to include any molecule which blocks, prevents, inhibits, neutralizes the normal functioning of *patched-2* in the *hedgehog (Hh)* signaling pathway. One particular form of antagonist includes a molecule that interferes with the interaction between *Dhh* and *patched*-2. Alternatively, an antagonist could also be a molecule which increases the levels of *patched-2*. In a similar manner, the term "agonist" is used herein to include any molecule which promotes, enhances or stimulates the binding of a *Hh* to *patched-2* in the *Hh* signaling pathway or otherwise upregulates it (e.g., blocking binding of *Ptch-2* (SEQ ID NO:2) to *Smo* (SEQ ID NO:17). Suitable molecules that affect the protein-protein interaction of *Hh* and *patched-2* and its binding proteins include fragments of the latter or small bioorganic molecules, *e.g.*, peptidomimetics, which will prevent or enhance, as the case may be, the binding of *Hh* to *patched*-2. Non-limiting examples include proteins, peptides, glycoproteins, glycopeptides, glycolipids, polysaccharides, oligosaccharides, nucleic acids, bioorganic molecules, peptidomimetics, pharmacological agents and their metabolites, transcriptional and translation control sequences, and the like.

Another preferred form of antagonist includes antisense oligonucleotides that inhibit proper transcription of wild type *patched-2*.

The term "modulation" or "modulating" means upregulation or downregulation of a signaling pathway. Cellular processes under the control of signal transduction may include, but are not limited to, transcription of specific genes; normal cellular functions, such as metabolism, proliferation, differentiation, adhesion, apoptosis and survival, as well as abnormal processes, such as transformation, blocking of differentiation and metastasis.

The techniques of "polymerase chain reaction," or "PCR", as used herein generally refers to a procedure wherein minute amounts of a specific piece of nucleic acid, RNA and/or DNA are amplified as described in U.S. Pat. No. 4,683,195 issued 28 July 1987. Generally, sequence information from the ends of the region of interest or beyond needs to be available, such that oligonucleotide primers can be designed; these primers will be identical or similar in sequence to opposite strands of the template to be amplified. The 5' terminal nucleotides of the two primers may coincide with the ends of the amplified material. PCR sequences form total genomic DNA, and cDNA transcribed from total cellular RNA, bacteriophage, or plasmid sequences, etc. See generally Mullis *et al.*, *Cold Spring Harbor Symp. Quant. Biol.* 51: 263 (1987); Erlich, Ed., PCR Technology, (Stockton Press, NY, 1989). As used herein, PCR is considered to be one, but not the only, example of a nucleic acid test sample comprising the use of a known nucleic acid as a primer and a nucleic acid polymerase to amplify or generate a specific piece of nucleic acid.

II. Compositions and Methods of the Invention

A. Full-length *patched-2*

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as *patched-2*. In particular, Applicants have identified and isolated cDNA encoding a human *patched-2* polypeptide, as disclosed in further detail in the Examples below. Using BLAST, BLAST-2 and FastA sequence alignment computer programs (set to the default parameters), Applicants found that a full-length native sequence human *patched-2* (i.e., *Ptch-2* in Figure 3, SEQ ID NO:2) has 53% amino acid sequence identity with a human *patched* (i.e., *Ptch*, SEQ ID NO:4). Moreover a human full-length *patched*-2 (i.e., *Ptch-2*, SEQ ID NO:2) has about a 91% sequence identity with murine *Ptch*-2 (SEQ ID NO:7) (Fig. 8). Accordingly, it is presently believed that the human *patched-2* (i.e., *Ptch-2*, SEQ ID NO:2) disclosed in the present application is a newly identified member of the mammalian hedgehog signaling cascade, specifically *Desert hedgehog*.

The full-length native sequence of human *patched-2* gene, or portions thereof, may be used as hybridization probes for a cDNA library to isolate the full-length gene or to isolate still other vertebrate homolog genes (for instance, those encoding naturally-occurring variants of *patched-2* or *patched-2* from other species) which have a desired sequence identity to the human *patched-2* sequence disclosed in Fig.1 (SEQ ID NO:2). Optionally, the length of the probes will be about 20 to about 50 bases. The hybridization probes may be derived from the nucleotide sequence of Fig. 1 (SEQ ID NO:1) or from genomic sequences including promoters, enhancer elements and introns of native sequence vertebrate *patched-2*. By way of example, a screening method will comprise isolating the coding region of the vertebrate *patched-2* gene using the known DNA sequence to synthesize a selected probe of about 40 bases. Hybridization probes may

be labeled by a variety of labels. including radionucleotides such as ^{32}P or ^{35}S. or enzymatic labels such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems. Labeled probes having a sequence complementary to that of the vertebrate *patched-2* gene of the present invention can be used to screen libraries of human cDNA, genomic DNA or mRNA to determine which members of such libraries the probe hybridizes to.

B. *Patched-2* Variants

In addition to the full-length native sequence *patched-2* described herein, it is contemplated that *patched-2* variants can be prepared. *Patched-2* variants can be prepared by introducing appropriate nucleotide changes into a known *patched-2* DNA, or by synthesis of the desired *patched-2* polypeptides. Those skilled in the art will appreciate that amino acid changes may alter post-translational processes of *patched-2*.

Variations in the native full-length sequence *patched-2* or in various domains of the *patched-2* described herein, can be made. for example, using any of the techniques and guidelines for conservative and non-conservative mutations set forth. for instance. in U.S. Patent No. 5,364,934. Variations may be a substitution. deletion or insertion of one or more codons encoding the *patched-2* that results in a change in the amino acid sequence of *patched-2* as compared with the native sequence *patched-2*. Optionally, the variation is by substitution of at least one amino acid with any other amino acid in one or more of the domains of *patched-2*. Guidance in determining which amino acid residue may be inserted. substituted or deleted without adversely affecting the desired activity may be found by comparing the sequence of the *patched-2* with that of homologous known protein molecules and minimizing the number of amino acid sequence changes made in regions of high homology. Amino acid substitutions can be the result of replacing one amino acid with another amino acid having similar structural and/or chemical properties. such as the replacement of a leucine with a serine, i.e., conservative amino acid replacements. Insertions or deletions may optionally be in the range of 1 to 5 amino acids. The variation allowed may be determined by systematically making insertions. deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity in the *in vitro* assay described in the Examples below.

The variations can be made using methods known in the art such as oligonucleotide-mediated (site-directed) mutagenesis. alanine scanning, and PCR mutagenesis. Site-directed mutagenesis [Carter et al., Nucl. Acids Res., 13:4331 (1986); Zoller et al., Nucl. Acids Res., 10: 6487 (1987)], cassette mutagenesis [Wells et al., Gene, 34:315 (1985)], restriction selection mutagenesis [Wells et al., Philos. Trans. R. Soc. London SerA, 317:415 (1986)] or other known techniques can be performed on the cloned DNA to produce the vertebrate *patched-2* variant DNA.

Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous sequence. Among the preferred scanning amino acids are relatively small. neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant. Alanine is also typically preferred because it is the most common amino acid. Further. it is frequently found in both buried and exposed positions [Creighton, The

Proteins. (W.H. Freeman & Co., N.Y.); Chothia. *J. Mol. Biol.*, 150:1 (1976)]. If alanine substitution does not yield adequate amounts of variant, an isoteric amino acid can be used.

In the comparison between human *patched* and *patched*-2 sequences depicted in Figure 3 (e.g., *Ptch*, SEQ ID NO:4 and *Ptch-2*. SEQ ID NO:2), the 12 transmembrane domains are identified in gray, while identical residues are boxed. Gaps are indicated by dashes (-) and are inserted to maximize the total identity score between the two sequences.

C. Modifications of *patched-2*

Covalent modifications of *patched-2* are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of *patched-2* with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C- terminal residues of the *patched-2*. Derivatization with bifunctional agents is useful, for instance. for crosslinking *patched-2* to a water-insoluble support matrix or surface for use in the method for purifying anti-*patched-2* antibodies, and vice-versa. Commonly used crosslinking agents include, *e.g.*, 1,1-bis(diazo-acetyl)-2-phenylethane, glutaraldehyde, N-hydroxy-succinimide esters, for example, esters with 4-azido-salicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis-(succinimidyl-propionate), bifunctional maleimides such as bis-N-maleimido-1,8-octane and agents such as methyl-3-[(p-azidophenyl)-dithio]proprioimidate.

Other modifications include deamidation of glutaminyl and asparaginyl residues to the corresponding glutamyl and aspartyl residues, respectively, hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α-amino groups of lysine, arginine, and histidine side chains [T.E. Creighton, Proteins: Structure and Molecular Properties, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)], acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

Another type of covalent modification of *patched-2* comprises linking the *patched-2* polypeptide to one of a variety of nonproteinaceous polymers, *e.g.*, polyethylene glycol. polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337. Such modifications would be expected in increase the half-life of the molecules in circulation in a mammalian system; Extended half-life of *patched-2* molecules might be useful under certain circumstances, such as where the patched-2 variant is administered as a therapeutic agent.

The *patched-2* of the present invention may also be modified in a way to form a chimeric molecule comprising *patched-2* bonded to another, heterologous polypeptide or amino acid sequence. In one embodiment, such a chimeric molecule comprises a fusion of *patched-2* with a tag polypeptide, which provides an epitope to which an anti-tag antibody can selectively bind. The epitope tag is generally placed at the amino- or carboxyl- terminus of the *patched-2*. The presence of such epitope-tagged forms of the *patched-2* can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables the *patched-2* to be readily purified by affinity purification using an anti-tag antibody or another type of affinity matrix that binds to the epitope tag. In an alternative embodiment, the chimeric molecule may comprise a fusion of the *patched-2* with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule, such a fusion could be to the Fc region of an IgG molecule.

Ordinarily, the C-terminus of a contiguous amino acid sequence of a patched-2 receptor is fused to the N-terminus of a contiguous amino acid sequence of an immunoglobulin constant region, in place of the variable region(s), however N-terminal fusions are also possible.

Typically, such fusions retain at least functionally active hinge, CH2 and CH3 domains of the constant region of an immunoglobulin heavy chain. Fusions are also made to the C-terminus of the Fc portion of a constant domain, or immediately N-terminal to the CH1 of the heavy chain or the corresponding region of the light chain. This ordinarily is accomplished by constructing the appropriate DNA sequence and expressing it in recombinant cell culture. Alternatively, immunoadhesins may be synthesized according to known methods.

The precise site at which the fusion is made is not critical; particular sites are well known and may be selected in order to optimize the biological activity, secretion or binding characteristics of the immunoadhesins.

In a preferred embodiment, the C-terminus of a contiguous amino acid sequence which comprises the binding site(s) of *patched*-2, at the N-terminal end, to the C-terminal portion of an antibody (in particular the Fc domain), containing the effector functions of an immunoglobulin, e.g. immunoglobulin G_1 (IgG-1). As herein above mentioned, it is possible to fuse the entire heavy chain constant region to the sequence containing the binding site(s). However, more preferably, a sequence beginning in the hinge region just upstream of the papain cleavage site (which defines IgG Fc chemically; residue 216, taking the first residue of heavy chain constant region to be 114 [Kobat *et al.*, supra], or analogous sites of other immunoglobulins) is used in the fusion. Although it was earlier thought that in immunoadhesins the immunoglobulin light chain would be required for efficient secretion of the heterologous protein-heavy chain fusion proteins, it has been found that even the immunoadhesins containing the whole IgG1 heavy chain are efficiently secreted in the absence of light chain. Since the light chain is unnecessary, the immunoglobulin heavy chain constant domain sequence used in the construction of the immunoadhesins of the present invention may be devoid of a light chain binding site. This can be achieved by removing or sufficiently altering immunoglobulin heavy chain sequence elements to which the light chain is ordinarily linked so that such binding is no longer possible. Thus, the CH1 domain can be entirely removed in certain embodiments of the *patched*-2/immunoglobulin chimeras.

In a particularly preferred embodiment, the amino acid sequence containing the extracellular domain(s) of *patched*-2 is fused to the hinge region and CH2, CH3; or CH1, hinge, CH2 and CH3 domains of an IgG-1, IgG-2, IgG-3, or IgG-4 heavy chain.

In some embodiments, the *patched*-2/immunoglobulin molecules (immunoadhesins) are assembled as monomers, dimers or multimers, and particularly as dimers or tetramers. Generally, these assembled immunoadhesins will have known unit structures similar to those of the corresponding immunoglobulins. A basic four chain structural unit (a dimer of two immunoglobulin heavy chain-light chain pairs) is the form in which IgG, IgA and IgE exist. A four chain unit is repeated in the high molecular weight immunoglobulins; IgM generally exists as a pentamer of basic four-chain units held together by disulfide bonds. IgA globulin, and occasionally IgG globulin, may also exist in a multimeric form in serum. In the case of multimers, each four chain unit may be the same or different.

It is not necessary that the entire immunoglobulin portion of the *patched*-2/immunoglobulin chimeras be from the same immunoglobulin. Various portions of different immunoglobulins may be combined, and

variants and derivatives of native immunoglobulins can be made as herein above described with respect to *patched*-2. in order to optimize the properties of the immunoadhesin molecules. For example, immunoadhesin constructs in which the hinge of IgG-1 was replaced with that of IgG-3 were found to be functional and showed pharmacokinetics comparable to those of immunoadhesins comprising the entire IgG-1 heavy chain.

Various tag polypeptides and their respective antibodies are well known in the art. Examples include poly-histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags; the flu HA tag polypeptide and its antibody 12CA5 [Field et al., Mol. Cell. Biol., 8: 2159-2165 (1988)]; the c-myc tag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibodies thereto [Evan et al., Molecular and Cellular Biology, 5:3610-3616 (1985)]: and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody [Paborsky et al., Protein Engineering, 3(6):547-553 (1990)]. Other tag polypeptides include the Flag-peptide [Hopp et al., BioTechnology, 6:1204-1210 (1988)]; the KT3 epitope peptide [Martin et al., Science, 255:192-194 (1992)]; an α-tubulin epitope peptide [Skinner et al., J. Biol. Chem., 266:15163-15166 (1991)]; and the T7 gene 10 protein peptide tag [Lutz-Freyermuth et al., Proc. Natl. Acad. Sci. USA, 87:6393-6397 (1990)]. A preferred tag is the influenza HA tag.

D. Preparation of *patched-2*

The description below relates primarily to production of a particular *patched-2* by culturing cells transformed or transfected with a vector containing *patched-2* nucleic acid. It is, of course, contemplated that alternative methods. which are well known in the art, may be employed to prepare *patched 2*. For instance. the *patched-2* sequence, or portions thereof, may be produced by direct peptide synthesis using solid-phase techniques [see, *e.g.*, Stewart et al., Solid-Phase Peptide Synthesis, W.H. Freeman Co., San Francisco, CA (1969): Merrifield, J. Am. Chem. Soc., 85:2149-2154 (1963)]. *In vitro* protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be accomplished, for instance, using an Applied Biosystems Peptide Synthesizer (Foster City, CA) using manufacturer's instructions. Various portions of the vertebrate *patched-2* may be chemically synthesized separately and combined using chemical or enzymatic methods to produce the full-length *patched-2*.

1. Isolation of DNA encoding vertebrate *patched-2*

DNA encoding *patched-2* may be obtained from a cDNA library prepared from tissue believed to possess the *patched*-2 mRNA and to express it at a detectable level. Accordingly, human *patched*-2 DNA can be conveniently obtained from a cDNA library prepared from human tissue, such as described in the Examples. The vertebrate patched-2-encoding gene may also be obtained from a genomic library or by oligonucleotide synthesis.

Libraries can be screened with probes (such as antibodies to the *patched-2* or oligonucleotides of at least about 20-80 bases) designed to identify the gene of interest or the protein encoded by it. Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the gene encoding vertebrate *patched-2* is to use PCR methodology [Sambrook et al., supra; Dieffenbach et al., PCR Primer: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1995)].

The Examples below describe techniques for screening a cDNA library. The oligonucleotide

sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized. The oligonucleotide is preferably labeled such that it can be detected upon hybridization to DNA in the library being screened. Methods of labeling are well known in the art. and include the use of radiolabels like ^{32}P-labeled ATP, biotinylation or enzyme labeling. Hybridization conditions, including moderate stringency and high stringency, are provided in Sambrook *et al., supra.*

Sequences identified in such library screening methods can be compared and aligned to other known sequences deposited and available in public databases such as GenBank or other private sequence databases. Sequence identity (at either the amino acid or nucleotide level) within defined regions of the molecule or across the full-length sequence can be determined through sequence alignment using computer software programs such as BLAST, BLAST-2, ALIGN, DNAstar, and INHERIT which employ various algorithms to measure homology.

Nucleic acid having protein coding sequence may be obtained by screening selected cDNA or genomic libraries using the deduced amino acid sequence disclosed herein for the first time, and, if necessary, using conventional primer extension procedures as described in Sambrook *et al., supra*, to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

2. Selection and Transformation of Host Cells

Host cells are transfected or transformed with expression or cloning vectors described herein for *patched-2* production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in *Mammalian Cell Biotechnology: A Practical Approach*, M. Butler, ed. (IRL Press, 1991) and Sambrook *et al., supra.*

Methods of transfection are known to the ordinarily skilled artisan, for example, $CaPO_4$ and electroporation. Depending on the host cell used, transformation is performed using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in Sambrook *et al.*, supra, or electroporation is generally used for prokaryotes or other cells that contain substantial cell-wall barriers. Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., *Gene*, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, *Virology* 52:456-457 (1978) can be employed. General aspects of mammalian cell host system transformations have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of Van Solingen *et al., J. Bact.*, 130:946 (1977) and Hsiao *et al., Proc. Natl. Acad. Sci. (USA)*, 76:3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, *e.g.*, polybrene, polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown *et al., Methods in Enzymology*, 185:527-537 (1990) and Mansour *et al., Nature*, 336:348-352 (1988).

Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote,

yeast. or higher eukaryote cells. Suitable prokaryotes include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriae such as *E. coli*. Various *E. coli* strains are publicly available, such as *E. coli* K12 strain MM294 (ATCC 31,446); *E. coli* X1776 (ATCC 31,537); *E. coli* strain W3110 (ATCC 27,325) and K5 772 (ATCC 53,635).

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for vertebrate *patched-2*-encoding vectors. *Saccharomyces cerevisiae* is a commonly used lower eukaryotic host microorganism.

Suitable host cells for the expression of vertebrate *patched-2* are derived from multicellular organisms. Examples of invertebrate cells include insect cells such as Drosophila S2 and Spodoptera Sf9, as well as plant cells. Examples of useful mammalian host cell lines include Chinese hamster ovary (CHO) and COS cells. More specific examples include monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham *et al., J. Gen Virol.*, 36:59 (1977)); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, *Proc. Natl. Acad. Sci. USA*, 77:4216 (1980)); mouse sertoli cells (TM4, Mather, *Biol. Reprod.* 23:243-251 (1980)); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); and mouse mammary tumor (MMT 060562, ATCC CCL51). The selection of the appropriate host cell is deemed to be within the skill in the art.

3. Selection and Use of a Replicable Vector

The nucleic acid (*e.g.*, cDNA or genomic DNA) encoding *patched-2* may be inserted into a replicable vector for cloning (amplification of the DNA) or for expression. Various vectors are publicly available. The vector may, for example, be in the form of a plasmid, cosmid, viral particle, or phage. The appropriate nucleic acid sequence may be inserted into the vector by a variety of procedures. In general, DNA is inserted into an appropriate restriction endonuclease site(s) using techniques known in the art. Vector components generally include, but are not limited to, one or more of a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Construction of suitable vectors containing one or more of these components employs standard ligation techniques, which are known to the skilled artisan.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. A preferred replicable expression vector is the plasmid is pRK5. *Holmes et al., Science*, 253:1278-1280 (1991).

Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, *e.g.*, ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, *e.g.*, the gene encoding D-alanine racemase for *Bacilli*.

An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the *patched-2* nucleic acid, such as DHFR or thymidine kinase.

An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described by Urlaub *et al., Proc. Natl. Acad. Sci. USA*, 77:4216 (1980). A suitable selection gene for use in yeast is the *trp*1 gene present in the yeast plasmid YRp7 [Stinchcomb *et al., Nature*, 282:39 (1979); Kingsman *et al., Gene*, 7:141 (1979); Tschemper *et al., Gene*, 10:157 (1980)]. The *trp*1 gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEP4-1 [Jones, *Genetics*, 85:12 (1977)].

Expression and cloning vectors usually contain a promoter operably linked to the *patched-2* nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a variety of potential host cells are well known. Promoters suitable for use with prokaryotic hosts include the β-lactamase and lactose promoter systems [Chang *et al., Nature*, 275:615 (1978); Goeddel *et al., Nature*, 281:544 (1979)], alkaline phosphatase, a tryptophan (trp) promoter system [Goeddel, *Nucleic Acids Res.*, 8:4057 (1980); EP 36,776], and hybrid promoters such as the tac promoter [deBoer *et al., Proc. Natl. Acad. Sci. USA*, 80:21-25 (1983)]. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding *patched-2*.

Examples of suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase [Hitzeman *et al., J. Biol. Chem.*, 255:2073 (1980)] or other glycolytic enzymes [Hess *et al., J. Adv. Enzyme Reg.*, 7:149 (1968); Holland, *Biochemistry*, 17:4900 (1978)], such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phospho-fructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in EP 73,657.

Patched-2 transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), from heterologous mammalian promoters, *e.g.*, the actin promoter or an immunoglobulin promoter, and from heat-shock promoters, provided such promoters are compatible with the host cell systems.

Inserting an enhancer sequence into the vector may increase transcription of a DNA encoding the *patched-2* by higher eukaryotes. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α-fetoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. The enhancer may be spliced into the vector at

a position 5' or 3' to the *patched-2* coding sequence, but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3', untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding *patched-2*.

Still other methods, vectors, and host cells suitable for adaptation to the synthesis of *patched-2* in recombinant vertebrate cell culture are described in Gething *et al.*, *Nature*, 293:620-625 (1981); Mantei *et al.*, *Nature*. 281:40-46 (1979); EP 117.060; and EP 117,058.

4. Detecting Gene Amplification/Expression

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA [Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 (1980)], dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of cells or tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal, and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a native sequence *patched-2* polypeptide or against a synthetic peptide based on the DNA sequences provided herein or against exogenous sequence to *patched-2* DNA and encoding a specific antibody epitope.

5. Purification of Polypeptide

Forms of *patched-2* may be recovered from host cell lysates. Since *patched-2* is membrane-bound, it can be released from the membrane using a suitable detergent solution (*e.g.* Triton-X 100) or by enzymatic cleavage. Cells employed in expression of *patched-2* can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

It may be desired to purify *patched-2* from recombinant cell proteins or polypeptides. The following procedures are exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex G-75; protein A Sepharose columns to remove contaminants such as IgG; and metal chelating columns to bind epitope-tagged forms of the *patched-2*. Various methods of protein purification may be employed and such methods are known in the art and described for example in Deutscher, Methods

in Enzymology, 182 (1990); Scopes, Protein Purification: Principles and Practice, Springer-Verlag, New York (1982). The purification step(s) selected will depend, for example, on the nature of the production process used and the particular *patched-2* produced.

E. Uses for *patched-2*

(1) Patched-2 is a specific receptor for desert hedgehog (Dhh)

The *hedgehog* (*Hh*) signaling pathway has been implicated in the formation of embryonic structures in mammals and invertebrates. The multi-pass transmembrane receptor *patched*, is a negative regulator of the *Hh* pathway, repressing the serpentine signaling molecule smoothened (*Smo*). Data have shown that loss of *Patched* leads to deregulation of the *Hh* pathway leading to formation of aberrant structures in the embryos and carcinoma in the adult.

Applicants' newly identified second human *patched* gene, termed *patched-2* (e.g., *Ptch-2*, SEQ ID NO:2), has a similar 12 transmembrane domain topology as does *patched*, and can bind to all the members of the *Hh* family and can complex with *Smo* (e.g., SEQ ID NO:17). However, the expression patterns of *patched-2* and *patched* do not overlap. *Patched-2* is expressed mainly in the developing spermatocytes, which are supported directly by the *Desert hedgehog* producing Sertoli cells, which suggests that *patched*-2 is a receptor for *Desert hedgehog*.

In the adult tubule, Sertoli cells, which are unusually large secretory cells, traverse the seminiferous tubule from the basal lamina to the luminal aspect, sending out cytoplasmic protrusions that engulf the germ cells. These contacts are particularly close during spermiogenesis, in which the haploid round spermatids undergo differentiation to produce the highly specialized, motile sperm. Tight junctions between adjacent Sertoli cells compartmentalize the tubule into a basal region, which contains mitotic spermatogonia and early spermtocytes, and an adluminal compartment, which contains meiotic spermatocytes and maturing spermatids. In fact, a Sertoli-derived cell line supports the meiotic progression of germ cells in culture, consistent with the view that factors derived from Sertoli cells contribute to germ cell maturation, Rassouizadegan, M., *et al.*. *Cell* 1993, 75: 997-1006. Loss of *Dhh* activity results in a recessive, sex-specific phentotype. Female mice homozygous for the mutation were fully viable and fertile, whereas male mice were viable but infertile. A gross examination indicated that, as early as 18.5 dpc, the testes of mutant males were noticeably smaller than those of heterozygous littermates. Bitgood *et al.*, *Curr. Biol.*, 1996 6(3): 298-304. Thus, Sertoli cells likely independently regulate mitotic and meiotic stages of germ cell development during postnatal development. Therefore, since *patched-2* appears to be the receptor for *Dhh* (SEQ ID NO:13), molecules which modulate the binding of *Dhh* (SEQ ID NO:13) to *patched*-2 would affect the activation of *Dhh* (SEQ ID NO:13) signaling, and thereby would have utility in the treatment of conditions which are modulated by *Dhh* (SEQ ID NO:13). (For example, testicular cancer). Alternatively, it is also provided that antagonists or agonists of *patched*-2 may be used for treating disorders or creating a desirable physiological condition effected by blocking *Dhh* signaling. (*E.g*, contraception, infertility treatment).

(2) General uses for patched-2

Nucleotide sequences (or their complement) encoding *patched-2* have various applications in the art of molecular biology, including uses as hybridization probes, in chromosome and gene mapping and in the generation of anti-sense RNA and DNA. *Patched-2* nucleic acid will also be useful for the preparation of

patched-2 polypeptides by the recombinant techniques described herein.

The full-length native sequence *patched-2* gene, or portions thereof. may be used as hybridization probes for a cDNA library to isolate the full-length gene or to isolate still other genes (for instance, those encoding naturally-occurring variants of *patched-2*) which have a desired sequence identity to the *patched-2* sequence disclosed in Fig. 1 (SEQ ID NO:1). Optionally, the length of the probes will be about 20 to about 50 bases. The hybridization probes may be derived from the nucleotide sequence of Fig. 1 (SEQ ID NO:1) or from genomic sequences including promoters. enhancer elements and introns of native sequence *patched-2*. By way of example, a screening method will comprise isolating the coding region of the *patched-2* gene using the known DNA sequence to synthesize a selected probe of about 40 bases. Hybridization probes may be labeled by a variety of labels, including radionucleotides such as ^{32}P or ^{35}S, or enzymatic labels such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems. Labeled probes having a sequence complementary to that of the *patched-2* gene of the present invention can be used to screen libraries of human cDNA, genomic DNA or mRNA to determine to which members of such libraries the probe hybridizes. Hybridization techniques are described in further detail in the Examples below.

The probes may also be employed in PCR techniques to generate a pool of sequences for identification of closely related *patched-2* sequences.

Nucleotide sequences encoding *patched-2* can also be used to construct hybridization probes for mapping the gene, which encodes *patched-2* and for the genetic analysis of individuals with genetic disorders. The nucleotide sequences provided herein may be mapped to a chromosome and specific regions of a chromosome using known techniques, such as *in situ* hybridization, linkage analysis against known chromosomal markers, and hybridization screening with libraries.

Patched-2 polypeptides can be used in assays to identify the other proteins or molecules involved in complexing with *patched-2* which ultimately results in the modulation of *hedgehog* signaling. Alternatively, these molecules can modulate the binding of *patched*-2 to *Dhh* (SEQ ID NO:13). By such methods, inhibitors of the binding interaction can be identified. Proteins involved in such binding interactions can also be used to screen for peptide or small molecule inhibitors or agonists of the binding interaction. Also, the substrate of *patched-2* can be used to isolate correlative complexing proteins. Screening assays can be designed to find lead compounds that mimic the biological activity of a native *patched-2* or to find those that act as a substrate for *patched-2*. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates. Such small molecule inhibitors could block the enzymatic action of *patched*-2, and thereby inhibit *hedgehog* signaling. Small molecules contemplated include synthetic organic or inorganic compounds. The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays and cell based assays, which are well characterized in the art.

Nucleic acids which encode *patched-2* or its modified forms can also be used to generate either transgenic animals or "knock out" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. A transgenic animal (*e.g.*, a mouse or rat) is an animal having cells that contain a transgene, was introduced into the animal or an ancestor of the animal at a prenatal, e.g., an embryonic stage. A transgene is a DNA sequence that is integrated into the genome of a cell from which a

transgenic animal develops. In one embodiment, cDNA encoding *patched-2* can be used to clone genomic DNA encoding *patched-2* in accordance with established techniques and the genomic sequences used to generate transgenic animals that contain cells which express DNA encoding *patched-2*. Methods for generating transgenic animals, particularly animals such as mice or rats, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009. Typically, particular cells would be targeted for *patched-2* transgene incorporation with tissue-specific enhancers. Transgenic animals that include a copy of a transgene encoding *patched-2* introduced into the germ line of the animal at an embryonic stage can be used to examine the effect of increased expression of DNA encoding *patched-2*. Such animals can be used as tester animals for reagents thought to confer protection from, for example, pathological conditions associated with its overexpression.

Non-human homologues of vertebrate patched-2 can be used to construct a *patched-2* "knock out" animal which has a defective or altered gene encoding *patched-2* as a result of homologous recombination between the endogenous gene encoding *patched-2* and altered genomic DNA encoding *patched-2* introduced into an embryonic cell of the animal. For example, cDNA encoding *patched-2* can be used to clone genomic DNA encoding *patched-2* in accordance with established techniques. A portion of the genomic DNA encoding *patched-2* can be deleted or replaced with another gene, such as a gene encoding a selectable marker that can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector [see e.g., Thomas and Capecchi, Cell, 51:503 (1987) for a description of homologous recombination vectors]. The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected [see e.g., Li *et al.*, *Cell*, 69:915 (1992)]. The selected cells are then injected into a blastocyst of an animal (e.g., a mouse or rat) to form aggregation chimeras [see e.g., Bradley, in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E. J. Robertson, ed. (IRL, Oxford, 1987), pp. 113-152]. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term to create a "knock out" animal. Progeny harboring the homologously recombined DNA in their germ cells can be identified by standard techniques and used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knockout animals can be characterized for instance, for their ability to defend against certain pathological conditions and for their development of pathological conditions due to absence of the *patched-2* polypeptide.

Suppression or inhibition (antagonism) of *Dhh* signaling is also an objective of therapeutic strategies. Since *patched*-2 can combine with all members of the hedgehog family (*i.e.*, *Shh, Dhh, Ihh*), antagonist molecules which prevent the binding of hedgehog molecules to *Ptch-2* (SEQ ID NO:2) have therapeutic utility. For example, *SHh* signaling is known to be activated in Basal Cell Carcinoma: *Dhh* (SEQ ID NO:13) is known to be involved in the regulation of spermatogenesis. Inhibitor or antagonist of *Hh* signaling would be effective therapeutics in the treatment of Basal Cell Carcinoma or male contraception, respectively.

The stimulation of *Dhh* signaling (agonism) is also an objective of therapeutic strategies. Since *Ptch*-2 (SEQ ID NO:2) also binds to the other members of the *Hh* family, *Ihh* and *Shh*, activating *Dhh* signaling would be useful in disease states or disorders characterized by inactive or insufficient *Hh* signaling.

For example, degenerative disorders of the nervous system, *e.g.*, Parkinson's disease, memory deficits, Alzheimer's disease, Lou Gehrig's disease, Huntington's disease, schizophrenia, stroke and drug addiction. Additionally, *patched-2* agonists could be used to treat gut diseases, bone diseases, skin diseases, diseases of the testis (including infertility), ulcers, lung diseases, diseases of the pancreas, diabetes, osteoporosis.

F. Anti-*patched-2* Antibodies

The present invention further provides anti- vertebrate *patched-2* antibodies. Exemplary antibodies include polyclonal, monoclonal, humanized, bispecific, and heteroconjugate antibodies.

1. Polyclonal Antibodies

The anti-*patched-2* antibodies may comprise polyclonal antibodies. Methods of preparing polyclonal antibodies are known to the skilled artisan. Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. The immunizing agent may include the *patched-2* polypeptide or a fusion protein thereof. It may be useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include but are not limited to keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. Examples of adjuvants that may be employed include Freund's complete adjuvant and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate). The immunization protocol may be selected by one skilled in the art without undue experimentation.

2. Monoclonal Antibodies

The anti-*patched-2* antibodies may, alternatively, be monoclonal antibodies. Monoclonal antibodies may be prepared using hybridoma methods, such as those described by Kohler and Milstein, Nature, 256:495 (1975). In a hybridoma method, a mouse, hamster, or other appropriate host animal, is typically immunized with an immunizing agent to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized *in vitro*.

The immunizing agent will typically include the *patched-2* polypeptide or a fusion protein thereof. Generally, either peripheral blood lymphocytes ("PBLs") are used if cells of human origin are desired, or spleen cells or lymph node cells are used if non-human mammalian sources are desired. The lymphocytes are then fused with an immortalized cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell [Goding, Monoclonal Antibodies: Principles and Practice, Academic Press, (1986) pp. 59-103]. Immortalized cell lines are usually transformed mammalian cells, particularly myeloma cells of rodent, bovine and human origin. Usually, rat or mouse myeloma cell lines are employed. The hybridoma cells may be cultured in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused immortalized cells. For example, if the parental cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine ("HAT medium"), which substances prevent the growth of HGPRT-deficient cells.

Preferred immortalized cell lines are those that fuse efficiently, support stable high level expression

of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. More preferred immortalized cell lines are murine myeloma lines, which can be obtained, for instance, from the Salk Institute Cell Distribution Center, San Diego, California and the American Type Culture Collection, Rockville, Maryland. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies [Kozbor, J. Immunol., 133:3001 (1984); Brodeur et al., Monoclonal Antibody Production Techniques and Applications, Marcel Dekker, Inc., New York, (1987) pp. 51-63].

The culture medium in which the hybridoma cells are cultured can then be assayed for the presence of monoclonal antibodies directed against *patched-2*. Preferably, the binding specificity of monoclonal antibodies produced by the hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA). Such techniques and assays are known in the art. The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson and Pollard, Anal. Biochem., 107:220 (1980).

After the desired hybridoma cells are identified, the clones may be subcloned by limiting dilution procedures and grown by standard methods [Goding, supra]. Suitable culture media for this purpose include, for example, Dulbecco's Modified Eagle's Medium and RPMI-1640 medium. Alternatively, the hybridoma cells may be grown *in vivo* as ascites in a mammal.

The monoclonal antibodies secreted by the subclones may be isolated or purified from the culture medium or ascites fluid by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

The monoclonal antibodies may also be made by recombinant DNA methods, such as those described in U.S. Patent No. 4,816,567. DNA encoding the monoclonal antibodies of the invention can be readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells of the invention serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. The DNA also may be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences [U.S. Patent No. 4,816,567; Morrison *et al., supra*] or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. Such a non-immunoglobulin polypeptide can be substituted for the constant domains of an antibody of the invention, or can be substituted for the variable domains of one antigen-combining site of an antibody of the invention to create a chimeric bivalent antibody.

The antibodies may be monovalent antibodies. Methods for preparing monovalent antibodies are well known in the art. For example, one method involves recombinant expression of immunoglobulin light chain and modified heavy chain. The heavy chain is truncated generally at any point in the Fc region so as to prevent heavy chain crosslinking. Alternatively, the relevant cysteine residues are substituted with another

amino acid residue or are deleted so as to prevent crosslinking.

In vitro methods are also suitable for preparing monovalent antibodies. Digestion of antibodies to produce fragments thereof, particularly, Fab fragments, can be accomplished using routine techniques known in the art.

3. Humanized Antibodies

The anti-*patched-2* antibodies of the invention may further comprise humanized antibodies or human antibodies. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', $F(ab')_2$ or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies may also comprise residues that are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin [Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-329 (1988); and Presta, Curr. Op. Struct. Biol., 2:593-596 (1992)].

Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source that is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers [Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-327 (1988); Verhoeyen et al., Science, 239:1534-1536 (1988)], by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Patent No. 4,816,567), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

Human antibodies can also be produced using various techniques known in the art, including phage display libraries [Hoogenboom and Winter, J. Mol. Biol., 227:381 (1991); Marks et al., J. Mol. Biol., 222:581 (1991)]. The techniques of Cole *et al.* and Boerner *et al.* are also available for the preparation of human monoclonal antibodies (Cole *et al.*, Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985) and Boerner et al., J. Immunol., 147(1):86-95 (1991)].

4. Bispecific Antibodies

Bispecific antibodies are monoclonal, preferably human or humanized, antibodies that have binding specificities for at least two different antigen s. In the present case, one of the binding specificities is for the vertebrate *patched-2*, the other one is for any other antigen, and preferably for a cell-surface protein or receptor or receptor subunit.

Methods for making bispecific antibodies are known in the art. Traditionally, the recombinant production of bispecific antibodies is based on the co-expression of two immunoglobulin heavy-chain/light-chain pairs, where the two heavy chains have different specificities [Milstein and Cuello, Nature, 305:537-539 (1983)]. Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of ten different antibody molecules, of which only one has the correct bispecific structure. The purification of the correct molecule is usually accomplished by affinity chromatography steps. Similar procedures are disclosed in WO 93/08829, published 13 May 1993, and in Traunecker *et al.*, EMBO J., 10:3655-3659 (1991).

Antibody variable domains with the desired binding specificities (antibody-antigen combining sites) can be fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy-chain constant domain, comprising at least part of the hinge, CH2, and CH3 regions. It is preferred to have the first heavy-chain constant region (CH1) containing the site necessary for light-chain binding present in at least one of the fusions. DNAs encoding the immunoglobulin heavy-chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host organism. For further details of generating bispecific antibodies see, for example, Suresh et al., Methods in Enzymology, 121:210 (1986).

5. Heteroconjugate Antibodies

Heteroconjugate antibodies are also within the scope of the present invention. Heteroconjugate antibodies are composed of two covalently joined antibodies. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells [U.S. Patent No. 4,676,980], and for treatment of HIV infection [WO 91/00360; WO 92/200373; EP 03089]. It is contemplated that the antibodies may be prepared *in vitro* using known methods in synthetic protein chemistry, including those involving crosslinking agents. For example, immunotoxins may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents for this purpose include iminothiolate and methyl-4-mercaptobutyrimidate and those disclosed, for example, in U.S. Patent No. 4,676,980.

G. Uses for anti-*patched-2* Antibodies

The anti-*patched-2* antibodies of the invention have various utilities. For example, anti-*patched-2* antibodies may be used in diagnostic assays for *patched-2*, *e.g.*, detecting its expression in specific cells, tissues, or serum. Various diagnostic assay techniques known in the art may be used, such as competitive binding assays, direct or indirect sandwich assays and immunoprecipitation assays conducted in either heterogeneous or homogeneous phases [Zola, Monoclonal Antibodies: A Manual of Techniques, CRC Press, Inc. (1987) pp. 147-158]. The antibodies used in the diagnostic assays can be labeled with a detectable moiety. The detectable moiety should be capable of producing, either directly or indirectly, a detectable signal. For example, the detectable moiety may be a radioisotope, such as ^{3}H, ^{14}C, ^{32}P, ^{35}S, or ^{125}I, a

fluorescent or chemiluminescent compound, such as fluorescein isothiocyanate, rhodamine, or luciferin, or an enzyme, such as alkaline phosphatase, beta-galactosidase or horseradish peroxidase. Any method known in the art for conjugating the antibody to the detectable moiety may be employed, including those methods described by Hunter et al., Nature, 144: 945 (1962); David *et al.*, Biochemistry, 13:1014 (1974); Pain *et al.*, J. Immunol. Meth., 40:219 (1981); and Nygren, J. Histochem. and Cytochem., 30:407 (1982).

Anti-*patched-2* antibodies also are useful for the affinity purification of *patched-2* from recombinant cell culture or natural sources. In this process, the antibodies against *patched-2* are immobilized on a suitable support, such a Sephadex resin or filter paper, using methods well known in the art. The immobilized antibody then is contacted with a sample containing the *patched-2* to be purified, and thereafter the support is washed with a suitable solvent that will remove substantially all the material in the sample except the *patched-2*, which is bound to the immobilized antibody. Finally, the support is washed with another suitable solvent that will release the *patched-2* from the antibody.

Basal cell carcinoma (BCC) is the most common human cancer. The *Hh* signaling pathway was found to be activated in all BCCs. Loss of *patched* function is thought to lead to unregulated *Smo* activity and is responsible for about half of all BCCs. *Patched* being a target of the Hh pathway itself, increases in *patched* mRNA levels have been detected in BCC [Gailani, *et al.*, *Nature Genet.* 14: 78-81 (1996)] as well as in animal models of BCC. *Oro et al.*, *Science* 276: 817-821 (1997); Xie *et al.*, *Nature* 391: 90-92 (1998). Abnormal activation of *Sh* signaling, such as that which occurs in BCC, was examined to confirm whether *patched-2* expression was increased. As shown in Fig. 9, an *in situ* hybridization for *Ptch* (SEQ ID NO:4) and *Ptch*-2 (SEQ ID NO:2) in *Smo*-M2 (SEQ ID NO:16) transgenic mice (Xie *et al.*, *supra*), while lower than *Ptch*, was still high in tumor cells. This suggests that therapeutic antibodies directed toward *Ptch*-2 (SEQ ID NO:2) may be useful for the treatment of BCC.

Anti-*patched*-2 antibodies also have utilities similar to those articulated for under the previous section "E. Uses of Patched-2". Depending on whether anti-*patched*-2 antibodies will bind *patched*-2 receptors so as to either inhibit *Hh* signaling (antagonist) or inhibit *patched*-2 complexing with *Smo* (SEQ ID NO:17) and thereby remove the normal inhibitory effect of *Smo* (SEQ ID NO:17) on *Hh* signaling (agonist) the antibody will have utilities corresponding to those articulated previously for *patched*-2.

H. *Patched-2* Antagonists

Several approaches may be suitably employed to create the *patched-2* antagonist and agonist compounds of the present invention. Any approach where the antagonist molecule can be targeted to the interior of the cell, which interferes or prevents wild type *patched-2* from normal operation is suitable. For example, competitive inhibitors, including mutant *patched-2* receptors which prevent wild type *patched-2* from properly binding with other proteins necessary for *Dhh* and *Hh* signaling. Additional properties of such antagonist or agonist molecules are readily determinable by one of ordinary skill, such as size, charge and hydrophobicity suitable for transmembrane transport.

Where mimics or other mammalian homologues of *patched-2* are to be identified or evaluated, the cells are exposed to the test compound and compared to positive controls which are exposed only to human *patched-2*, and to negative controls which were not exposed to either the compound or the natural ligand. Where antagonists or agonists of *patched-2* signal modulation are to be identified or evaluated, the cells are

exposed to the compound of the invention in the presence of the natural ligand and compared to controls which are not exposed to the test compound.

Detection assays may by employed as a primary screen to evaluate the *Hh* signaling inhibition/enhancing activity of the antagonist/agonist compounds of the invention. The assays may also be used to assess the relative potency of a compound by testing a range of concentrations, in a range from 100 mM to 1 pM, for example, and computing the concentration at which the amount of phosphorylation or signal transduction is reduced or increased by 50% (IC_{50}) compared to controls.

Assays can be performed to identify compounds that affect *Hh* signaling of *patched-2* substrates. Specifically, assays can be performed to identify compounds that increase the phosphorylation activity of *patched-2* or assays can be performed to identify compounds that decrease the *Hh* signaling of *patched-2* substrates. These assays can be performed either on whole cells themselves or on cell extracts. The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays, cell based assays, etc. Such assay formats are well known in the art.

The screening assays of the present invention are amenable to high-throughput screening of chemical libraries, and are particularly suitable for identifying small molecule drug candidates.

(1) Antagonist and agonist molecules

To screen for antagonists and/or agonists of *patched-2* signaling, the assay mixture is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, *patched-2* induces hedgehog signaling with a reference activity. The mixture components can be added in any order that provides for the requisite hedgehog activity. Incubation may be performed at any temperature that facilitates optimal binding, typically between about 4° and 40°C, more commonly between about 15° and 40°C. Incubation periods are likewise selected for optimal binding but also minimized to facilitate rapid, high-throughput screening, and are typically between about 0.1 and 10 hours, preferably less than 5 hours, more preferably less than 2 hours. After incubation, the effect of the candidate pharmacological agent on the *patched-2* signaling is determined in any convenient way. For cell-free binding-type assays, a separation step is often used to separate bound and unbound components. Separation may, for example, be effected by precipitation (*e.g.* TCA precipitation, immunoprecipitation, etc.), immobilization (e.g. on a solid substrate), followed by washing. The bound protein is conveniently detected by taking advantage of a detectable label attached to it, *e.g.* by measuring radioactive emission, optical or electron density, or by indirect detection using, *e.g.* antibody conjugates.

For example, a method of screening for suitable *patched-2* antagonists and/or agonists could involve the application of *Dhh* and other hedgehog ligands. Such a screening assay could compare *in situ* hybridization in the presence and absence of the candidate antagonist and/or agonist in a *patched-2* expressing tissue as well as confirmation or absence of *patched-2* modulated cellular development. Typically these methods involve exposing an immobilized *patched-2* to a molecule suspected of binding thereto and determining the level of ligand binding downstream activation of reporter constructs and/or evaluating whether or not the molecule activates (or blocks activation of) *patched-2*. In order to identify such *patched-2* binding ligands, *patched-2* can be expressed on the surface of a cell and used to screen

libraries of synthetic candidate compounds or naturally-occurring compounds (e.g., from endogenous sources such as serum or cells).

Suitable molecules that affect the protein-protein interaction of *patched-2* and its binding proteins include fragments of the latter or small molecules, *e.g.*, peptidomimetics, which will inhibit ligand-receptor interaction. Such small molecules, which are usually less than 10 K molecular weight, are preferable as therapeutics since they are more likely to be permeable to cells, are less susceptible to degradation by various cellular mechanisms, and are not as apt to elicit an immune response as proteins. Small molecules include but are not limited to synthetic organic or inorganic compounds. Many pharmaceutical companies have extensive libraries of such molecules, which can be conveniently screened by using the assays of the present invention. Non-limiting examples include proteins, peptides, glycoproteins, glycopeptides, glycolipids, polysaccharides, oligosacchardies, nucleic acids, bioorganic molecules, peptidomimetics, pharmacological agents and their metabolites, transcriptional and translation control sequences, and the like.

A preferred technique for identifying molecules which bind to *patched-2* utilizes a chimeric substrate (*e.g.*, epitope-tagged *patched-2* or *patched-2* immunoadhesin) attached to a solid phase, such as the well of an assay plate. The binding of the candidate molecules, which are optionally labeled (e.g., radiolabeled), to the immobilized receptor can be measured. Alternatively, competition for various *Hh* pathways, especially *Dhh* (SEQ ID NO:13) can be measured. In screening for antagonists and/or agonists, *patched-2* can be exposed to a *patched-2* substrate followed by the putative antagonist and/or agonist, or the *patched-2* binding protein and antagonist and/or agonist can be added simultaneously, and the ability of the antagonist and/or agonist to block *patched-2* activation can be evaluated.

(2) Detection assays

The *patched-2* polypeptides are useful in assays for identifying lead compounds for therapeutically active agents that modulate *patched-2* receptor/ligand hedgehog signaling. Specifically, lead compounds that either prevent the formation of *patched-2* signaling complexes or prevent or attenuate *patched-2* modulated hedgehog signaling (*e.g*, binding to *patched*-2) can be conveniently identified.

Various procedures known in the art may be used for identifying, evaluating or assaying the inhibition of activity of the *patched-2* proteins of the invention. As *patched-2* is believed to be a receptor for *Dhh* (SEQ ID NO:13), but also binds *Shh* (SEQ ID NO:14) and *Ihh* (SEQ ID NO:29), techniques known for use with identifying ligand/receptor modulators may also be employed with the present invention. In general, such assays involve exposing target cells in culture to the compounds and a) biochemically analyzing cell lysates to assess the level and/or identity of binding; or (b) scoring phenotypic or functional changes in treated cells as compared to control cells that were not exposed to the test substance. Such screening assays are described in U.S.P. 5,602171, U.S.P. 5,710,173, WO 96/35124 and WO 96/40276.

(a) Biochemical detection techniques

Biochemical analysis can be evaluated by a variety of techniques. One typical assay mixture which can be used with the present invention contains *patched-2* and a ligand protein with which *patched-2* is normally associated (*e.g.*, *Dhh* (SEQ ID NO:13)) usually in an isolated, partially pure or pure form. One or both of these components may be *patched-2* to another peptide or polypeptide, which may, for example, provide or enhance protein-protein binding, improve stability under assay conditions, *etc.*

In addition, one of the components usually comprises or is coupled to a detectable label. The label may provide for direct detection by measuring radioactivity, luminescence, optical or electron density, etc., or indirect detection such as an epitope tag, an enzyme, etc. The assay mixture can additionally comprise a candidate pharmacological agent, and optionally a variety of other components, such as salts, buffers, carrier proteins, e.g. albumin, detergents, protease inhibitors, nuclease inhibitors, antimicrobial agents, etc., which facilitate binding, increase stability, reduce non-specific or background interactions, or otherwise improve the efficiency or sensitivity of the assay.

The following detection methods may also be used in a cell-free system wherein cell lysate containing the signal transducing substrate molecule and *patched-2* is mixed with a compound of the invention. To assess the activity of the compound, the reaction mixture may be analyzed by the SDS-PAGE technique or it may be added to substrate-specific anchoring antibody bound to a solid support, and a detection procedure as described above is performed on the separated or captured substrate to assess the presence or absence of a *patched*-2 binding ligand. The results are compared to those obtained with reaction mixtures to which the compound is not added. The cell-free system does not require the natural ligand or knowledge of its identity. For example, Posner *et al.* (U.S.P. 5,155,031 describes the use of insulin receptor as a substrate and rat adipocytes as target cells to demonstrate the ability of pervanadate to inhibit PTP activity. Another example, Burke *et al., Biochem. Biophys. Res. Comm.* 204: 129-134 (1994) describes the use of autophosphorylated insulin receptor and recombinant PTP1B in assessing the inhibitory activity of a phosphotyrosyl mimetic.

(i) Whole cell detection

A common technique involves incubating cells with *patched-2* and radiolabeled ligand, lysing the cells, separating cellular protein components of the lysate using an SDS-polyacrylamide gel (SDS-PAGE) technique, in either one or two dimensions, and detecting the presence of labeled proteins by exposing X-ray film. Detection can also be effected without using radioactive labeling. In such a technique, the protein components (e.g., separated by SDS-PAGE) are transferred to a nitrocellulose membrane where the presence of patched-ligand complexes is detected using an anti-ligand antibody.

Alternatively, the anti-*patched*-2 ligand antibody can be conjugated with an enzyme, such as horseradish peroxidase, and detected by subsequent addition of a colorimetric substrate for the enzyme. A further alternative involves detecting the anti-*patched*-2 ligand by reacting with a second antibody that recognizes anti-*patched*-2 ligand, this second antibody being labeled with either a radioactive moiety or an enzyme as previously described. Examples of these and similar techniques are described in Hansen *et al., Electrophoresis* 14: 112-126 (1993); Campbell *et al., J. Biol. Chem.* 268: 7427-7434 (1993); Donato *et al., Cell Growth Diff.* 3: 258-268 (1992); Katagiri *et al., J. Immunol.* 150: 585-593 (1993). Additionally, the anti-*patched*-2 ligand can be detected by labeling it with a radioactive substance, followed by scanning the labeled nitrocellulose to detect radioactivity or exposure of X-ray film.

Further detection methods may be developed which are preferred to those described above. Especially for use in connection with high-throughput screening, it is expected that such methods would

exhibit good sensitivity and specificity, extended linear range, low background signal, minimal fluctuation, compatibility with other reagents, and compatibility with automated handling systems.

The *in vivo* efficacy of the treatment of the present invention can be studied against chemically induced tumors in various rodent models. Tumor cell lines propagated in *in vitro* cell cultures can be introduced in experimental rodents, e.g. mice by injection, for example by the subcutaneous route. Techniques for chemical inducement of tumors in experimental animals are well known in the art.

(ii) Kinase assays

Because *patched*-2 is a negative regulator of *Hh* signaling, which when activated by *Hh* releases the normal inhibitory effect on *Smo*, the inhibition of *patched*-2 binding to *Smo* can be measured by activation of various kinase substrate associated with *Hh* signaling. When the screening methods of the present invention for *patched*-2 antagonists/agonists are carried out as an *ex vivo* assay, the target kinase (*e.g. fused*) can be a substantially purified polypeptide. The kinase substrate (*e.g.*, MBP, *Gli*) is a substantially purified substrate, which in the assay is phosphorylated in a reaction with a substantially purified phosphate source that is catalyzed by the kinase. The extent of phosphorylation is determined by measuring the amount of substrate phosphorylated in the reaction. A variety of possible substrates may be used, including the kinase itself in which instance the phosphorylation reaction measured in the assay is autophosphorylation. Exogenous substrates may also be used, including standard protein substrates such as myelin basic protein (MBP); yeast protein substrates; synthetic peptide substrates, and polymer substrates. Of these, MBP and other standard protein substrates may be regarded as preferred. Other substrates may be identified, however, which are superior by way of affinity for the kinase, minimal perturbation of reaction kinetics, possession of single or homogenous reaction sites, ease of handling and post-reaction recover, potential for strong signal generation, and resistance or inertness to test compounds.

Measurement of the amount of substrate phosphorylated in the *ex vivo* assay of the invention may be carried out by means of immunoassay, radioassay or other well-known methods. In an immunoassay measurement, an antibody (such as a goat or mouse anti-phosphoserine/threonine antibody) may be used which is specific for phosphorylated moieties formed during the reaction. Using well-known ELISA techniques, the phosphoserine/threonine antibody complex would itself be detected by a further antibody linked to a label capable of developing a measurable signal (as for example a fluorescent or radioactive label). Additionally, ELISA-type assays in microtitre plates may be used to test purified substrates. Peraldi *et al., J. Biochem.* 285: 71-78 (1992); Schraag *et al., Anal. Biochem.* 211: 233-239 (1993); Cleavland, *Anal. Biochem.* 190: 249-253 (1990); Farley, *Anal. Biochem.* 203: 151-157 (1992) and Lozaro, *Anal. Biochem.* 192: 257-261 (1991).

For example, detection schemes can measure substrate depletion during the kinase reaction. Initially, the phosphate source may be radiolabeled with an isotope such as ^{32}P or ^{33}P, and the amount of substrate phosphorylation may be measured by determining the amount of radiolabel incorporated into the substrate during the reaction. Detection may be accomplished by: (a) commercially available scintillant-containing plates and beads using a beta-counter, after adsorption to a filter or a microtitre well surface, or (b) photometric means after binding to a scintillation proximity assay bead or scintillant plate. Weemink and

Kijken. *J. Biochem. Biophs. Methods* 31: 49, 1996; Braunwalder *et al., Anal. Biochem.* 234: 23 (1996); Kentrup *et al., J. Biol. Chem.* 271: 3488 (1996) and Rusken *et al., Meth. Enzymol.* 200: 98 (1991).

Preferably, the substrate is attached to a solid support surface by means of non-specific or, preferably, specific binding. Such attachment permits separation of the phosphorylated substrate from unincorporated, labeled phosphate source (such as adenosine triphosphate prior to signal detection. In one embodiment, the substrate may be physically immobilized prior to reaction, as through the use of Nunc™ high protein binding plate (Hanke *et al., J. Biol. Chem.* 271: 695 (1996)) or Wallac ScintiStrip™ plates (Braunwalder *et al., Anal. Biochem.* 234: 23 (1996). Substrate may also be immobilized after reaction by capture on, for example, P81 phophocellulose (for basic peptides), PEI/acidic molybdate resin or DEAE, or TCA precipitation onto Whatman™ 3MM paper, Tiganis *et al., Arch. Biochem. Biophys.* 325: 289 (1996); Morawetz *et al., Mol. Gen. Genet.* 250; 17 (1996); Budde *et al. Int J. Pharmacognosy* 33: 27 (1995) and Casnellie, *Meth. Enz.* 200: 115 (1991). Yet another possibility is the attachment of the substrate to the support surface, as by conjugation with binding partners such as glutathione and streptavidin (in the case of GST and biotin), respectively) which have been attached to the support, or via antibodies specific for the tags which are likewise attached to the support.

Further detection methods may be developed which are preferred to those described above. Especially for use in connection with high-throughput screening, it is expected that such methods would exhibit good sensitivity and specificity, extended linear range, low background signal, minimal fluctuation, compatibility with other reagents, and compatibility with automated handling systems.

The *in vivo* efficacy of the treatment of the present invention can be studied against chemically induced tumors in various rodent models. Tumor cell lines propagated in *in vitro* cell cultures can be introduced in experimental rodents, e.g. mice by injection, for example by the subcutaneous route. Techniques for chemical inducement of tumors in experimental animals are well known in the art.

(b) Biological detection techniques:

The ability of the antagonist/agonist compounds of the invention to modulate the activity of *patched-2*, which itself modulates *hedgehog* signaling, may also be measured by scoring for morphological or functional changes associated with ligand binding. Any qualitative or quantitative technique known in the art may be applied for observing and measuring cellular processes which comes under the control of *patched*-2. The activity of the compounds of the invention can also be assessed in animals using experimental models of disorders caused by or related to dysfunctional *hedgehog* signaling. For example, ineffective *Dhh hedgehog* signaling in mice leads to viable but sterile mice. Additionally, proper *Shh* signaling is critical to murine embryonic development at the notochord and floor plate, neural tube, distal limb structures, spinal column and ribs. Improper *Shh* signaling, is also correlative with cyclopia. Any of these phenotypic properties could be evaluated and quantified in a screening assay for *patched-2* antagonists and/or agonist. Disease states associated with overexpression of hedgehog is associated with basal cell carcinoma while inactive *Shh* signaling leads to improper neural development.

The data obtained from these cell culture assays and animal studies can be used in formulating a range of dosages for use in humans. The dosage of the compounds of the invention should lie within a range

of circulating concentrations with little or no toxicity. The dosage may vary within this range depending on the dosage form employed and the route of administration.

(2) *Antisense oligonucleotides*

Another preferred class of antagonists involves the use of gene therapy techniques. include the administration of antisense oligonucleotides. Applicable gene therapy techniques include single or multiple administrations of therapeutically effective DNA or mRNA. Antisense RNAs and DNAs can be used as therapeutic agents for blocking the expression of certain genes *in vivo*. Reference short antisense oligonucleotides can be imported into cells where they act as inhibitors, despite their low intracellular concentrations caused by restricted uptake by the cell membrane, Zamecnik *et al.*, *Proc. Natl. Acad. Sci. USA* 83: 4143-4146 (1986). The anti-sense oligonucleotides can be modified to enhance their uptake, e.g., by substituting their negatively charged phophodiester groups by uncharged groups.

There are a variety of techniques known for introducing nucleic acids into viable cells. The techniques vary depending upon whether the nucleic acid is transferred into cultured cells *in vitro*, *ex vivo*, or *in vivo* in the cells of the intended host. Techniques suitable for the transfer of nucleic acid into mammalian cells *in vitro* include the use of liposomes, electroporation, microinjection, cell fusion, DEAE-dextran, the calcium phosphate precipitation method, etc. The currently preferred *in vivo* gene transfer techniques include transfection with viral (typically retroviral) vectors and viral coat protein-liposome mediated transfection, Dzau *et al.*, *Trends Biotech.* 11: 205-210 (1993). In some situations it is desirable to provide the nucleic acid source with an agent that targets the cells, such as an antibody specific for a cell surface membrane protein associated with endocytosis may be used for targeting and/or to facilitate uptake, *e.g.*, capsid proteins or fragments thereof tropic for a particular cell type, antibodies for proteins which undergo internalization in cycling, and proteins that target intracellular localization and enhance intracellular half-life. The technique of receptor-mediated endocytosis is described, for example, by Wu *et al.*, *J. Biol. Chem.* 262: 4429-4432 (1987); Wagner *et al.*, *Proc. Natl. Acad. Sci. USA* 87: 3410-3414 (1990). For a review of known gene targeting and gene therapy protocols, see Anderson *et al.*, *Science* 256: 808-813 (1992).

In one embodiment of the invention, *patched-2* expression may be reduced by providing *patched-2*-expressing cells with an amount of *patched-2* antisense RNA or DNA effective to reduce expression of the *patched-2* protein.

I. Diagnostic Uses

Another use of the compounds of the invention (*e.g.*, *patched-2*, *patched-2* and anti-*patched-2* antibodies) described herein is to help diagnose whether a disorder is driven, to some extent, by *patched-2* or hedgehog signaling. For example, basal cell carcinoma cells are associated with active *hedgehog* signaling, spermatocyte formation is associated with *Dhh* signaling, and defective *patched* and *patched-2* suppression may be associated with testicular carcinomas.

A diagnostic assay to determine whether a particular disorder is driven by *patched-2* modulated *hedgehog* signaling, can be carried out using the following steps: (1) culturing test cells or tissues; (2) administering a compound which can prevent *patched-2* binding with *Smo* (SEQ ID NO:17), thereby activating the *Hh* signaling pathway; and (3) measuring the amount of *Hh* signaling. The steps can be carried

out using standard techniques in light of the present disclosure. For example. standard techniques can be used to isolate cells or tissues and culturing or *in vivo*.

Compounds of varying degree of selectivity are useful for diagnosing the role of *patched-2*. For example. compounds which inhibit *patched*-2 in addition to another form of kinase can be used as an initial test compound to determine if one of several signaling ligands drive the disorder. The selective compounds can then be used to further eliminate the possible role of the other ligands in driving the disorder. Test compounds should be more potent in inhibiting ligand-*patched*-2 binding activity than in exerting a cytotoxic effect (*e.g.*, an IC_{50}/LD_{50} of greater than one). The IC_{50} and LD_{50} can be measured by standard techniques, such as an MTT assay. or by measuring the amount of LDH released. The degree of IC_{50}/LD_{50} of a compound should be taken into account in evaluating the diagnostic assay. For example, the larger the IC_{50}/LD_{50} ratio the more relative the information. Appropriate controls take into account the possible cytotoxic effect of a compound of a compound, such as treating cells not associated with a cell proliferative disorder (*e.g.*, control cells) with a test compound. can also be used as part of the diagnostic assay. The diagnostic methods of the invention involve the screening for agents that modulate the effects of *patched*-2 upon *hedgehog* signaling. Exemplary detection techniques include radioactive labeling and immunoprecipitating (U.S.P. 5,385,915).

J. <u>Pharmaceutical Compositions and Dosages</u>

Therapeutic formulations of the compositions of the invention are prepared for storage as lyophilized formulations or aqueous solutions by mixing the *patched*-2 molecule, agonist and/or antagonist having the desired degree of purity with optional "pharmaceutically-acceptable" or "physiologically-acceptable" carriers, excipients or stabilizers typically employed in the art (all of which are termed "excipients"). For example. buffering agents, stabilizing agents, preservatives. isotonifiers. non-ionic detergents. antioxidants and other miscellaneous additives. (See Remington's Pharmaceutical Sciences, 16th Ed. . A. Osol. Ed. (1980)). Such additives must be nontoxic to the recipients at the dosages and concentrations employed.

Buffering agents help to maintain the pH in the range which approximates physiological conditions. They are preferably present at concentration ranging from about 2mM to about 50 mM. Suitable buffering agents for use with the present invention include both organic and inorganic acids and salts thereof such as citrate buffers (e.g., monosodium citrate-disodium citrate mixture. citric acid-trisodium citrate mixture, citric acid-monosodium citrate mixture, etc.), succinate buffers (e.g., succinic acid-monosodium succinate mixture, succinic acid-sodium hydroxide mixture, succinic acid-disodium succinate mixture, etc.), tartrate buffers (e.g., tartaric acid-sodium tartrate mixture, tartaric acid-potassium tartrate mixture, tartaric acid-sodium hydroxide mixture, etc.), fumarate buffers (e.g., fumaric acid-monosodium fumarate mixture, etc.). fumarate buffers (e.g., fumaric acid-monosodium fumarate mixture, fumaric acid-disodium fumarate mixture, monosodium fumarate-disodium fumarate mixture, etc.), gluconate buffers (e.g., gluconic acid-sodium glyconate mixture, gluconic acid-sodium hydroxide mixture, gluconic acid-potassium glyconate mixture, etc.), oxalate buffer (e.g., oxalic acid-sodium oxalate mixture. oxalic acid-sodium hydroxide mixture, oxalic acid-potassium oxalate mixture. etc.), lactate buffers (e.g., lactic acid-sodium lactate mixture. lactic acid-

sodium hydroxide mixture. lactic acid-potassium lactate mixture. etc.) and acetate buffers (e.g., acetic acid-sodium acetate mixture. acetic acid-sodium hydroxide mixture. etc.). Additionally, there may be mentioned phosphate buffers. histidine buffers and trimethylamine salts such as Tris.

Preservatives are added to retard microbial growth. and are added in amounts ranging from 0.2% - 1% (w/v). Suitable preservatives for use with the present invention include phenol. benzyl alcohol, *meta*-cresol. methyl paraben. propyl paraben, octadecyldimethylbenzyl ammonium chloride. benzalconium halides (e.g., chloride. bromide. iodide), hexamethonium chloride. alkyl parabens such as methyl or propyl paraben, catechol. resorcinol. cyclohexanol, and 3-pentanol.

Isotonifiers sometimes known as "stabilizers" are present to ensure isotonicity of liquid compositions of the present invention and include polyhydric sugar alcohols, preferably trihydric or higher sugar alcohols, such as glycerin. erythritol, arabitol, xylitol, sorbitol and mannitol. Polyhydric alcohols can be present in an amount between 0.1% to 25% by weight, preferably 1% to 5% taking into account the relative amounts of the other ingredients.

Stabilizers refer to a broad category of excipients which can range in function from a bulking agent to an additive which solubilizes the therapeutic agent or helps to prevent denaturation or adherence to the container wall. Typical stabilizers can be polyhydric sugar alcohols (enumerated above); amino acids such as arginine. lysine. glycine. glutamine. asparagine. histidine, alanine, ornithine. L-leucine. 2-phenylalanine, glutamic acid. threonine, etc., organic sugars or sugar alcohols. such as lactose. trehalose. stachyose, mannitol. sorbitol. xylitol. ribitol, myoinisitol, galactitol, glycerol and the like. including cyclitols such as inositol; polyethylene glycol; amino acid polymers; sulfur containing reducing agents. such as urea. glutathione. thiocitic acid, sodium thioglycolate, thioglycerol, α-monothioglycerol and sodium thio sulfate; low molecular weight polypeptides (i.e. < 10 residues); proteins such as human serum albumin. bovine serum albumin. gelatin or immunoglobulins; hydrophilic polymers. such as polyvinylpyrrolidone monosaccharides. such as xylose, mannose. fructose, glucose; disaccharides such as lactose. maltose. sucrose and trisaccacharides such as raffinose; polysaccharides such as dextran. Stabilizers can be present in the range from 0.1 to 10.000 weights per part of weight active protein.

Non-ionic surfactants or detergents (also known as "wetting agents") are present to help solubilize the therapeutic agent as well as to protect the therapeutic protein against agitation-induced aggregation, which also permits the formulation to be exposed to shear surface stressed without causing denaturation of the protein. Suitable non-ionic surfactants include polysorbates (20, 80, etc.). polyoxamers (184. 188 etc.), Pluronic® polyols, polyoxyethylene sorbitan monoethers (Tween®-20, Tween®-80, etc.). Non-ionic surfactants are present in a range of about 0.05 mg/ml to about 1.0 mg/ml, preferably about 0.07 mg/ml to about 0.2 mg/ml.

Additional miscellaneous excipients include bulking agents, (e.g. starch). chelating agents (*e.g.* EDTA). antioxidants (e.g., ascorbic acid, methionine, vitamin E), and cosolvents.

The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely

affect each other. For example, it may be desirable to further provide an immunosuppressive agent. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsule prepared, for example, by coascervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsule and poly-(methylmethacylate) microcapsule, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in *Remington's Pharmaceutical Sciences*, 16th edition, A. Osal, Ed. (1980).

The formulations to be used for in vivo administration must be sterile. This is readily accomplished, for example, by filtration through sterile filtration membranes.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semi-permeable matrices of solid hydrophobic polymers containing the compounds of the invention, which matrices are in the form of shaped articles, *e.g.*, films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No.3,773,919), copolymers of L-glutamic acid and ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT™ (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated compounds of the invention remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

The amount of therapeutic polypeptide, antibody or fragment thereof which will be effective in the treatment of a particular disorder or condition will depend on the nature of the disorder or condition, and can be determined by standard clinical techniques. Where possible, it is desirable to determine the dose-response curve and the pharmaceutical compositions of the invention first *in vitro*, and then in useful animal model systems prior to testing in humans. However, based on common knowledge of the art, a pharmaceutical composition effective in modulating *Dhh* and *Hh* signaling may provide a local *patched*-2 protein concentration of between about 10 and 1000 ng/ml, preferably between 100 and 800 ng/ml and most preferably between about 200 ng/ml and 600 ng/ml of *Ptch*-2 (SEQ ID NO:2).

In a preferred embodiment, an aqueous solution of therapeutic polypeptide, antibody or fragment thereof is administered by subcutaneous injection. Each dose may range from about 0.5 μg to about 50 μg per kilogram of body weight, or more preferably, from about 3 μg to about 30 μg per kilogram body weight.

The dosing schedule for subcutaneous administration may vary from once a week to daily depending on a number of clinical factors, including the type of disease, severity of disease, and the subject's sensitivity to the therapeutic agent.

Patched-2 polypeptide may comprise an amino acid sequence or subsequence thereof as indicated in Fig. 1, active amino acid sequence derived therefrom, or functionally equivalent sequence as this subsequence is believed to comprise the functional portion of the *patched*-2 polypeptide.

If the subject manifests undesired side effects such as temperature elevation, cold or flu-like symptoms, fatigue, etc., it may be desirable to administer a lower dose at more frequent intervals. One or more additional drugs may be administered in combination with *patched*-2 to alleviate such undesired side effects, for example, an anti-pyretic, anti-inflammatory or analgesic agent.

The following examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way.

All patent and literature references cited in the present specification are hereby incorporated by reference in their entirety.

EXAMPLES

Commercially available reagents referred to in the examples were used according to manufacturer's instructions unless otherwise indicated. The source of those cells identified in the following examples, and throughout the specification, by ATCC accession numbers is the American Type Culture Collection, Rockville, Maryland.

EXAMPLE 1

Introduction

At the cell surface, *Hh* function appears to be mediated by a multicomponent receptor complex involving *Ptch* and *Smo* (SEQ ID NO:17), two multi-transmembrane proteins initially identified as segment polarity genes in *Drosophila* and later characterized in vertebrates. Nakano, Y. *et al.*, *Nature* 341: 508-513 (1989); Goodrich *et al.*, *Gene Dev.* 10: 301-312 (1996); Marigo *et al.*, *Develop.* 122: 1225-1233 (1996); van den Heuvel *et al.*, *Nature* 382: 547-551 (1996); Alcedo *et al.*, *Cell* 86: 221-232 (1996); Stone *et al. Nature* 384: 129-34 (1996). Both genetic and biochemical evidence support the existence of a receptor complex where *Ptch* (SEQ ID NO:4) is the ligand binding subunit, and where *Smo* (SEQ ID NO:17), a G-protein coupled receptor-like molecule, is the signaling component. Stone *et al.*, *Nature* 384: 129-134 (1996), Marigo *et al.*, *Nature* 384: 176-79 (1996), Chen *et al.*, *Cell* 87: 553-63 (1996). Upon binding of *Hh* to *Ptch* (SEQ ID NO:4), the normal inhibitory effect of *Ptch* (SEQ ID NO:4) on *Smo* (SEQ ID NO:17) is relieved, allowing *Smo* (SEQ ID NO:17) to transduce the *Hh* signal across the plasma membrane.

Results

It remains to be established if the *PTCH/SMO* receptor complex mediates the action of all 3 mammalian *Hhs* or if specific components exist. Recently, a second murine *Patched* gene, *mPatched-2* (SEQ ID NO:7) was recently isolated [Motoyama *et al.*, *Nature Genet.* 18: 104-106 (1998)] but its function as a *Hh* receptor has not been established. In order to characterize *patched*-2 (SEQ ID NO:2)and compare it to *Patched* (SEQ ID NO:4) with respect to the biological function of the various *Hh* family members, we have screened EST databases with the *Patched* (SEQ ID NO:4) protein and identified 2 EST candidates for a

novel human *patched* gene. A full length cDNA encoding human *Ptch-2* (SEQ ID NO:2) was cloned from a testis library. The initiation ATG defines a 3612 nucleotide open reading frame encoding a 1204 amino acid long protein with a predicted molecular weight of approximately 131 kDa. The overall identity between human *Ptch* (SEQ ID NO:4) and *Ptch-2* (SEQ ID NO:2) is 54% (Fig. 1), while the identity between human *PTCH*-2 and the recently described mouse *Ptch*-2 (SEQ ID NO:7) is 90% (Fig. 8). The most obvious structural difference between the two human *Patched* proteins is a truncated C-terminal cytoplasmic domain in *Ptch-2* (SEQ ID NO:2). In addition, only one of the two glycosylation sites present in *Ptch* (SEQ ID NO:4) is conserved in *Ptch*-2 (SEQ ID NO:2).

To determine if *Patched*-2 is a *Hh* receptor and if the two *Patched* molecules are capable of discriminating between the various *Hh* ligands through specific binding, Applicants transfected human 293 embryonic kidney cells with *Ptch* (SEQ ID NO:4) or *Ptch-2* (SEQ ID NO:2) expression constructs and analyzed the cells for binding of *Shh*, *Dhh* and *Ihh*. As shown on Figure 7A, binding of ^{125}I-*Shh* can be competed with an excess of *Shh*, *Dhh* or *Ihh* (SEQ ID NOS: 14, 13 and 29), respectively. Scatchard analysis of the displacement curves indicates that all *Hhs* have similar affinity for *Ptch* (SEQ ID NO:4) (*Shh*, 1.0nM (SEQ ID NO:14); *Dhh*, 2.6nM (SEQ ID NO:13); *Ihh*, 1.0nM (SEQ ID NO:29) and *Ptch*-2 (SEQ ID NO:2) (*Shh*, 1.8nM (SEQ ID NO:14); *Dhh*, 0.6nM (SEQ ID NO:13); *Ihh*, 0.4nM (SEQ ID NO:29) indicating that both *PTCH* (SEQ ID NO:4) and *PTCH*-2 (SEQ ID NO:2) can serve as physiological receptors for the 3 mammalian *Hh* proteins.

Applicants next determined whether, like *Patched*, *Patched*-2 forms a physical complex with *Smo* (SEQ ID NO:17). Expression constructs for Flag-tagged *Ptch* (SEQ ID NO:4) or *Ptch-2* (SEQ ID NO:2) were transiently co-transfected in 293 cells with Myc-tagged *Smo* (SEQ ID NO:17). As described previously [Stone *et al.*, *Nature* 384: 129-34 (1996)], in cells expressing *Ptch* (SEQ ID NO:4) and *Smo* (SEQ ID NO:15), *Ptch* (SEQ ID NO:4) can be immunoprecipitated with antibodies against the epitope-tagged *Smo* (SEQ ID NO:15) (Fig. 7B). Similarly, *Patched*-2 can be immunoprecipitated with antibodies against the epitope-tagged *Smo* (SEQ ID NO:15) when the two proteins are co-expressed in 293 cells. Together, these results suggest a model where *Patched*-2 forms a multicomponent *Hh* receptor complex with *Smo* (SEQ ID NO:17) similar to the one described for *PTCH* (Stone *et al., supra*). Interestingly, these results also demonstrate that the long C-terminal tail which is missing in *Patched*-2 is not required for the interaction with *Smo* (SEQ ID NO:17) as was already suggested by the analysis of truncated *Patched* (Stone *et al., supra*). However, it remains possible that the absence of a C-terminal domain affects the capacity of *Patched*-2 to block signaling by *Smo* (SEQ ID NO:17) or leads to difference in signaling by *Patched* compared to *Patched*-2.

To further investigate whether *Patched*-2 could mediate the action of a specific Hh molecule based on its expression profile, Applicants have compared the expression pattern of *Ptch* (SEQ ID NO:4) and *Ptch-2* (SEQ ID NO:2). First, Northern blot analysis using a probe specific for *Ptch*-2 (SEQ ID NO:1) revealed high levels of *PTCH2* mRNA in the testis (Fig. 4). By this method, *Ptch*-2 (SEQ ID NO:1) expression was not detected in any other tissue analyzed including embryonic tissues (data not shown). This profile is very different from the one observed for *Ptch* (SEQ ID NO:18) which was not found in testis by Northern blot but in a large number of adult and embryonic tissues [Goodrich *et al.*, *Genes Dev.* 10: 301-312

(1996)]. More detailed analysis of the expression pattern of *Ptch* (SEQ ID NO:18) and *Ptch*-2 (SEQ ID NO:1) was performed by *in situ* hybridization with particular attention to testis. As previously described (Motoyama *et al., supra*), low levels of *Ptch-2* (SEQ ID NO:1) expression were detected in epithelial cells of the developing tooth and skin (data not shown). High levels of *Ptch-2* (SEQ ID NO:2) encoding mRNA are expressed inside the seminiferous tubule, on the primary and secondary spermatocytes (Fig. 6B,6E) while only low levels of *Ptch* (SEQ ID NO:4) encoding mRNA can be detected on the Leydig cells located in the interstitium of the seminiferous tubules (Fig. 6A). The primary and secondary spermatocytes are in close contact with the supporting Sertoli cells, the source of *Dhh* (SEQ ID NO:13) in the testis [Bitgood *et al., Curr. Biol.* 6: 298-304 (1996)]. To determine which one of the 2 receptors is the most relevant mediator of *Dhh* (SEQ ID NO:13) activity in the testis, we have analyzed the expression profile of FuRK (SEQ ID NO:10), a Fused Related Kinase that is believed to be a component of the *Hh* signaling pathway. Consistent with the idea that *Patched*-2 is the target of *Dhh* in the testis, we found that FuRK (SEQ ID NO:10) is expressed only in germ cells where it colocalizes with *Ptch*-2 (SEQ ID NO:2) (Figure 4c,f). *Dhh* (SEQ ID NO:13) is required for proper differentiation of germ cells since male *Dhh*-deficient mice are sterile due to lack of mature sperm (Bitgood *et al., supra*). Our data suggest that *Dhh* (SEQ ID NO:13) acts directly on germ cells through *Ptch*-2 (SEQ ID NO:2) while the function of *Ptch* (SEQ ID NO:4) expressed at low levels on testosterone producing Leydig cells is unclear.

Discussion

Loss of heterozygosity (LOH) for *Patched* was reported to occur with high frequency in familial as well as sporadic basal cell carcinoma [Johnson *et al.*. *Science* 272: 1668-71 (1996); Hahn *et al.*, *Cell* 85: 841-51 (1996); Gailani *et al.*. *Nature Genetics* 14: 78-81; Xie *et al.*, *Cancer Res.* 57: 2369-72 (1997)], suggesting that it functions as a tumor suppressor. According to the receptor model described above, loss of *Patched* function may result in aberrant signaling by *Smo* (SEQ ID NO:17), leading to hyperproliferation of the skin basal cell layer. If, as suggested above, *Patched-2* mediates the function of *Dhh*, loss of *Patched-2* may lead to tumor formation in tissues where *Smo* (SEQ ID NO:17) activity is controlled by *Patched-2*. The gene encoding *Ptch*-2 (SEQ ID NO:2) was mapped by fluorescence *in situ* hybridization and by PCR using a radiation hybrid panel to human chromosome 1p33-34 (data not shown). Interestingly, recent analysis of recurrent chromosomal abnormalities in testicular tumors, including seminomas, revealed a deletion of the region 1p32-36 [Summersgill *et al.*, *B. J. Cancer* 77: 305-313 (1998)]. Loss of this region encompassing the *Patched-2* locus was consistent in 36% of the germ cell tumor cases. These data raise the possibility that, like *Patched* in basal cell carcinoma and medulloblastoma, *Patched*-2 may be a tumor suppressor in *Dhh* (SEQ ID NO: 13) target cells such as spermatocytes, further implicating *Hh* signaling in cancer.

In summary, our data demonstrate that both *Patched* and *Patched*-2 are genuine *Hh* receptors and that they are both capable of forming a complex with *Smo* (SEQ ID NO:17). Although binding data indicate that *Patched* and *Patched*-2 do not discriminate between the various *Hh* ligands through affinity differences, the distinct tissue distribution of these 2 receptors suggests that *in vivo*, *Patched* may be the primary receptor for *Shh* whereas *Patched*-2 will mediate mainly *Dhh* signaling. The function of *Patched* expression in Leydig cells in the absence of some of the *Hh* signaling components remain to be explained. Similarly, it will be of interest to determine if *Patched-2* plays a role when expressed in *Shh* expressing cells present in the

developing tooth and skin Motoyama *et al.*, *Nature Genet.* 18: 104-106 (1998). Finally, the existence of *Patched*-2 raises the question of whether additional *patched* receptors exist. in particular one that mediates the function of *Ihh* (SEQ ID NO: 29).

Material and Methods

1. Isolation of human patched-2 cDNA clones

An expressed sequence tag (EST) DNA database (LIFESEQ™, Incyte Pharmaceuticals. Palo Alto, CA) was searched for a human homologue of the Drosophila segment polarity gene patched-2. Two ESTs (Incyte #905531 and 1326258) (Fig. 2) were identified as a potential candidates. In order to identify human cDNA libraries containing human patched-2 clones, human cDNA libraries in pRK5 were first screened by PCR using the following primers:

5'-905531(A): 5'-AGGCGGGGATCACAGCA-3' (SEQ ID NO:19)

3'-905531(A): 5'-ATACCAAAGAGTTCCACT-3' (SEQ ID NO:20)

A fetal lung library was selected and enriched for *patched*-2 cDNA clones by extension of single stranded DNA from plasmid libraries grown in dut⁻/ung⁻ host using the 3'-905531(A) primer in a reaction containing 10μl of 10x PCR Buffer (Klentaq®), 1μl dNTP (200 μM), 1 μl library DNA (200 ng), 0.5 μl primer, 86.5 μl H_2O and 1 μl of Klentaq® (Clontech) added after a hot start. The reaction was denatured for 1 min. at 95°C, annealed for 1 min. at 60°C then extended for 20 min. at 72°C. DNA was extracted with phenol/$CHCl_3$, ethanol precipitated, then transformed by electroporation into DH10B (Gibco/BRL) host bacteria. Colonies from each transformation were replica plated on nylon membranes and screened with an overlapping oligo probe derived from the EST sequence (#905531) of the following sequence:

5'-ptch2 probe: 5'-CTGCGGCGCTGCTTCCTGCTGGCCGTCTGCATCCTGCTGGTGTGC-3

(SEQ ID NO:21)

3'-ptch2probe: 5'-AGAGCACAGACGAGGAAAGTGCACACCAGCAGGATGCAGACGGCC-3'

(SEQ ID NO:22)

The oligo probe was labeled with [γ-^{32}P]-ATP and T4 polynucleotide kinase. Filters were hybridized overnight at 42°C in 50% formamide, 5xSSC, 10xDenhardt's, 0.05M sodium phosphate (pH 6.5), 0.1% sodium pyrophosphate, 50 μg/ml of sonicated salmon sperm DNA. The filters were then rinsed in 2x SSC and washed in 0.1x SSC, 0.1% SDS then exposed to Kodak® X Ray films. Using this procedure, a partial clone was isolated from the fetal brain library (clone 3A – Fig. 10) (SEQ ID NO:8). In order to isolate the missing 5'-sequence, a testis library (see northern blot analysis, infra) was screened. The primer set used to amplify a 204 bp probe from clone 3A to probe the testis library was:

RACE 5: 5'-ACTCCTGACTTGTAGCAGATT-3' (SEQ ID NO. 23) and

RACE 6: 5'-AGGCTGCATACACCTCTCAGA-3' (SEQ ID NO:24).

The amplified probe was purified by excision from an agarose gel and labeled with a random primer labeling kit (Boehringer Mannheim). Several clones were isolated, including one (clone 16.1 – Fig. 11) (SEQ ID NO:9) containing a potential initiation methionine. A full length cDNA encoding a *Patched-2* was reconstructed by assembling several of these clones. The full length cDNA encoding human *Ptch*-2 (Fig. 1

(SEQ ID NO:1)) has a 3612 nucleotide long open reading frame encoding a 1204 amino acid protein with a 144 kDa predicted molecular weight. Alignment with human *Ptch* (SEQ ID NO:4) reveals a 53% identity between the 2 molecules at the amino acid level (Fig. 3). All 12 transmembrane domains are conserved. The most significant difference is a shorter C-terminal intracellular domain in *Ptch*-2 (SEQ ID NO:2) compared to *Ptch* (SEQ ID NO:4).

2. *Northern blot analysis:*

In order to determine the best tissue source for isolation of the complete full length *Patched*-2 cDNA as well as to determine its expression profile, we probed human multiple tissue northern blots (Clontech) with a 752 bp fragment amplified from the 3' untranslated region of *Patched*-2 using the following primers:

TM2: TM2 5-GCTTAGGCCCGAGGAGAT-3' (SEQ ID NO:25)

UTR2: 5'-AACTCACAACTTTCTCTCCA-3' (SEQ ID NO:26).

The resulting fragment was gel purified and labeled by random priming. The blots were hybridized in ExpressHyb® hybridization solution (Clontech) in the presence of 1 x 10^6 cpm/ml ^{32}P-labeled probe at 42°C overnight. The blots were washed in 2x SSC at room temperature for 10 minutes and washed in 0.1 x SSC/0.1 % SDS at 42°C for 30 minutes then exposed to x-ray film overnight. Fig. 4 shows that Ptch-2 message is expressed at high levels in only the testis.

3. *Chromosomal localization:*

The primers TM2 (SEQ ID NO:25) and UTR2 (SEQ ID NO:26) described above were used to screen the Genome Systems (St. Louis, MO) BAC library. Two individual BAC clones were obtained from this library and chromosomal localization of both of the clones by FISH indicated that *Ptch-2* (SEQ ID NO:2) maps to human chromosome 1p33-34 (FIG 5). Loss of heterozyosity (LOH) for *Patched* was reported to occur with high frequency in basal cell carcinoma . Loss of *Patched* function is thought to lead to constitutive signaling by Smoothened (*Smo*) (SEQ ID NO:17), resulting in hyperproliferation of the basal layer of the dermis. A similar mechanism may lead to the formation of germ cell tumors. This model proposes that the first step in the progression of a germ cell tumor is an initial loss of DNA by a germ cell precursor, leading to a neoplastic germ cell which then forms a seminoma [De Jong *et al.*, *Cancer Genet. Cytogenet.* 48: 143-167 (1990)]. From the invasive seminoma, all other forms of germ cell tumor types develop. Approximately 80% of all germ cell tumors correlate with an isochromosome 12p (i12p) and is found at a higher frequency in non-seminomas than seminomas [Rodriguez *et al.*, *Cancer Res.* 52: 2285-2291 (1992)]. However, analysis of recurrent chromosomal abnormalities in testicular tumors including seminomas revealed a deletion of the region 1p32-36. Loss of this region was consistent in 36% of the germ cell tumor cases of in a recent study Summersgill *et al.*, *B. J. Cancer* 57: 305-313 (1998)]. A similar deletion of chromosome 1p32-36 has been reported at a frequency of 28% in oligodendrogliomas; Bello, *et al.*, *Int. J. Cancer* 57: 172-175 (1994). While expression of *Ptch*-2 (SEQ ID NO:2) in the brain was not examined here in detail, *Ptch*-2 (SEQ ID NO:2) is thought to be the *Dhh* receptor (see below) and expression of *Dhh* by murine Schwann cells was previously reported [Bitgood *et al*, *Develop. Biol.* 172: 126-138 (1995)]. Since *Ptch*-2 (SEQ ID NO:2) localizes to chromosome 1p33-34 it is possible that *Patched*-2

regulates *Smo* (SEQ ID NO:17) signaling in *Dhh* target cells and that loss of *Patched*-2 function leads to abnormal *Smo* (SEQ ID NO:17) signaling in these cells and subsequent tumor formation.

4. *In situ hybridization:*

Mouse testis sections were cut at 16 μm, and processed for *in situ* hybridization by the method described in Phillips *et al.*, *Science* 250: 290-294 (1990). ^{33}P-UTP labeled RNA probes were generated as described in Melton *et al.*. *Nucleic Acids Res.* 12: 7035-7052 (1984). Sense and antisense probes were synthesized from the 3' non-coding region of the mouse *Patched* or *Patched*-2 and from a mouse FuRK cDNA fragment corresponding to the region encoding amino acid 317-486 of the human sequence using T3 and T7, respectively.

PTCH:

503 (Anti-sense)

5'GGATTCTAATACGACTCACTATAGGGCCCAATGGCCTAAACCGACTGC3' (SEQ ID NO:27)

503 (Sense)

5'CTATGAAATTAACCCTCACTAAAGGGACCCACGGCCTCTCCTCACA3' (SEQ ID NO:28)

PTCH2:

504 (Anti-sense)

5'GGATTCTAATACGACTCACTATAGGGCCCCTAAACTCCGCTGCTCCAC3' (SEQ ID NO:12)

504 (Sense)

5'CTATGAAATTAACCCTCACTAAAGGGAGCTCCCGTGAGTCCCTATGTG3' (SEQ ID NO:11)

FuRK sense and antisense were synthesized from a mouse fused DNA fragment using T3 and T7, respectively, corresponding to the region encoding amino acid residues 317-486 of the human.

Figure 6 illustrates that, although both *Ptch* (SEQ ID NO:4) and *Ptch*-2 (SEQ ID NO:2) are expressed in testis, their expression pattern does not overlap. *Ptch* (SEQ ID NO:4) is expressed in the Leydig cells of the interstitium while *Ptch*-2 (SEQ ID NO:2) is expressed in the primary and secondary spermatocytes.

The expression of *Patched*-2 specifically in the developing spermatogonia suggest that *Patched*-2 is the immediate target of *Dhh* (SEQ ID NO. 13). *Dhh* (SEQ ID NO. 13) is expressed by Sertoli cells and mice deficient in *Dhh* (SEQ ID NO. 13) are sterile because of a defect in sperm production [Bitgood *et al.*, *Curr. Biol.* 6: 298-304 (1996)]. Although this effect on germ cells was though to be indirect and mediated by *Patched* present on Leydig cells, our data suggest that *Dhh* directly acts on germ cells through *Patched*-2. This is further demonstrated by the localization of FuRK (SEQ ID NO. 10), an intracellular kinase homologous to *Drosophila* Fused and involved in transducing the Hedgehog (*Hh*) signal. As shown in Figure 6, FuRK (SEQ ID NO:10) is colocalizes with *Ptch-2* (SEQ ID NO:2) in germ cells and not with *Ptch* (SEQ ID NO:4) in Leydig cells, suggesting that *Patched-2* and not *Patched* will be able to transduce the *Dhh* signal. These results suggest that *Patched*-2 is a *Dhh* receptor.

Ptch-2 mRNA levels in Smo-M2 (SEQ ID NO. 16) transgenic mice [a Smo mutation which results in autonomous phenotypes similar to BCC, Xie *et al.*. *Nature* 391: 90-92 (1998)] can be increased upon

abnormal activation of the *Hh* signaling pathway. As indicated in Fig. 9, *Ptch-2* (SEQ ID NO:2) levels were high in tumor cells (although lower than *Ptch* (SEQ ID NO:4) levels). This suggests that antibodies directed toward *Ptch-2* (SEQ ID NO:2) may be useful in the treatments of BCC.

5. *Immunoprecipitation with Smo:*

The binding of *Patched*-2 to *Smo* (SEQ ID NO:17) was assessed by cotransfection using a transient transfection system of a myc-epitope tagged *Smo* (SEQ ID NO:15) and a FLAG-epitope tagged *Patched or Patched-2* expression construct in 293 cells using standard techniques (Gorman, C., *DNA Cloning: A Practical Approach*, Clover, DM ed., Vol. II, pp. 143-190, IRL Press, Washington, D.C.). 36 hours after transfection, the cells were lysed in 1% NP-40 and immunoprecipitated overnight with the 9E10 anti-myc antibody or with the M2 anti-FLAG antibody (IBI-Kodak) followed by protein A Sepharose, and then separated on a denatured 6% polyacrylamide gel. Proteins were detected by transfer to nitrocellulose and probing with antibodies to Flag or Myc epitopes, using the ECL detection system (Amersham). Figure 7B indicates that both *Ptch* (SEQ ID NO:4) or *Ptch*-2 (SEQ ID NO:2) are expressed at the same level (IP Flag, Blot Flag) and that like *Ptch* (SEQ ID NO:4), *Ptch-2* (SEQ ID NO:2) forms a physical complex with *Smo* (SEQ ID NO:17). These results suggest that like *Patched*, *Patched*-2 controls *Hh* signaling through its interaction with *Smo* (SEQ ID NO:17).

6. *Hh Binding:*

To determine whether *Patched*-2 is able to bind to the various hedgehog ligands, 293 cells were transfected with *Ptch* (SEQ ID NO:4) or *Ptch-2* (SEQ ID NO:2) using standard procedures. Cells were incubated with 100 pM ^{125}I-*Shh* (19kD amino terminal fragment of murine *Shh* (SEQ ID NO:14)) in the presence or absence of excess unlabeled *Shh* (SEQ ID NO:14) or *Dhh* (SEQ ID NO:13) for 2h at room temperature. After equilibrium was reached, the ligand bound cells were centrifuged through a continuous sucrose gradient to separate unincorporated and then counted in a scintillation counter. Figure 7A shows that both *Dhh* (SEQ ID NO:13) and *Shh* (SEQ ID NO:14) bind to *Ptch* (SEQ ID NO:4) and *Ptch-2* (SEQ ID NO:2). Varying concentrations of cold competitor indicate that the 2 ligands have similar affinity for *Ptch* (SEQ ID NO:4) and *Ptch-2* (SEQ ID NO:2).

Example 2

Expression of *patched-2* in *E. coli*

The DNA sequence encoding human *patched-2* is initially amplified using selected PCR primers. The primers should contain restriction enzyme sites that correspond to the restriction enzyme sites on the selected expression vector. A variety of expression vectors may be employed. An example of a suitable vector is pBR322 (derived from *E. coli*; see Bolivar *et al., Gene*, 2:95 (1977)) which contains genes for ampicillin and tetracycline resistance. The vector is digested with restriction enzyme and dephosphorylated. The PCR amplified sequences are then ligated into the vector. The vector will preferably include sequences that encode for an antibiotic resistance gene, a trp promoter, a polyhis leader (including the first six STII codons, polyhis sequence, and enterokinase cleavage site), the vertebrate patched-2 coding region, lambda transcriptional terminator, and an argU gene.

The ligation mixture is then used to transform a selected *E. coli* strain using the methods described in Sambrook *et al., supra*. Transformants are identified by their ability to grow on LB plates and antibiotic

resistant colonies are then selected. Plasmid DNA can be isolated and confirmed by restriction analysis and DNA sequencing.

Selected clones can be grown overnight in liquid culture medium such as LB broth supplemented with antibiotics. The overnight culture may subsequently be used to inoculate a larger scale culture. The cells are then grown to a desired optical density, during which the expression promoter is turned on.

After culturing the cells for several more hours, the cells can be harvested by centrifugation. The cell pellet obtained by the centrifugation can be solubilized using various agents known in the art, and the solubilized vertebrate *patched-2* protein can then be purified using a metal chelating column under conditions that allow tight binding of the protein.

Example 3

Expression of patched-2 in mammalian cells

The vector, pRK5 (see EP 307,247, published March 15, 1989), is employed as the expression vector. Optionally, the vertebrate patched-2 DNA is ligated into pRK5 with selected restriction enzymes to allow insertion of the vertebrate patched-2 DNA using ligation methods such as described in Sambrook *et al., supra*. The resulting vector is called pRK5-patched-2.

In one embodiment, the selected host cells may be 293 cells. Human 293 cells (ATCC CCL 1573) are grown to confluence in tissue culture plates in medium such as DMEM supplemented with fetal calf serum and optionally, nutrient components and/or antibiotics. About 10 μg pRK5-patched-2 DNA is mixed with about 1 μg DNA encoding the VA RNA gene [Thimmappaya *et al., Cell.* 31:543 (1982)] and dissolved in 500 μl of 1 mM Tris-HCl, 0.1 mM EDTA, 0.227 M $CaCl_2$. To this mixture is added, dropwise, 500 μl of 50 mM HEPES (pH 7.35), 280 mM NaCl, 1.5 mM $NaPO_4$, and a precipitate is allowed to form for 10 minutes at 25°C. The precipitate is suspended and added to the 293 cells and allowed to settle for about four hours at 37°C. The culture medium is aspirated off and 2 ml of 20% glycerol in PBS is added for 30 seconds. The 293 cells are then washed with serum free medium. fresh medium is added and the cells are incubated for about 5 days.

Approximately 24 hours after the transfections, the culture medium is removed and replaced with culture medium (alone) or culture medium containing 200 μCi/ml ^{35}S-cysteine and 200 μCi/ml ^{35}S-methionine. After a 12 hour incubation, the conditioned medium is collected, concentrated on a spin filter, and loaded onto a 15% SDS gel. The processed gel may be dried and exposed to film for a selected period of time to reveal the presence of vertebrate *patched-2* polypeptide. The cultures containing transfected cells may undergo further incubation (in serum free medium) and the medium is tested in selected bioassays.

In an alternative technique, vertebrate *patched-2* may be introduced into 293 cells transiently using the dextran sulfate method described by Somparyrac *et al., Proc. Natl. Acad. Sci.,* 12:7575 (1981). 293 cells are grown to maximal density in a spinner flask and 700 μg pRK5-*patched-2* DNA is added. The cells are first concentrated from the spinner flask by centrifugation and washed with PBS. The DNA-dextran precipitate is incubated on the cell pellet for four hours. The cells are treated with 20% glycerol for 90 seconds, washed with tissue culture medium, and re-introduced into the spinner flask containing tissue culture medium, 5 μg/ml bovine insulin and 0.1 μg/ml bovine transferrin. After about four days, the

conditioned media is centrifuged and filtered to remove cells and debris. The sample containing expressed vertebrate *patched-2* can then be concentrated and purified by any selected method, such as dialysis and/or column chromatography.

In another embodiment, vertebrate *patched-2* can be expressed in CHO cells. The pSVi-*patched-2* can be transfected into CHO cells using known reagents such as $CaPO_4$ or DEAE-dextran. As described above, the cell cultures can be incubated, and the medium replaced with culture medium (alone) or medium containing a radiolabel such as ^{35}S-methionine. After determining the presence of vertebrate *patched-2* polypeptide, the culture medium may be replaced with serum free medium. Preferably, the cultures are incubated for about 6 days, and then the conditioned medium is harvested. The medium containing the expressed vertebrate *patched-2* can then be concentrated and purified by any selected method.

Epitope-tagged vertebrate *patched-2* may also be expressed in host CHO cells. The vertebrate *patched-2* may be subcloned out of the pRK5 vector. The subclone insert can undergo PCR to fuse in frame with a selected epitope tag such as a poly-his tag into an expression vector. The poly-his tagged vertebrate *patched-2* insert can then be subcloned into a SV40 driven vector containing a selection marker such as DHFR for selection of stable clones. Finally, the CHO cells can be transfected (as described above) with the SV40 driven vector. Labeling may be performed, as described above, to verify expression. The culture medium containing the expressed poly-His tagged vertebrate *patched-2* can then be concentrated and purified by any selected method, such as by Ni^{2+}-chelate affinity chromatography.

Example 4

Expression of vertebrate *patched-2* in Yeast

The following method describes recombinant expression of vertebrate *patched-2* in yeast.

First, yeast expression vectors are constructed for intracellular production or secretion of vertebrate *patched-2* from the ADH2/GAPDH promoter. DNA encoding vertebrate *patched-2*: a selected signal peptide and the promoter is inserted into suitable restriction enzyme sites in the selected plasmid to direct intracellular expression of vertebrate *patched-2*. For secretion, DNA encoding vertebrate *patched-2* can be cloned into the selected plasmid, together with DNA encoding the ADH2/GAPDH promoter, the yeast alpha-factor secretory signal/leader sequence, and linker sequences (if needed) for expression of vertebrate *patched-2*.

Yeast cells, such as yeast strain AB110, can then be transformed with the expression plasmids described above and cultured in selected fermentation media. The transformed yeast supernatants can be analyzed by precipitation with 10% trichloroacetic acid and separation by SDS-PAGE, followed by staining of the gels with Coomassie Blue stain.

Recombinant vertebrate *patched-2* can subsequently be isolated and purified by removing the yeast cells from the fermentation medium by centrifugation and then concentrating the medium using selected cartridge filters. The concentrate containing vertebrate *patched-2* may further be purified using selected column chromatography resins.

Example 5

Expression of vertebrate *patched-2* in Baculovirus-Infected Insect Cells

The following method describes recombinant expression of vertebrate *patched-2* in Baculovirus-infected insect cells.

The vertebrate *patched-2* is patched-2 upstream of an epitope tag contained within a baculovirus expression vector. Such epitope tags include poly-his tags and immunoglobulin tags (like Fc regions of IgG). A variety of plasmids may be employed, including plasmids derived from commercially available plasmids such as pVL1393 (Novagen). Briefly, the vertebrate *patched-2* or the desired portion of the vertebrate *patched-2* (such as the sequence encoding the extracellular domain of a transmembrane protein) is amplified by PCR with primers complementary to the 5' and 3' regions. The 5' primer may incorporate flanking (selected) restriction enzyme sites. The product is then digested with those selected restriction enzymes and subcloned into the expression vector.

Recombinant baculovirus is generated by co-transfecting the above plasmid and BaculoGold™ virus DNA (Pharmingen) into *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711) using lipofectin (commercially available from GIBCO-BRL). After 4 - 5 days of incubation at 28°C, the released viruses are harvested and used for further amplifications. Viral infection and protein expression is performed as described by O'Reilley et al., Baculovirus expression vectors: A laboratory Manual, Oxford: Oxford University Press (1994).

Expressed poly-his tagged vertebrate *patched-2* can then be purified, for example, by Ni^{2+}-chelate affinity chromatography as follows. Extracts are prepared from recombinant virus-infected Sf9 cells as described by Rupert *et al.*, *Nature*, 362:175-179 (1993). Briefly, Sf9 cells are washed, resuspended in sonication buffer (25 mL Hepes, pH 7.9; 12.5 mM $MgCl_2$; 0.1 mM EDTA; 10% Glycerol; 0.1% NP-40; 0.4 M KCl), and sonicated twice for 20 seconds on ice. The sonicates are cleared by centrifugation, and the supernatant is diluted 50-fold in loading buffer (50 mM phosphate, 300 mM NaCl, 10% Glycerol, pH 7.8) and filtered through a 0.45 μm filter. A Ni^{2+}-NTA agarose column (commercially available from Qiagen) is prepared with a bed volume of 5 mL, washed with 25 mL of water and equilibrated with 25 mL of loading buffer. The filtered cell extract is loaded onto the column at 0.5 mL per minute. The column is washed to baseline A_{280} with loading buffer, at which point fraction collection is started. Next, the column is washed with a secondary wash buffer (50 mM phosphate; 300 mM NaCl, 10% Glycerol, pH 6.0), which elutes nonspecifically bound protein. After reaching A_{280} baseline again, the column is developed with a 0 to 500 mM Imidazole gradient in the secondary wash buffer. One mL fractions are collected and analyzed by SDS-PAGE and silver staining or western blot with Ni^{2+}-NTA-conjugated to alkaline phosphatase (Qiagen). Fractions containing the eluted His_{10}-tagged vertebrate *patched-2* are pooled and dialyzed against loading buffer.

Alternatively, purification of the IgG tagged (or Fc tagged) vertebrate *patched-2* can be performed using known chromatography techniques, including for instance, Protein A or protein G column

chromatography

Example 6

Preparation of Antibodies that Bind Vertebrate *patched-2*

This example illustrates preparation of monoclonal antibodies. which can specifically bind vertebrate *patched-2*.

Techniques for producing the monoclonal antibodies are known in the art and are described, for instance. in Goding, supra. Immunogens that may be employed include purified vertebrate *patched-2*, fusion proteins containing vertebrate *patched-2*. and cells expressing recombinant vertebrate *patched-2* on the cell surface. Selection of the immunogen can be made by the skilled artisan without undue experimentation.

Mice. such as Balb/c, are immunized with the vertebrate *patched-2* immunogen (*e.g.*, extracellular portions or cells expressed *ptch*-2) emulsified in complete Freund's adjuvant and injected subcutaneously or intraperitoneally in an amount from 1-100 micrograms. Alternatively, the immunogen is emulsified in MPL-TDM adjuvant (Ribi Immunochemical Research, Hamilton, MT) and injected into the animal's hind foot pads. The immunized mice are then boosted 10 to 12 days later with additional immunogen emulsified in the selected adjuvant. Thereafter, for several weeks, the mice may also be boosted with additional immunization injections. Serum samples may be periodically obtained from the mice by retro-orbital bleeding for testing in ELISA assays to detect vertebrate *patched-2* antibodies.

After a suitable antibody titer has been detected, the animals "positive" for antibodies can be injected with a final intravenous injection of vertebrate *patched-2*. Three to four days later. the mice are sacrificed and the spleen cells are harvested. The spleen cells are then patched-2 (using 35% polyethylene glycol) to a selected murine myeloma cell line such as P3X63AgU.1, available from ATCC, No. CRL 1597. The fusions generate hybridoma cells which can then be plated in 96 well tissue culture plates containing HAT (hypoxanthine, aminopterin, and thymidine) medium to inhibit proliferation of non-patched-2 cells, myeloma hybrids, and spleen cell hybrids.

The hybridoma cells will be screened in an ELISA for reactivity against vertebrate *patched-2*. Determination of "positive" hybridoma cells secreting the desired monoclonal antibodies against vertebrate *patched-2* is within the skill in the art.

The positive hybridoma cells can be injected intraperitoneally into syngeneic Balb/c mice to produce ascites containing the anti- vertebrate *patched-2* monoclonal antibodies. Alternatively, the hybridoma cells can be grown in tissue culture flasks or roller bottles. Purification of the monoclonal antibodies produced in the ascites can be accomplished using ammonium sulfate precipitation. followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can be employed.

Example 7

Gli Luciferase Assay

The following assay may be used to measure the activation of the transcription factor *GLI*, the mammalian homologue of the *Drosophila cubitus interruptus* (Ci). It has been shown that *GLI* is a transcription factor activated upon *SHh* stimulation of cells.

Nine (9) copies of a GLI binding site present in the HNF3β enhancer. (Sasaki *et al.*. *Development*

124: 1313-1322 (1997)). are introduced in front of a thymidine kinase minimal promoter driving the luciferase reporter gene in the pGL3 plasmid (Promega). The sequence of the *GLI* binding sequence is: TCGACAAGCAGGGAACACCCAAGTAGAAGCTC (p9XGliLuc) (SEQ ID NO:31), while the negative control sequence is: TCGACAAGCAGGGAAGTGGGAAGTAGAAGCTC (p9XmGliLuc) (SEQ ID NO:32). These constructs are cotransfected with the full length *Ptch*-2 and *Smo* in C3H10T1/2 cells grown in F12. DMEM (50:50). 10% FCS heat inactivated. The day before transfection 1 x 10^5 cells per well was inoculated in 6 well plates. in 2 ml of media. The following day, 1 µg of each construct is cotransfected in duplicate with 0.025 mg ptkRenilla luciferase plasmid using lipofectamine (Gibco-BRL) in 100 µl OptiMem (with GlutaMAX) as per manufacturer's instructions for 3 hours at 37°C. Serum (20%, 1 ml) is then added to each well and the cells were incubated for 3 more hours at 37°C. Cells are then washed twice with PBS, then incubated for 48 hours at 37°C in 2 ml of media. Each well is then washed with PBS. and the cells lysed in 0.5 ml Passive Lysis Buffer (Promega) for 15 min. at room temperature on a shaker. The lysate is transferred in eppendorf tubes on ice. spun in a refrigerated centrifuge for 30 seconds and the supernatant saved on ice. For each measure. 20 µl of cell lysate is added to 100 µl of LARII (luciferase assay reagent. Promega) in a polypropylene tube and the luciferase light activity measured. The reaction is stopped by the addition of Stop and Glow buffer (Promega), mixed by pipetting up and down 3 to 5 times and *Renilla* luciferase lights activity is measured on the luminometer.

Deposit of Material

The following materials have been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD, USA (ATCC):

Designation:	ATCC Dep. No.	Deposit Date
pRK7.hptc2.Flag-1405	209778	4/14/98

This deposit was made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and the Regulations thereunder (Budapest Treaty). This assures maintenance of a viable culture of the deposit for 30 years from the date of deposit. The deposit will be made available by ATCC under the terms of the Budapest Treaty, and subject to an agreement between Genentech, Inc. and ATCC, which assures permanent and unrestricted availability of the progeny of the culture of the deposit to the public upon issuance of the pertinent U.S. patent or upon laying open to the public of any U.S. or foreign patent application, whichever comes first. and assures availability of the progeny to one determined by the U.S. Commissioner of Patents and Trademarks to be entitled thereto according to 35 USC § 122 and the Commissioner's rules pursuant thereto (including 37 CFR § 1.14 with particular reference to 886 OG 638).

The assignee of the present application has agreed that if a culture of the materials on deposit should die or be lost or destroyed when cultivated under suitable conditions, the materials will be promptly replaced on notification with another of the same. Availability of the deposited material is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any government in accordance with its patent laws.

The foregoing written specification is considered to be sufficient to enable one skilled in the art to

practice the invention. The present invention is not to be limited in scope by the construct deposited, since the deposited embodiment is intended as a single illustration of certain aspects of the invention and any constructs that are functionally equivalent are within the scope of this invention. The deposit of material herein does not constitute an admission that the written description herein contained is inadequate to enable the practice of any aspect of the invention, including the best mode thereof, nor is it to be construed as limiting the scope of the claims to the specific illustrations that it represents. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and fall within the scope of the appended claims.

What is claimed is:

1. Isolated nucleic acid comprising DNA having at least a 95% sequence identity to (a) a DNA molecule encoding a *patched*-2 polypeptide comprising the sequence of amino acids 1 to about 1203 of Fig. 1 (SEQ ID NO:2). or (b) the complement of (a); and encoding a polypeptide having *patched*-2 biological activity.

2. An isolated nucleic acid comprising DNA having at least a 95% sequence identity to (a) a DNA molecule encoding the same mature polypeptide encoded by the cDNA in ATCC Deposit No. 209778 (designation: pRK7.hptc2.Flag-1405), or (b) the complement of the DNA molecule of (a).

3. (Amended) The isolated nucleic acid of claim 2 comprising human *patched-2* encoding sequence of the cDNA in ATCC deposit No. 209778 (designation: pRK7.hptc2.Flag-1405), or a sequence which hybridizes thereto under stringent conditions.

4. A vector comprising the nucleic acid of claim 1.

5. The vector of claim 4 operably linked to control sequences recognized by a host cell transformed with the vector.

6. A host cell transformed with the vector of claim 4.

7. The host cell of claim 6 which is mammalian.

8. The host cell of claim 7 wherein said cell is a CHO cell.

9. The host cell of claim 6 which is prokaryotic.

10. The host cell of claim 9 wherein said cell is an *E. coli*.

11. The host cell of claim 7 wherein said cell is a yeast cell.

12. The host cell of claim 11 which is *Saccharomyces cerevisiae*.

13. A process for producing *patched-2* polypeptides comprising culturing the host cell of claim 9 under conditions suitable for expression of vertebrate *patched-2* and recovering *patched-2* from the cell culture.

14. (Amended) Isolated native sequence human *patched-2* polypeptide comprising amino acid residues

1 to 1203 of Fig. 1 (SEQ ID NO:2).

15. (Amended) Isolated native sequence human *patched-2* polypeptide encoded by the nucleotide deposited under accession number ATCC 209778 (Designation: pRK7.hptc2.Flag-1405) having *patched-2* biological activity.

16. A chimeric molecule comprising vertebrate *patched-2* polypeptide patched-2 to a heterologous amino acid sequence.

17. The chimeric molecule of claim 16 wherein said heterologous amino acid sequence is an epitope tag sequence.

18. The chimeric molecule of claim 17 wherein said heterologous amino acid sequence is a constant region of an immunoglobulin.

19. An antagonist of *patched*-2 which blocks. prevents, inhibits and/or neutralizes the *Dhh* function in the *Dhh* signaling pathway.

20. The antagonist of claim 19 which is a small bioorganic molecule.

21. The antagonist of claim 19 which is an antisense nucleotide.

22. An agonist of *patched-2* with stimulates or enhances the normal functioning of *patched-2* in the *Dhh* signaling pathway.

23. (Amended) The agonist of claim 22 which prevents *Smo* (SEQ ID NO:17) inactivation of *ptch*-2 (SEQ ID NO:2)

24. The agonist of claim 22 which is a small bioorganic molecule.

25. The agonist of claim 24 which is a small bioorganic molecule.

26. A method of screening for antagonists or agonists of *patched-2* biological activity comprising:

(a) exposing *patched*-2 expressing target cells in culture to a candidate compound and *Dhh*; and

(b) analyzing cells for binding of *Dhh* to *patched*-2; or

(c) scoring phenotypic or functional changes in the treated cells:

and comparing the results to control cells which were not exposed to the candidate compound.

27. A method of screening for antagonist or agonist molecule of *patched-2* biological activity comprising:

(a) exposing a *patched-2* ligand and a compound having *patched-2* biological activity to a candidate antagonist or agonist; and

(b) analyzing the substrate for binding of the ligand to the compound; and

comparing the results to control reactions which were not exposed to the candidate molecule.

28. A method of diagnosing to determine whether a particular disorder is modulated by *Dhh* signaling, comprising:

(a) culturing test cells or tissues;

(b) administering a compound which can inhibit *patched-2* modulated *Dhh* signaling; and

(c) analyzing the level of *Dhh* binding to *patched*-2 or *Dhh* mediated phenotypic effects in the test cells.

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  1 GTTATTTCAG GCCATGGTGT TGCGCCGAAT TAATTCCCGA TCCAGACATG ATAAGATACA TTGATGAGTT TGGACAAACC ACAACTAGAA TGCAGTGAAA
    CAATAAAGTC CGGTACCACA ACGCGGCTTA ATTAAGGGCT AGGTCTGTAC TATTCTATGT AACTACTCAA ACCTGTTTGG TGTTGATCTT ACGTCACTTT
    (SEQ ID NO: 1)

101 AAAATGCTTT ATTTGTGAAA TTTGTGATGC TATTGCTTTA TTTGTAACCA TTATAAGCTG CAATAAACAA GTTGGGCCAT GGCGGCCAAG CTTCTGCAGG
    TTTTACGAAA TAAACACTTT AAACACTACG ATAACGAAAT AAACATTGGT AATATTCGAC GTTATTTGTT CAACCCGGTA CCGCCGGTTC GAAGACGTCC

201 TCGACTCTAG AGGATCCCCG GGGAATTCCG GCATGACTCG ATCGCGCCCC CTCAGAGAGC TGCCCCCGAG TTACACACCC CCAGCTCGAA CCGCAGCACC
    AGCTGAGATC TCCTAGGGGC CCCTTAAGGC CGTACTGAGC TAGCGCGGGG GAGTCTCTCG ACGGGGGCTC AATGTGTGGG GGTCGAGCTT GGCGTCGTGG
  1                                  M  T  R   S  P  P   L  R  E  L   P  P  S   Y  T  P   P  A  R  T   A  A  P
              ^insert starts here  (SEQ ID NO: 2)

301 CCAGATCCTA GCTGGGAGCC TGAAGGCTCC ACTCTGGCTT CGTGCTTACT TCCAGGGCCT GCTCTTCTCT CTGGGATGCG GGATCCAGAG ACATTGTGGC
    GGTCTAGGAT CGACCCTCGG ACTTCCGAGG TGAGACCGAA GCACGAATGA AGGTCCCGGA CGAGAAGAGA GACCCTACGC CCTAGGTCTC TGTAACACCG
 24  Q  I  L   A  G  S  L   K  A  P   L  W  L   R  A  Y  F   Q  G  L   L  F  S   L  G  C  G   I  Q  R   H  C  G

401 AAAGTGCTCT TTCTGGGACT GTTGGCCTTT GGGGCCCTGG CATTAGGTCT CCGCATGGCC ATTATTGAGA CAAACTTGGA ACAGCTCTGG GTAGAAGTGG
    TTTCACGAGA AAGACCCTGA CAACCGGAAA CCCCGGGACC GTAATCCAGA GGCGTACCGG TAATAACTCT GTTTGAACCT TGTCGAGACC CATCTTCACC
 57 K  V  L  F   L  G  L   L  A  F   G  A  L  A   L  G  L   R  M  A   I  I  E  T   N  L  E   Q  L  W   V  E  V  G

501 GCAGCCGGGT GAGCCAGGAG CTGCATTACA CCAAGGAGAA GCTGGGGGAG GAGGCTGCAT ACACCTCTCA GATGCTGATA CAGACCGCAC GCCAGGAGGG
    CGTCGGCCCA CTCGGTCCTC GACGTAATGT GGTTCCTCTT CGACCCCCTC CTCCGACGTA TGTGGAGAGT CTACGACTAT GTCTGGCGTG CGGTCCTCCC
 91  S  R  V   S  Q  E   L  H  Y  T   K  E  K   L  G  E   E  A  A  Y   T  S  Q   M  L  I   Q  T  A  R   Q  E  G

601 AGAGAACATC CTCACACCCG AAGCACTTGG CCTCCACCTC CAGGCAGCCC TCACTGCCAG TAAAGTCCAA GTATCACTCT ATGGGAAGTC CTGGGATTTG
    TCTCTTGTAG GAGTGTGGGC TTCGTGAACC GGAGGTGGAG GTCCGTCGGG AGTGACGGTC ATTTCAGGTT CATAGTGAGA TACCCTTCAG GACCCTAAAC
124  E  N  I   L  T  P  E   A  L  G   L  H  L   Q  A  A  L   T  A  S   K  V  Q   V  S  L  Y   G  K  S   W  D  L

701 AACAAAATCT GCTACAAGTC AGGAGTTCCC CTTATTGAAA ATGGAATGAT TGAGTGGATG ATTGAGAAGC TGTTTCCGTG CGTGATCCTC ACCCCCCTCG
    TTGTTTTAGA CGATGTTCAG TCCTCAAGGG GAATAACTTT TACCTTACTA ACTCACCTAC TAACTCTTCG ACAAAGGCAC GCACTAGGAG TGGGGGGAGC
157 N  K  I  C   Y  K  S   G  V  P   L  I  E  N   G  M  I   E  W  M   I  E  K  L   F  P  C   V  I  L   T  P  L  D

801 ACTGCTTCTG GGAGGGAGCC AAACTCCAAG GGGGCTCCGC CTACCTGCCC GGCCGCCCGG ATATCCAGTG GACCAACCTG GATCCAGAGC AGCTGCTGGA
    TGACGAAGAC CCTCCCTCGG TTTGAGGTTC CCCCGAGGCG GATGGACGGG CCGGCGGGCC TATAGGTCAC CTGGTTGGAC CTAGGTCTCG TCGACGACCT
191  C  F  W   E  G  A   K  L  Q  G   G  S  A   Y  L  P   G  R  P  D   I  Q  W   T  N  L   D  P  E  Q   L  L  E
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FIG. 1A

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 901 GGAGCTGGGT CCCTTTGCCT CCCTTGAGGG CTTCCGGGAG CTGCTAGACA AGGCACAGGT GGGCCAGGCC TACGTGGGGC GGCCCTGTCT GCACCCTGAT
     CCTCGACCCA GGGAAACGGA GGGAACTCCC GAAGGCCCTC GACGATCTGT TCCGTGTCCA CCCGGTCCGG ATGCACCCCG CCGGGACAGA CGTGGGACTA
 224  E  L  G   P  F  A  S   L  E  G   F  R  E   L  L  D  K   A  Q  V   G  Q  A   Y  V  G  R   P  C  L   H  P  D

1001 GACCTCCACT GCCCACCTAG TGCCCCCAAC CATCACAGCA GGCAGGCTCC CAATGTGGCT CACGAGCTGA GTGGGGGCTG CCATGGCTTC TCCCACAAAT
     CTGGAGGTGA CGGGTGGATC ACGGGGGTTG GTAGTGTCGT CCGTCCGAGG GTTACACCGA GTGCTCGACT CACCCCCGAC GGTACCGAAG AGGGTGTTTA
 257 D  L  H  C   P  P  S   A  P  N   H  H  S  R   Q  A  P   N  V  A   H  E  L  S   G  G  C   H  G  F   S  H  K  F

1101 TCATGCACTG GCAGGAGGAA TTGCTGCTGG GAGGCATGGC CAGAGACCCC CAAGGAGAGC TGCTGAGGGC AGAGGCCCTG CAGAGCACCT TCTTGCTGAT
     AGTACGTGAC CGTCCTCCTT AACGACGACC CTCCGTACCG GTCTCTGGGG GTTCCTCTCG ACGACTCCCG TCTCCGGGAC GTCTCGTGGA AGAACGACTA
 291   M  H  W   Q  E  E   L  L  L  G   G  M  A   R  D  P   Q  G  E  L   L  R  A   E  A  L   Q  S  T  F   L  L  M

1201 GAGTCCCCGC CAGCTGTACG AGCATTTCCG GGGTGACTAT CAGACACATG ACATTGGCTG GAGTGAGGAG CAGGCCAGCA CAGTGCTACA AGCCTGGCAG
     CTCAGGGGCG GTCGACATGC TCGTAAAGGC CCCACTGATA GTCTGTGTAC TGTAACCGAC CTCACTCCTC GTCCGGTCGT GTCACGATGT TCGGACCGTC
 324  S  P  R   Q  L  Y  E   H  F  R   G  D  Y   Q  T  H  D   I  G  W   S  E  E   Q  A  S  T   V  L  Q   A  W  Q

1301 CGGCGCTTTG TGCAGCTGGC CCAGGAGGCC CTGCCTGAGA ACGCTTCCCA GCAGATCCAT GCCTTCTCCT CCACCACCCT GGATGACATC CTGCATGCGT
     GCCGCGAAAC ACGTCGACCG GGTCCTCCGG GACGGACTCT TGCGAAGGGT CGTCTAGGTA CGGAAGAGGA GGTGGTGGGA CCTACTGTAG GACGTACGCA
 357 R  R  F  V   Q  L  A   Q  E  A   L  P  E  N   A  S  Q   Q  I  H   A  F  S  S   T  T  L   D  D  I   L  H  A  F

1401 TCTCTGAAGT CAGTGCTGCC CGTGTGGTGG GAGGCTATCT GCTCATGCTG GCCTATGCCT GTGTGACCAT GCTGCGGTGG GACTGCGCCC AGTCCCAGGG
     AGAGACTTCA GTCACGACGG GCACACCACC CTCCGATAGA CGAGTACGAC CGGATACGGA CACACTGGTA CGACGCCACC CTGACGCGGG TCAGGGTCCC
 391   S  E  V   S  A  A   R  V  V  G   G  Y  L   L  M  L   A  Y  A  C   V  T  M   L  R  W   D  C  A  Q   S  Q  G

1501 TTCCGTGGGC CTTGCCGGGG TACTGCTGGT GGCCCTGGCG GTGGCCTCAG GCCTTGGGCT CTGTGCCCTG CTCGGCATCA CCTTCAATGC TGCCACTACC
     AAGGCACCCG GAACGGCCCC ATGACGACCA CCGGGACCGC CACCGGAGTC CGGAACCCGA GACACGGGAC GAGCCGTAGT GGAAGTTACG ACGGTGATGG
 424  S  V  G   L  A  G  V   L  L  V   A  L  A   V  A  S  G   L  G  L   C  A  L   L  G  I  T   F  N  A   A  T  T

1601 CAGGTGCTGC CTTTCTTGGC TCTGGGAATC GGCGTGGATG ACGTATTCCT GCTGGCGCAT GCCTTCACAG AGGCTCTGCC TGGCACCCCT CTCCAGGAGC
     GTCCACGACG GAAAGAACCG AGACCCTTAG CCGCACCTAC TGCATAAGGA CGACCGCGTA CGGAAGTGTC TCCGAGACGG ACCGTGGGGA GAGGTCCTCG
 457 Q  V  L  P   F  L  A   L  G  I   G  V  D  D   V  F  L   L  A  H   A  F  T  E   A  L  P   G  T  P   L  Q  E  R

1701 GCATGGGCGA GTGTCTGCAG CGCACGGGCA CCAGTGTCGT ACTCACATCC ATCAACAACA TGGCCGCCTT CCTCATGGCT GCCCTCGTTC CCATCCCTGC
     CGTACCCGCT CACAGACGTC GCGTGCCCGT GGTCACAGCA TGAGTGTAGG TAGTTGTTGT ACCGGCGGAA GGAGTACCGA CGGGAGCAAG GGTAGGGACG
 491   M  G  E   C  L  Q   R  T  G  T   S  V  V   L  T  S   I  N  N  M   A  A  F   L  M  A   A  L  V  P   I  P  A
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FIG. 1B

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801 GCTGCGAGCC TTCTCCCTAC AGGCGGCCAT AGTGGTTGGC TGCACCTTTG TAGCCGTGAT GCTTGTCTTC CCAGCCATCC TCAGCCTGGA CCTACGGCGG
    CGACGCTCGG AAGAGGGATG TCCGCCGGTA TCACCAACCG ACGTGGAAAC ATCGGCACTA CGAACAGAAG GGTCGGTAGG AGTCGGACCT GGATGCCGCC
524  L  R  A   F  S  L  Q   A  A  I   V  V  G   C  T  F  V   A  V  M   L  V  F   P  A  I  L   S  L  D   L  R  R

901 CGCCACTGCC AGCGCCTTGA TGTGCTCTGC TGCTTCTCCA GTCCCTGCTC TGCTCAGGTG ATTCAGATCC TGCCCCAGGA GCTGGGGGAC GGGACAGTAC
    GCGGTGACGG TCGCGGAACT ACACGAGACG ACGAAGAGGT CAGGGACGAG ACGAGTCCAC TAAGTCTAGG ACGGGGTCCT CGACCCCCTG CCCTGTCATG
557 R  H  C  Q   R  L  D   V  L  C   C  F  S  S   P  C  S   A  Q  V   I  Q  I  L   P  Q  E   L  G  D   G  T  V  P

001 CAGTGGGCAT TGCCCACCTC ACTGCCACAG TTCAAGCCTT TACCCACTGT GAAGCCAGCA GCCAGCATGT GGTCACCATC CTGCCTCCCC AAGCCCACCT
    GTCACCCGTA ACGGGTGGAG TGACGGTGTC AAGTTCGGAA ATGGGTGACA CTTCGGTCGT CGGTCGTACA CCAGTGGTAG GACGGAGGGG TTCGGGTGGA
591  V  G  I   A  H  L   T  A  T  V   Q  A  F   T  H  C   E  A  S  S   Q  H  V   V  T  I   L  P  P  Q   A  H  L

101 GGTGCCCCCA CCTTCTGACC CACTGGGCTC TGAGCTCTTC AGCCCTGGAG GGTCCACACG GGACCTTCTA GGCCAGGAGG AGGAGACAAG GCAGAAGGCA
    CCACGGGGGT GGAAGACTGG GTGACCCGAG ACTCGAGAAG TCGGGACCTC CCAGGTGTGC CCTGGAAGAT CCGGTCCTCC TCCTCTGTTC CGTCTTCCGT
624 V  P  P   P  S  D  P   L  G  S   E  L  F   S  P  G  G   S  T  R   D  L  L   G  Q  E  E   E  T  R   Q  K  A

201 GCCTGCAAGT CCCTGCCCTG TGCCCGCTGG AATCTTGCCC ATTTCGCCCG CTATCAGTTT GCCCCGTTGC TGCTCCAGTC ACATGCCAAG GCCATCGTGC
    CGGACGTTCA GGGACGGGAC ACGGGCGACC TTAGAACGGG TAAAGCGGGC GATAGTCAAA CGGGGCAACG ACGAGGTCAG TGTACGGTTC CGGTAGCACG
557 A  C  K  S   L  P  C   A  R  W   N  L  A  H   F  A  R   Y  Q  F   A  P  L  L   L  Q  S   H  A  K   A  I  V  L

301 TGGTGCTCTT TGGTGCTCTT CTGGGCCTGA GCCTCTACGG AGCCACCTTG GTGCAAGACG GCCTGGCCCT GACGGATGTG GTGCCTCGGG GCACCAAGGA
    ACCACGAGAA ACCACGAGAA GACCCGGACT CGGAGATGCC TCGGTGGAAC CACGTTCTGC CGGACCGGGA CTGCCTACAC CACGGAGCCC CGTGGTTCCT
91   V  L  F   G  A  L   L  G  L  S   L  Y  G   A  T  L   V  Q  D  G   L  A  L   T  D  V   V  P  R  G   T  K  E

401 GCATGCCTTC CTGAGCGCCC AGCTCAGGTA CTTCTCCCTG TACGAGGTGG CCCTGGTGAC CCAGGGTGGC TTTGACTACG CCCATTCCCA ACGCGCCCTC
    CGTACGGAAG GACTCGCGGG TCGAGTCCAT GAAGAGGGAC ATGCTCCACC GGGACCACTG GGTCCCACCG AAACTGATGC GGGTAAGGGT TGCGCGGGAG
24   H  A  F   L  S  A  Q   L  R  Y   F  S  L   Y  E  V  A   L  V  T   Q  G  G   F  D  Y  A   H  S  Q   R  A  L

01 TTTGATCTGC ACCAGCGCTT CAGTTCCCTC AAGGCGGTGC TGCCCCCACC GGCCACCCAG GCACCCCGCA CCTGGCTGCA CTATTACCGC AACTGGCTAC
   AAACTAGACG TGGTCGCGAA GTCAAGGGAG TTCCGCCACG ACGGGGGTGG CCGGTGGGTC CGTGGGGCGT GGACCGACGT GATAATGGCG TTGACCGATG
57 F  D  L  H   Q  R  F   S  S  L   K  A  V  L   P  P  P   A  T  Q   A  P  R  T   W  L  H   Y  Y  R   N  W  L  Q

01 AGGGAATCCA GGCTGCCTTT GACCAGGACT GGGCTTCTGG GCGCATCACC CGCCACTCGT ACCGCAATGG CTCTGAGGAT GGGGCCCTGG CCTACAAGCT
   TCCCTTAGGT CCGACGGAAA CTGGTCCTGA CCCGAAGACC CGCGTAGTGG GCGGTGAGCA TGGCGTTACC GAGACTCCTA CCCCGGGACC GGATGTTCGA
91  G  I  Q   A  A  F   D  Q  D  W   A  S  G   R  I  T   R  H  S  Y   R  N  G   S  E  D   G  A  L  A   Y  K  L
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FIG. 1C

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2701 GCTCATCCAG ACTGGAGACG CCCAGGAGCC TCTGGATTTC AGCCAGCTGA CCACAAGGAA GCTGGTGGAC AGAGAGGGAC TGATTCCACC CGAGCTCTTC
     CGAGTAGGTC TGACCTCTGC GGGTCCTCGG AGACCTAAAG TCGGTCGACT GGTGTTCCTT CGACCACCTG TCTCTCCCTG ACTAAGGTGG GCTCGAGAAG
 824 L I Q  T G D A  Q E P  L D F  S Q L T  T R K  L V D  R E G L  I P P  E L F

2801 TACATGGGGC TGACCGTGTG GGTGAGCAGT GACCCCCTGG GTCTGGCAGC CTCACAGGCC AACTTCTACC CCCCACCTCC TGAATGGCTG CACGACAAAT
     ATGTACCCCG ACTGGCACAC CCACTCGTCA CTGGGGGACC CAGACCGTCG GAGTGTCCGG TTGAAGATGG GGGGTGGAGG ACTTACCGAC GTGCTGTTTA
 857 Y M G L  T V W  V S S  D P L G  L A A  S Q A  N F Y P  P P P  E W L  H D K Y

2901 ACGACACCAC GGGGGAGAAC CTTCGCATCC CGCCAGCTCA GCCCTTGGAG TTTGCCCAGT TCCCCTTCCT GCTGCGTGGC CTCCAGAAGA CTGCAGACTT
     TGCTGTGGTG CCCCCTCTTG GAAGCGTAGG GCGGTCGAGT CGGGAACCTC AAACGGGTCA AGGGGAAGGA CGACGCACCG GAGGTCTTCT GACGTCTGAA
 891  D T T  G E N  L R I P  P A Q  P L E  F A Q F  P F L  L R G  L Q K T  A D F

3001 TGTGGAGGCC ATCGAGGGGG CCCGGGCAGC ATGCGCAGAG GCCGGCCAGG CTGGGGTGCA CGCCTACCCC AGCGGCTCCC CCTTCCTCTT CTGGGAACAG
     ACACCTCCGG TAGCTCCCCC GGGCCCGTCG TACGCGTCTC CGGCCGGTCC GACCCCACGT GCGGATGGGG TCGCCGAGGG GGAAGGAGAA GACCCTTGTC
 924  V E A  I E G A  R A A  C A E  A G Q A  G V H  A Y P  S G S P  F L F  W E Q

3101 TATCTGGGCC TGCGGCGCTG CTTCCTGCTG GCCGTCTGCA TCCTGCTGGT GTGCACTTTC CTCGTCTGTG CTCTGCTGCT CCTCAACCCC TGGACGGCTG
     ATAGACCCGG ACGCCGCGAC GAAGGACGAC CGGCAGACGT AGGACGACCA CACGTGAAAG GAGCAGACAC GAGACGACGA GGAGTTGGGG ACCTGCCGAC
 957 Y L G L  R R C  F L L  A V C I  L L V  C T F  L V C A  L L L  L N P  W T A G

3201 GCCTCATAGT GCTGGTCCTG GCGATGATGA CAGTGGAACT CTTTGGTATC ATGGGTTTCC TGGGCATCAA GCTGAGTGCC ATCCCCGTGG TGATCCTTGT
     CGGAGTATCA CGACCAGGAC CGCTACTACT GTCACCTTGA GAAACCATAG TACCCAAAGG ACCCGTAGTT CGACTCACGG TAGGGGCACC ACTAGGAACA
 991  L I V  L V L  A M M T  V E L  F G I  M G F L  G I K  L S A  I P V V  I L V

3301 GGCCTCTGTA GGCATTGGCG TTGAGTTCAC AGTCCACGTG GCTCTGGGCT TCCTGACCAC CCAGGGCAGC CGGAACCTGC GGGCCGCCCA TGCCCTTGAG
     CCGGAGACAT CCGTAACCGC AACTCAAGTG TCAGGTGCAC CGAGACCCGA AGGACTGGTG GGTCCCGTCG GCCTTGGACG CCCGGCGGGT ACGGGAACTC
1024  A S V  G I G V  E F T  V H V  A L G F  L T T  Q G S  R N L R  A A H  A L E

3401 CACACATTTG CCCCCGTGAC CGATGGGGCC ATCTCCACAT TGCTGGGTCT GCTCATGCTT GCTGGTTCCC ACTTTGACTT CATTGTAAGG TACTTCTTTG
     GTGTGTAAAC GGGGGCACTG GCTACCCCGG TAGAGGTGTA ACGACCCAGA CGAGTACGAA CGACCAAGGG TGAAACTGAA GTAACATTCC ATGAAGAAAC
1057 H T F A  P V T  D G A  I S T L  L G L  L M L  A G S H  F D F  I V R  Y F F A

3501 CGGCGCTGAC AGTGCTCACG CTCCTGGGCC TCCTCCATGG ACTCGTGCTG CTGCCTGTGC TGCTGTCCAT CCTGGGCCCG CCGCCAGAGG TGATACAGAT
     GCCGCGACTG TCACGAGTGC GAGGACCCGG AGGAGGTACC TGAGCACGAC GACGGACACG ACGACAGGTA GGACCCGGGC GGCGGTCTCC ACTATGTCTA
1091  A L T  V L T  L L G L  L H G  L V L  L P V L  L S I  L G P  P P E V  I Q M
```

FIG. 1D

```
601 GTACAAGGAA AGCCCAGAGA TCCTGAGTCC ACCAGCTCCA CAGGGAGGCG GGCTTAGGTG GGGGGCATCC TCCTCCCTGC CCCAGAGCTT TGCCAGAGTG
    CATGTTCCTT TCGGGTCTCT AGGACTCAGG TGGTCGAGGT GTCCCTCCGC CCGAATCCAC CCCCCGTAGG AGGAGGGACG GGGTCTCGAA ACGGTCTCAC
124  Y  K  E   S  P  E  I   L  S  P   P  A  P   Q  G  G  G   L  R  W   G  A  S   S  S  L  P   Q  S  F   A  R  V

701 ACTACCTCCA TGACCGTGGC CATCCACCCA CCCCCCCTGC CTGGTGCCTA CATCCATCCA GCCCCTGATG AGCCCCCTTG GTCCCCTGCT GCCACTAGCT
    TGATGGAGGT ACTGGCACCG GTAGGTGGGT GGGGGGGACG GACCACGGAT GTAGGTAGGT CGGGGACTAC TCGGGGGAAC CAGGGGACGA CGGTGATCGA
157 T  T  S  M   T  V  A   I  H  P   P  P  L  P   G  A  Y   I  H  P   A  P  D  E   P  P  W   S  P  A   A  T  S  S

801 CTGGCAACCT CAGTTCCAGG GGACCAGGTC CAGCCACTGG GTGAAAGAGC AGCTGAAGCA CAGAGACCAT GTGTGGGGCG TGTGGGGTCA CTGGGAAGCA
    GACCGTTGGA GTCAAGGTCC CCTGGTCCAG GTCGGTGACC CACTTTCTCG TCGACTTCGT GTCTCTGGTA CACACCCCGC ACACCCCAGT GACCCTTCGT
191  G  N  L   S  S  R   G  P  G  P   A  T  G  O
                                               1203

901 CTGGGTCTGG TGTTAGACGC AGGACGGACC CCTGGAGGGC CCTGCTGCTG CTGCATCCCC TCTCCCGACC CAGCTGTCAT GGGCCTCCCT GATATCGAAT
    GACCCAGACC ACAATCTGCG TCCTGCCTGG GGACCTCCCG GGACGACGAC GACGTAGGGG AGAGGGCTGG GTCGACAGTA CCCGGAGGGA CTATAGCTTA
                            ^T to C  (silent)                 pRK follows, this is the 5prime end of vector^

001 TCAATCGATA GAACCGAGGT GCAGTTGGAC
    AGTTAGCTAT CTTGGCTCCA CGTCAACCTG
```

FIG. 1E

```
(SEQ ID NO: 3)            30        40        50        60        70
905531        GCTGGGGTGCACGCCTACCNCAGCGGNTCCCCCTTCCTCTTCTGGGAACA
                                  ******* ***** ****************** **
                 ::: :: :
hpatched      CTGGGGCTGTCCAGTTACCCCAACGGCTACCCCTTCCTCTTCTGGGAGCA
(SEQ ID NO: 4) 3010      3020      3030      3040      3050

                          80        90       100       110       120
905531        GTATCTGGGCCTGCGGCGCTGCTTCCTGCTGGCCGTCTGCATCCTGCTGG
              ***  * ***** ** * ***  * ******  * ** ** *  ** ***
hpatched      GTACATCGGCCTCCGCCACTGGCTGCTGCTGTTCATCAGCGTGGTGTTGG
           3060      3070      3080      3090      3100

                         130       140       150       160       170
905531        TGTGCACTTTCCTCGTCTGTGCTCTGCTGCTCCTNAACCCCTGGACGGCT
                ***** ******** ** *** *  * ** *****************
hpatched      CCTGCACATTCCTCGTGTGCGCTGTCTTCCTTCTGAACCCCTGGACGGCC
           3110      3120      3130      3140      3150

                         180       190       200       210       220
905531        GGCCTNATAGTGCTGGTCCTGGCGATGATGACAGTGGAACTCTTTGGTAT
              **  **** *** *********** ******* ** ** ** ** ** **
hpatched      GGGATCATTGTGATGGTCCTGGCGCTGATGACGGTCGAGCTGTTCGGCAT
           3160      3170      3180      3190      3200

                         230       240       250
905531        CATGGGTTTNCTGGGCATCAAGCTGAGT
                *****  ** * ** ******** ***
hpatched      GATGGGCCTCATCGGAATCAAGCTCAGT
           3210      3220      3230


                      80        90       100       110       120
905531        TCTGGGCCTGCGGCGCTGCTTCCTGCTGGCCGTCTGCATCCTGCTGGTGT
               :::   :::       * ** *  ** *****     ****  * ** ****
hpatched      GCTGCTGCTGTTCATCAGCGTGGTGTTGGCC---TGCACATTCCTCGTGT
                 3090      3100      3110           3120

                     130       140       150
905531        GCACTTTCCTCGTCTGTGCTCTGCTGCT
              ** ** ** ** *          :    :
hpatched      GCGCTGTCTTCCTTCTGAACCCCTGGAC
             3130      3140      3150
```

FIG. 2A

```
                          30        40        50        60        70
(SEQ ID NO: 5) 1326258    GCTGGGGTGCACGCCTACCCCAGCGGCTCCCCCTTCCTCTTCTGGAACA
                             ::: ::  :   ******* ***** ***************** **
               hpatched   CTGGGGCTGTCCAGTTACCCCAACGGCTACCCCTTCCTCTTCTGGAGCA
                       3010      3020      3030      3040      3050

                          80        90        100       110       120
               1326258    GTATCTGGGCCTGCGGCGCTGCTTCCTGCTGGCCGTCTGCATCCTGCTGG
                          ***  * ***** ** * ***  * ******  * ** ** *  ** ***
               hpatched   GTACATCGGCCTCCGCCACTGGCTGCTGCTGTTCATCAGCGTGGTGTTGG
                       3060      3070      3080      3090      3100

                         130       140       150
               1326258    TGTGCACTTTCCTCNTCTGTGCTCT
                            ***** ******** ** *** *
               hpatched   CCTGCACATTCCTCGTGTGCGCTGT
                       3110      3120      3130
```

```
                                90        100       110       120       130
               1326258    TCTGGGCCTGCGGCGCTGCTTCCTGCTGGCCGTCTGCATCCTGCTGGTGT
                           :::   :::     * ** *  ** *****   ****   * ** ****
               hpatched   GCTGCTGCTGTTCATCAGCGTGGTGTTGGCC---TGCACATTCCTCGTGT
                                3090      3100      3110         3120

                                140       150
               1326258    GCACTTTCCTCNTCTGTGCTCT
                          ** ** ** ****       :
               hpatched   GCGCTGTCTTCCTTCTGAACCC
                         3130      3140
```

```
                                   10        20        30        40        50
               1326258       CCGGGCAGCATGCGCAGAGGCCGGCCAGGCTGGGGTGCACGCCTACCCCA
                             ****** *****    ** ** * *** ******* *     :
(SEQ ID NO: 6) hpatched.RC   CCGGGCGGCATG--GCGAAGCGGACCACGCTGGGGGTGGCTCAGGGGAG
                            710       720         730       740       750
```

FIG. 2B

(SEQ ID NO:4) PTCH
(SEQ ID NO:2) PTCH2

PTCH	1	MASAGNAAEPQDRGGGGSGCIGAPGRPAGGGRRRRTGGLRRAAAPDRDYL
PTCH2	1	---------------------------------------MTRSPPLREL-
PTCH	51	HRPSYCDAAFALEQISKGKATGRKAPLWLRAKFQRLLFKLGCYIQKNCGK
PTCH2	11	-PPSYTPP--ARTAAPQILAGSLKAPLWLRAYFQGLLFSLGCGIQRHCGK

TM1

PTCH	101	FLVVGLLIFGAFAVGLKAANLETNVEELWVEVGGRVSRELNYTRQKIGEE
PTCH2	58	VLFLGLLAFGALALGLRMAIIETNLEQLWVEVGSRVSQELHYTKEKLGEE
PTCH	151	AMFNPQLMIQTPKEEGANVLTTEALLQHLDSALQASRVHVYMYNRQWKLE
PTCH2	108	AAYTSQMLIQTARQEGENILTPEALGLHLQAALTASKVQVSLYGKSWDLN
PTCH	201	HLCYKSGELITETGYMDQIIEYLYPCLIITPLDCFWEGAKLQSGTAYLLG
PTCH2	158	KICYKSGVPLIENGMIEWMIEKLFPCVILTPLDCFWEGAKLQGGSAYLPG
PTCH	251	KPPLRWTNFDPLEFLEELKKINYQVDSWEEMLNKAEVGHGYMDRPCLNPA
PTCH2	208	RPDIQWTNLDPEQLLEELGPFA-SLEGFRELLDKAQVGQAYVGRPCLHPD
PTCH	301	DPDCPATAPNKNSTKPLDMALVLNGGCHGLSRKYMHWQEELIVGGTVKNS
PTCH2	257	DLHCPPSAPNHHSRQAPNVAHELSGGCHGFSHKFMHWQEELLLGGMARDP
PTCH	351	TGKLVSAHALQTMFQLMTPKQMYEHFKGY-EYVSH-INWNEDKAAAILEAW
PTCH2	307	QGELLRAEALQSTFLLMSPRQLYEHFRG-DYQTHDIGWSEEQASTVLQAW

TM2

PTCH	400	QRTYVEVVHQSVAQNSTQKVLSFTTTTLDDILKSFSDVSVIRVASGYLLM
PTCH2	356	QRRFVQLAQEALPENASQQIHAFSSTTLDDILHAFSEVSAARVVGGYLLM

FIG. 3A

```
                                                              TM3
PTCH   450  LAYACLTMLRWDCSKSQGAVGLAGVLLVALSVAAGLGLCSLIGISFNAAT
PTCH2  406  LAYACVTMLRWDCAQSQGSVGLAGVLLVALAVASGLGLCALLGITFNAAT

                      TM4
PTCH   500  TQVLPFLALGVGVDDVFLLAHAFSETGQNKRIPFEDRTGECLKRTGASVA
PTCH2  456  TQVLPFLALGIGVDDVFLLAHAFTEALPG--TPLQERMGECLQRTGTSVV

                      TM5                                TM6
PTCH   550  LTSISNVTAFFMAALIPIPALRAFSLQAAVVVVFNFAMVLLIFPAILSMD
PTCH2  504  LTSINNMAAFLMAALVPIPALRAFSLQAAIVVGCTFVAVMLVFPAILSLD

PTCH   600  LYRREDRRLDIFCCFTSPCVSRVIQVEPQAYTDTHDNTRYSPPPPYSSHS
PTCH2  554  LRRRHCQRLDVLCCFSSPCSAQVIQILPQELGDGT------------VPVG

PTCH   650  FAHETQITMQSTVQLRTEYDPHTHVYYTTAEPRSEISVQPVTVTQDTLSC
PTCH2  593  IAH-----LTATVQAFTHCEASSQHVVTILPPQAHLVPPP----SDPLGS

PTCH   700  QSPESTSSTRDLLSQFSDSSLH--CLEPPCTKWTLSSFAEKHYAPFLLKP
PTCH2  634  ELFSPGGSTRDLLGQEEETRQKAACKSLPCARWNLAHFARYQFAPLLLQS

                        TM7
PTCH   748  KAKVVVIFLFLGLLGVSLYGTTRVRDGLDLTDIVPRETREYDFIAAQFKY
PTCH2  684  HAKAIVLVLFGALLGLSLYGATLVQDGLALTDVVPRGTKEHAFLSAQLRY

PTCH   798  FSFYNMYIVTQKA-DYPNIQHLLYDLHRSFSNVKYVMLEENKQLPKMWLH
PTCH2  734  FSLYEVALVTQGGFDYAHSQRALFDLHQRFSSLKAVLPPPATQAPRTWLH
```

FIG. 3B

```
                                         ***
PTCH   847  YFRDWLQGLQDAFDSDWETGKIMPNNYKNGSDDGVLAYKLLVQTGSRDKP
PTCH2  784  YYRNWLQGIQAAFDQDWASGRITRHSYRNGSEDGALAYKLLIQTGDAQEP

PTCH   897  IDISQLTKQRLVDADGIINPSAFYIYLTAWVSNDPVAYAASQANIRPHRP
PTCH2  834  LDFSQLTTRKLVDREGLIPPELFYMGLTVWVSSDPLGLAASQANFYPPPP

PTCH   947  EWVHDKADYMPETRLRIPAAEPIEYAQFPFYLNGLRDTSDFVEAIEKVRT
PTCH2  884  EWLHDKYD-TTGENLRIPPAQPLEFAQFPFLLRGLQKTADFVEAIEGARA

                                                   TM8
PTCH   997  ICSNYTSLGLSSYPNGYPFLFWEQYIGLRHWLLLFISVVLACTFLVCAVF
PTCH2  933  ACAEAGQAGVHAYPSGSPFLFWEQYLGLRRCFLLAVCILLVCTFLVCALL

                      TM9                           TM10
PTCH  1047  LLNPWTAGIIVMVLALMTVELFGMMGLIGIKLSAVPVVILIASVGIGVEF
PTCH2  983  LLNPWTAGLIVLVLAMMTVELFGIMGFLGIKLSAIPVVILVASVGIGVEF

                                                TM11
PTCH  1097  TVHVALAFLTAIGDKNRRAVLALEHMFAPVLDGAVSTLLGVLMLAGSEFD
PTCH2 1033  TVHVALGFLTTQGSRNLRAAHALEHTFAPVTDGAISTLLGLLMLAGSHFD

                               TM12
PTCH  1147  FIVRYFFAVLAILTILGVLNGLVLLPVLLSFFGPYPEVSPANGLNRLPTP
PTCH2 1083  FIVRYFFAALTVLTLLGLLHGLVLLPVLLSILGPPPEVIQMYKESPEILS

PTCH  1197  SPEPPPSVVRFAMPPGHTHSGSDSSDSEYSSQTTVSGLSEELRHYEAQQG
PTCH2 1133  PPAPQGGGLRWGASSSLPQS-FARVTTSMTVAIHPPPLPGAYIHPAPDEP
```

FIG. 3C

```
PTCH   1247 A G G P A H Q V I V E A T E N P V F A H S T V V H P E S R H H P P S N P R Q Q P H L D S G S L P P G
PTCH2  1182 P W S P A A T S S G N L S S R G P G P A T G

PTCH   1297 R Q G Q Q P R R D P P R E G L W P P L Y R P R R D A F E I S T E G H S G P S N R A R W G P R G A R S

PTCH   1347 H N P R N P A S T A M G S S V P G Y C Q P I T T V T A S A S V T V A V H P P P V P G P G R N P R G G

PTCH   1397 L C P G Y P E T D H G L F E D P H V P F H V R C E R R D S K V E V I E L Q D V E C E E R P R G S S S

PTCH   1447 N
```

FIG. 3D

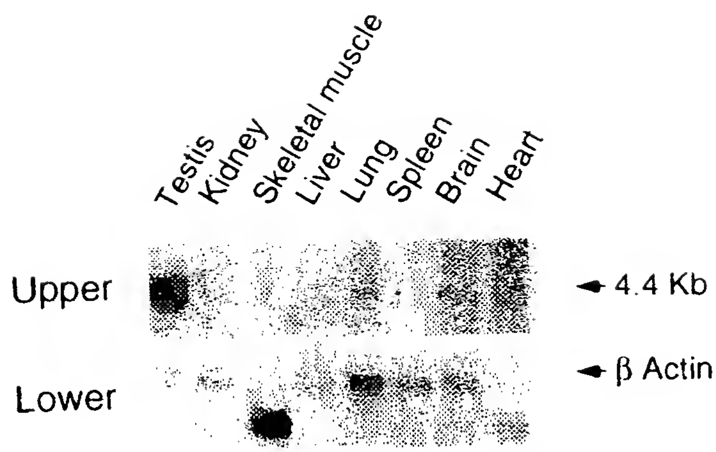

Testis
Kidney
Skeletal muscle
Liver
Lung
Spleen
Brain
Heart
Upper
Lower
4.4 Kb
β Actin

FIG. 4

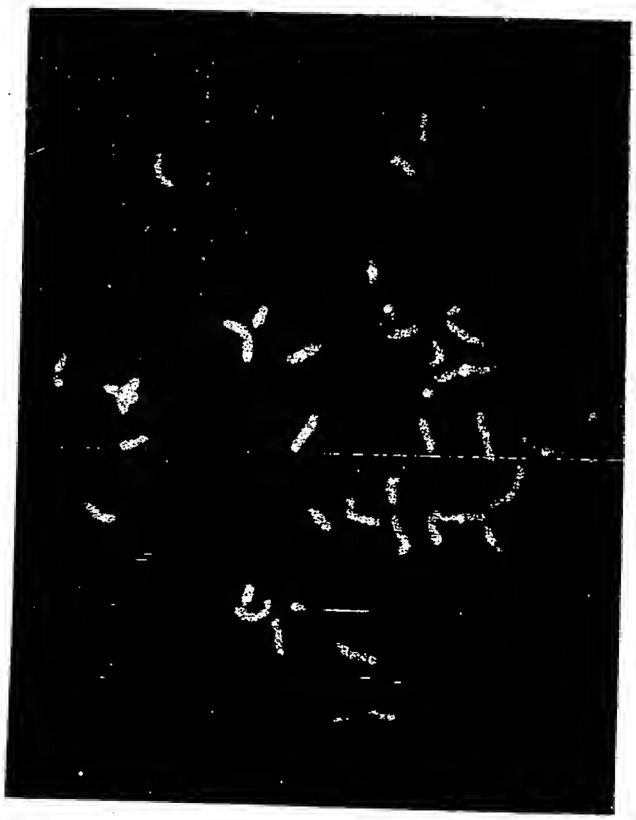


FIG. 5

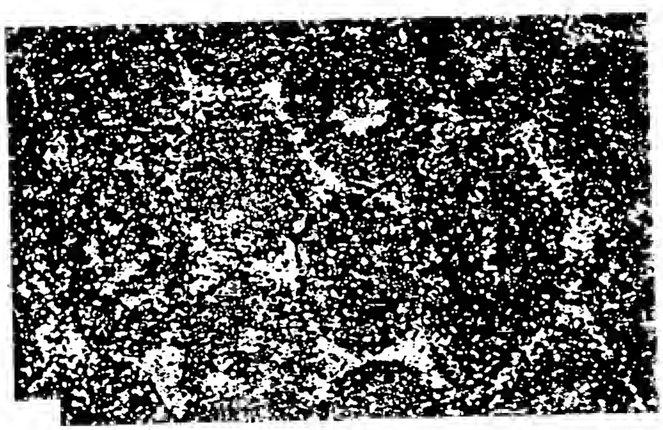


FIG. 6A

FIG. 6B

FIG. 6C

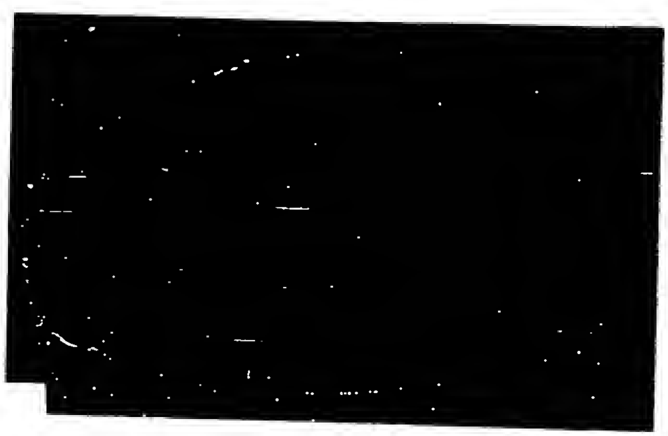


FIG. 6D

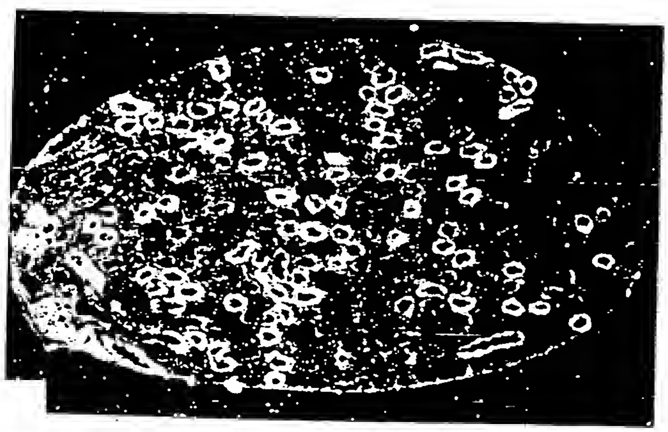


FIG. 6E

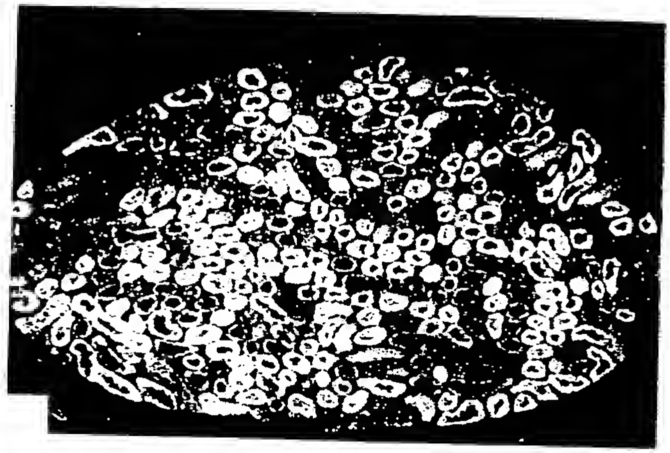
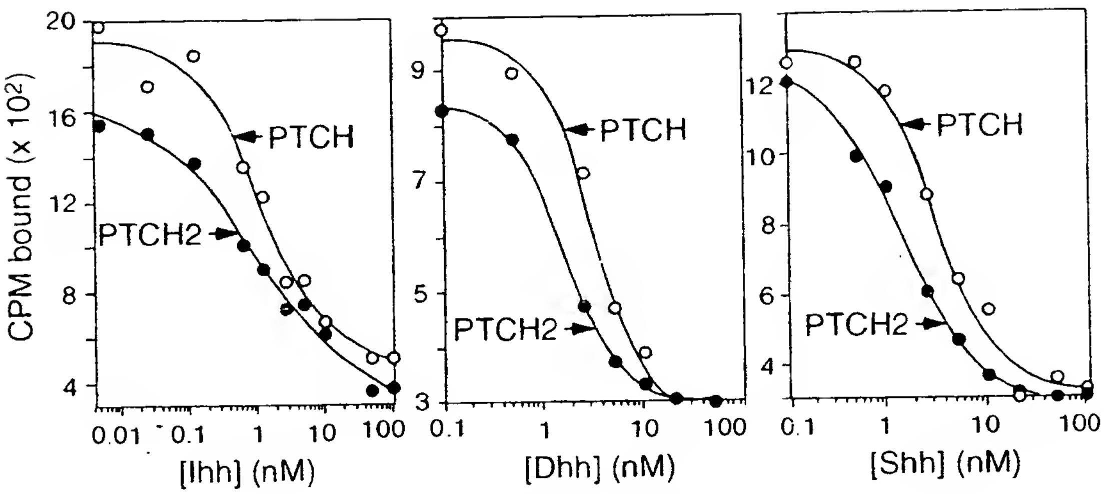
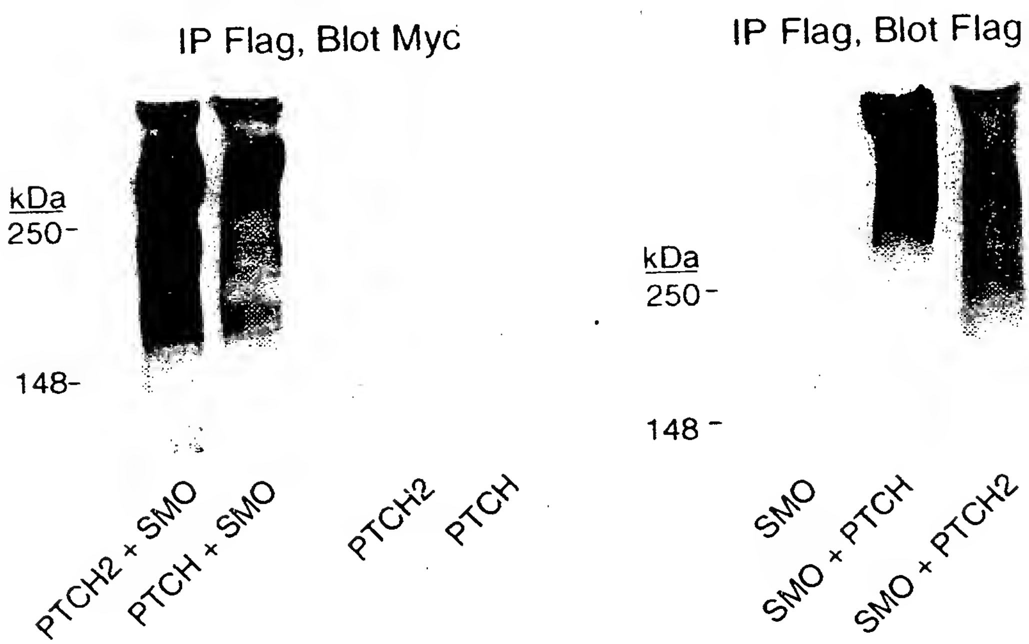


FIG. 6F



CPM bound (x 10²)
20
16
12
8
4
PTCH
PTCH2
0.01
0.1
1
10
100
[Ihh] (nM)
9
7
5
3
0.1
1
10
100
[Dhh] (nM)
12
10
8
6
4
0.1
1
10
100
[Shh] (nM)

FIG. 7A



IP Flag, Blot Myc
kDa
250-
148-
PTCH2 + SMO
PTCH + SMO
PTCH2
PTCH
IP Flag, Blot Flag
kDa
250-
148-
SMO
SMO + PTCH
SMO + PTCH2

FIG. 7B

```
                    10        20        30        40        50
hPtch-2     MTRSPPLRELPPSYTPPARTAAPQILAGSLKAPLWLRAYFQGLLFSLGCG
            * *. .* ***********..**.******.******************
mPatched2   MVRPLSLGELPPSYTPPARSSAPHILAGSLQAPLWLRAYFQGLLFSLGCR
(SEQ ID NO:7)       10        20        30        40        50

                    60        70        80        90       100
hPtch-2     IQRHCGKVLFLGLLAFGALALGLRMAIIETNLEQLWVEVGSRVSQELHYT
            **.**********.**********.*.***.*******************
mPatched2   IQKHCGKVLFLGLVAFGALALGLRVAVIETDLEQLWVEVGSRVSQELHYT
                    60        70        80        90       100

                   110       120       130       140       150
hPtch-2     KEKLGEEAAYTSQMLIQTARQEGENILTPEALGLHLQAALTASKVQVSLY
            ********************.*** *.******.****************
mPatched2   KEKLGEEAAYTSQMLIQTAHQEGGNVLTPEALDLHLQAALTASKVQVSLY
                   110       120       130       140       150

                   160       170       180       190       200
hPtch-2     GKSWDLNKICYKSGVPLIENGMIEWMIEKLFPCVILTPLDCFWEGAKLQG
            *************************.************************
mPatched2   GKSWDLNKICYKSGVPLIENGMIERMIEKLFPCVILTPLDCFWEGAKLQG
                   160       170       180       190       200

                   210       220       230       240       250
hPtch-2     GSAYLPGRPDIQWTNLDPEQLLEELGPFASLEGFRELLDKAQVGQAYVGR
            *****************.********************************
mPatched2   GSAYLPGRPDIQWTNLDPQQLLEELGPFASLEGFRELLDKAQVGQAYVGR
                   210       220       230       240       250

                   260       270       280       290       300
hPtch-2     PCLHPDDLHCPPSAPNHHSRQAPNVAHELSGGCHGFSHKFMHWQEELLLG
            ***.*** ********.*********.***********************
mPatched2   PCLDPDDPHCPPSAPNRHSRQAPNVAQELSGGCHGFSHKFMHWQEELLLG
                   260       270       280       290       300

                   310       320       330       340       350
hPtch-2     GMARDPQGELLRAEALQSTFLLMSPRQLYEHFRGDYQTHDIGWSEEQAST
            * *** **.****************************************
mPatched2   GTARDLQGQLLRAEALQSTFLLMSPRQLYEHFRGDYQTHDIGWSEEQASM
                   310       320       330       340       350
```

FIG. 8A

```
                   360       370       380       390       400
hPtch-2     VLQAWQRRFVQLAQEALPENASQQIHAFSSTTLDDILHAFSEVSAARVVG
            ****************** *******************.******..***
mPatched2   VLQAWQRRFVQLAQEALPANASQQIHAFSSTTLDDILRAFSEVSTTRVVG
                   360       370       380       390       400

                   410       420       430       440       450
hPtch-2     GYLLMLAYACVTMLRWDCAQSQGSVGLAGVLLVALAVASGLGLCALLGIT
            ***********************.**************************
mPatched2   GYLLMLAYACVTMLRWDCAQSQGAVGLAGVLLVALAVASGLGLCALLGIT
                   410       420       430       440       450

                   460       470       480       490       500
hPtch-2     FNAATTQVLPFLALGIGVDDVFLLAHAFTEALPGTPLQERMGECLQRTGT
            ********************.******** * *.*** *******. ***
mPatched2   FNAATTQVLPFLALGIGVDDIFLLAHAFTKAPPDTPLPERMGECLRSTGT
                   460       470       480       490       500

                   510       520       530       540       550
hPtch-2     SVVLTSINNMAAFLMAALVPIPALRAFSLQAAIVVGCTFVAVMLVFPAIL
            ** ***.*** **.*********************** * **********
mPatched2   SVALTSVNNMVAFFMAALVPIPALRAFSLQAAIVVGCNFAAVMLVFPAIL
                   510       520       530       540       550

                   560       570       580       590       600
hPtch-2     SLDLRRRHCQRLDVLCCFSSPCSAQVIQILPQELGDGTVPVGIAHLTATV
            ******** *******************.******* .************
mPatched2   SLDLRRRHRQRLDVLCCFSSPCSAQVIQMLPQELGDRAVPVGIAHLTATV
                   560       570       580       590       600

                   610       620       630       640       650
hPtch-2     QAFTHCEASSQHVVTILPPQAHLVPPPSDPLGSELFSPGGSTRDLLGQEE
            ***********************..*.********.**********.***
mPatched2   QAFTHCEASSQHVVTILPPQAHLLSPASDPLGSELYSPGGSTRDLLSQEE
                   610       620       630       640       650

                   660       670       680       690       700
hPtch-2     ETRQKAACKSLPCARWNLAHFARYQFAPLLLQSHAKAIVLVLFGALLGLS
             *  .***..* **.* ***************..***.**..********
mPatched2   GTGPQAACRPLLCAHWTLAHFARYQFAPLLLQTRAKALVLLFFGALLGLS
                   660       670       680       690       700
```

FIG. 8B

```
                   710       720       730       740       750
hPtch-2     LYGATLVQDGLALTDVVPRGTKEHAFLSAQLRYFSLYEVALVTQGGFDYA
            **************************************************
mPatched2   LYGATLVQDGLALTDVVPRGTKEHAFLSAQLRYFSLYEVALVTQGGFDYA
                   710       720       730       740       750

                   760       770       780       790       800
hPtch-2     HSQRALFDLHQRFSSLKAVLPPPATQAPRTWLHYYRNWLQGIQAAFDQDW
            ************************************.*************
mPatched2   HSQRALFDLHQRFSSLKAVLPPPATQAPRTWLHYYRSWLQGIQAAFDQDW
                   760       770       780       790       800

                   810       820       830       840       850
hPtch-2     ASGRITRHSYRNGSEDGALAYKLLIQTGDAQEPLDFSQLTTRKLVDREGL
            ****** *********************.*****************.***
mPatched2   ASGRITCHSYRNGSEDGALAYKLLIQTGNAQEPLDFSQLTTRKLVDKEGL
                   810       820       830       840       850

                   860       870       880       890       900
hPtch-2     IPPELFYMGLTVWVSSDPLGLAASQANFYPPPPEWLHDKYDTTGENLRIP
            **************************************************
mPatched2   IPPELFYMGLTVWVSSDPLGLAASQANFYPPPPEWLHDKYDTTGENLRIP
                   860       870       880       890       900

                   910       920       930       940       950
hPtch-2     PAQPLEFAQFPFLLRGLQKTADFVEAIEGARAACAEAGQAGVHAYPSGSP
            .*************.*******************.***************
mPatched2   AAQPLEFAQFPFLLHGLQKTADFVEAIEGARAACTEAGQAGVHAYPSGSP
                   910       920       930       940       950

                   960       970       980       990      1000
hPtch-2     FLFWEQYLGLRRCFLLAVCILLVCTFLVCALLLLNPWTAGLIVLVLAMMT
            **********************************.***************
mPatched2   FLFWEQYLGLRRCFLLAVCILLVCTFLVCALLLLSPWTAGLIVLVLAMMT
                   960       970       980       990      1000

                  1010      1020      1030      1040      1050
hPtch-2     VELFGIMGFLGIKLSAIPVVILVASVGIGVEFTVHVALGFLTTQGSRNLR
            *************************.****************..******
mPatched2   VELFGIMGFLGIKLSAIPVVILVASIGIGVEFTVHVALGFLTSHGSRNLR
                  1010      1020      1030      1040      1050
```

FIG. 8C

```
                        1060      1070      1080      1090      1100
hPtch-2      AAHALEHTFAPVTDGAISTLLGLLMLAGSHFDFIVRYFFAALTVLTLLGL
             ** ***.*********.************.****.****  *********
mPatched2    AASALEQTFAPVTDGAVSTLLGLLMLAGSNFDFIIRYFFVVLTVLTLLGL
                        1060      1070      1080      1090      1100

                        1110      1120      1130      1140      1150
hPtch-2      LHGLVLLPVLLSILGPPPEVIQMYKESPEILSPPAPQGGGLRWGASSSLP
             ****.*************.*.*.*****. *...*** *****. ...**
mPatched2    LHGLLLLPVLLSILGPPPQVVQVYKESPQTLNSAAPQRGGLRWDRPPTLP
                        1110      1120      1130      1140      1150

                        1160      1170      1180      1190      1200
hPtch-2      QSFARVTTSMTVAIHPPPLPGAYIHPAPDEPPWSPAATSSGNLSSRGPGP
             *************.*********.***..**
mPatched2    QSFARVTTSMTVALHPPPLPGAYVHPASEEPT
                        1160      1170      1180

hPtch-2      ATG
```

FIG. 8D

PTCH2

PTCH

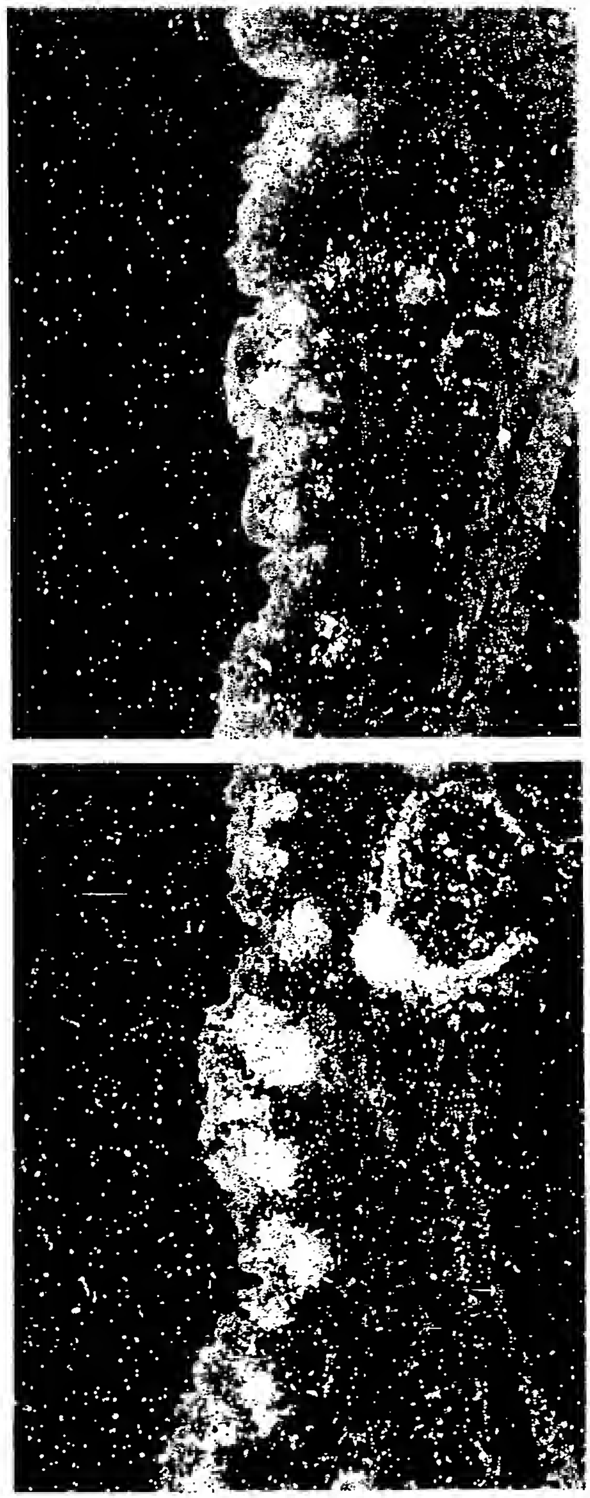


FIG. 9

> Consensus Sequence of human patched 2 cDNA clone
> length: 4004 bp

(SEQ ID NO:8)

```
  1 CCCACGCGTC CGGGAGAAGC TGGGGGAGGA GGCTGCATAC ACCTCTCAGA TGCTGATACA GACCGCACGC CAGGAGGGAG AGAACATCCT CACACCCGAA
    GGGTGCGCAG GCCCTCTTCG ACCCCCTCCT CCGACGTATG TGGAGAGTCT ACGACTATGT CTGGCGTGCG GTCCTCCCTC TCTTGTAGGA GTGTGGGCTT
                                                          race 6

101 GCACTTGGCC TCCACCTCCA GGCAGCCCTC ACTGCCAGTA AAGTCCAAGT ATCACTCTAT GGGAAGTCCT GGGATTTGAA CAAAATCTGC TACAAGTCAG
    CGTGAACCGG AGGTGGAGGT CCGTCGGGAG TGACGGTCAT TTCAGGTTCA TAGTGAGATA CCCTTCAGGA CCCTAAACTT GTTTTAGACG ATGTTCAGTC
                                                                                                              race 5'

201 GAGTTCCCCT TATTGAAAAT GGAATGATTG AGCGGATGAT TGAGAAGCTG TTTCCGTGCG TGATCCTCAC CCCCCTCGAC TGCTTCTGGG AGGGAGCCAA
    CTCAAGGGGA ATAACTTTTA CCTTACTAAC TCGCCTACTA ACTCTTCGAC AAAGGCACGC ACTAGGAGTG GGGGGAGCTG ACGAAGACCC TCCCTCGGTT

301 ACTCCAAGGG GGCTCCGCCT ACCTGCCGCT CCCAATGTGG CTCACGAGCT GAGTGGGGGC TGCCATGGCT TCTCCCACAA ATTCATGCAC TGGCAGGAGG
    TGAGGTTCCC CCGAGGCGGA TGGACGGCGA GGGTTACACC GAGTGCTCGA CTCACCCCCG ACGGTACCGA AGAGGGTGTT TAAGTACGTG ACCGTCCTCC

401 AATTGCTGCT GGGAGGCATG GCCAGAGACC CCCAAGGAGA GCTGCTGAGG GCAGAGGCCC TGCAGAGCAC CTTCTTGCTG ATGAGTCCCC GCCAGCTGTA
    TTAACGACGA CCCTCCGTAC CGGTCTCTGG GGGTTCCTCT CGACGACTCC CGTCTCCGGG ACGTCTCGTG GAAGAACGAC TACTCAGGGG CGGTCGACAT

501 CGAGCATTTC CGGGGTGACT ATCAGACACA TGACATTGGC TGGAGTGAGG AGCAGGCCAG CACAGTGCTA CAAGCCTGGC AGCGGCGCTT TGTGCAGGTC
    GCTCGTAAAG GCCCCACTGA TAGTCTGTGT ACTGTAACCG ACCTCACTCC TCGTCCGGTC GTGTCACGAT GTTCGGACCG TCGCCGCGAA ACACGTCCAG

601 GGTATGGACA AGGACAGGGG GGTGCCCTGA GGCCATTCCC TCCTCCTGCC CCCTCCTATC CACCCTGTTT CTCCAGCTGG CCCAGGAGGC CCTGCCTGAG
    CCATACCTGT TCCTGTCCCC CCACGGGACT CCGGTAAGGG AGGAGGACGG GGGAGGATAG GTGGGACAAA GAGGTCGACC GGGTCCTCCG GGACGGACTC

701 AACGCTTCCC AGCAGATCCA TGCCTTCTCC TCCACCACCC TGGATGACAT CCTGCATGCG TTCTCTGAAG TCAGTGCTGC CCGTGTGGTG GGAGGCTATC
    TTGCGAAGGG TCGTCTAGGT ACGGAAGAGG AGGTGGTGGG ACCTACTGTA GGACGTACGC AAGAGACTTC AGTCACGACG GGCACACCAC CCTCCGATAG

801 TGCTCATGGT GGGTCTTGCA CCTGGCACCT TGCCCCCACC CCACCTCCAA CCAGTGCCCA CCCTGGGGAG CCCCTGAGAC TGCCCTTTCC CCCCACAGCT
    ACGAGTACCA CCCAGAACGT GGACCGTGGA ACGGGGGTGG GGTGGAGGTT GGTCACGGGT GGGACCCCTC GGGGACTCTG ACGGGAAAGG GGGGTGTCGA
```

FIG. 10A

901 GGCCTATGCC TGTGTGACCA TGCTGCGGTG GGACTGCGCC CAGTCCCAGG GTTCCGTGGG CCTTGCCGGG GTACTGCTGG TGGCCCTGGC GGTGGCCTCA
CCGGATACGG ACACACTGGT ACGACGCCAC CCTGACGCGG GTCAGGGTCC CAAGGCACCC GGAACGGCCC CATGACGACC ACCGGGACCG CCACCGGAGT

1001 GGCCTTGGGC TCTGTGCCCT GCTCGGCATC ACCTTCAATG CTGCCACTAC CCAGGTACGC CAGGACTGCA GGGCAGACTC AGTGCCAGTC ACCAGGCTTC
CCGGAACCCG AGACACGGGA CGAGCCGTAG TGGAAGTTAC GACGGTGATG GGTCCATGCG GTCCTGACGT CCCGTCTGAG TCACGGTCAG TGGTCCGAAG

1101 ACGGGTCCTC AGCTGCCCGC TCCTCTGCCC CTCCAGGTGC TGCCCTTCTT GACTCTGGGA ATCGGCGTGG ATGACGTATT CCTGCTGGCG CATGCCTTCA
TGCCCAGGAG TCGACGGGCG AGGAGACGGG GAGGTCCACG ACGGGAAGAA CTGAGACCCT TAGCCGCACC TACTGCATAA GGACGACCGC GTACGGAAGT

1201 CAGAGGCTCT GCCTGGCACC CCTCTCCAGG TGGGGCCTTG TCCCCCAGGG CTCATCTGAG GCAGCTCAGC TTACTGGTTA AGAGCCTCTT GGTTCAAGTG
GTCTCCGAGA CGGACCGTGG GGAGAGGTCC ACCCCGGAAC AGGGGGTCCC GAGTAGACTC CGTCGAGTCG AATGACCAAT TCTCGGAGAA CCAAGTTCAC

1301 ACCTTGGGCT GCTAATGAAC CTCGGTGCCT CTTGTCCCCA TGTGTAAACA GGGGAAATAA TAGTGCTGTG TCCTAAGGGT TATTGTTTGG ATCAGTGAAG
TGGAACCCGA CGATTACTTG GAGCCACGGA GAACAGGGGT ACACATTTGT CCCCTTTATT ATCACGACAC AGGATTCCCA ATAACAAACC TAGTCACTTC

1401 TAACTCAAGT TGAATGCTTA GAACAGCCCA TCATACGTAC ATGGTACCCA ATAAATGCTA GCCACTGTGT TATGACTGCC CCACCTCTGC ACCCCAAGTT
ATTGAGTTCA ACTTACGAAT CTTGTCGGGT AGTATGCATG TACCATGGGT TATTTACGAT CGGTGACACA ATACTGACGG GGTGGAGACG TGGGGTTCAA

1501 CCTGAGCCTC CCCTTCACTC CACTTTGACA CGGCCCCTCC CTTGTGACCT GAGGGCAGGT CCCCACTCTG TCCTGGCAGG AGCGCATGGG CGAGTGTCTG
GGACTCGGAG GGGAAGTGAG GTGAAACTGT GCCGGGGAGG GAACACTGGA CTCCCGTCCA GGGGTGAGAC AGGACCGTCC TCGCGTACCC GCTCACAGAC

1601 CAGCGCACGG GCACCAGTGT TGTACTCACA TCCATCAACA ACATGGCCGC CTTCCTCATG GCTGCCCTCG TTCCCATCCC TGCGCTGCGA GCCTTCTCCC
GTCGCGTGCC CGTGGTCACA ACATGAGTGT AGGTAGTTGT TGTACCGGCG GAAGGAGTAC CGACGGGAGC AAGGGTAGGG ACGCGACGCT CGGAAGAGGG

1701 TACAGCCTGG ACCTACGGCG GCGCCACTGC CAGCGCCTTG ATGTGCTCTG CTGCTTCTCC AGGTACTGCC TGCGCCCCAG CCCCTTCCTC CCGTGACCCA
ATGTCGGACC TGGATGCCGC CGCGGTGACG GTCGCGGAAC TACACGAGAC GACGAAGAGG TCCATGACGG ACGCGGGGTC GGGGAAGGAG GGCACTGGGT

1801 CGCCAGCCTG TCCCCTCACC AGCATTTCAA GGCACAGACC TGTCATCCAC TCTCTACCTC TTCCAGTCCC TGCTCTGCTC AGGTGATTCA GATCCTGCCC
GCGGTCGGAC AGGGGAGTGG TCGTAAAGTT CCGTGTCTGG ACAGTAGGTG AGAGATGGAG AAGGTCAGGG ACGAGACGAG TCCACTAAGT CTAGGACGGG

1901 CAGGAGCTGG GGGACGGGAC AGTACCAGTG GGCATTGCCC ACCTCACTGC CACAGTTCAA GCCTTTACCC ACTGTGAAGC CAGCAGCCAG CATGTGGTCA
GTCCTCGACC CCCTGCCCTG TCATGGTCAC CCGTAACGGG TGGAGTGACG GTGTCAAGTT CGGAAATGGG TGACACTTCG GTCGTCGGTC GTACACCAGT

2001 CCATCCTGCC TCCCCAAGCC CACCTGGTGC CCCCACCTTC TGACCCACTG GGCTCTGAGC TCTTCAGCCC TGGAGGGTCC ACACGGGACC TTCTAGGCCA
GGTAGGACGG AGGGGTTCGG GTGGACCACG GGGGTGGAAG ACTGGGTGAC CCGAGACTCG AGAAGTCGGG ACCTCCCAGG TGTGCCCTGG AAGATCCGGT

FIG. 10B

```
2101 GGAGGAGGAG ACAAGGCAGA AGGCAGCCTG CAAGTCCCTG CCCTGTGCCC GCTGGAATCT TGCCCATTTC GCCCGCTATC AGTTTGCCCC GTTGCTGCTC
     CCTCCTCCTC TGTTCCGTCT TCCGTCGGAC GTTCAGGGAC GGGACACGGG CGACCTTAGA ACGGGTAAAG CGGGCGATAG TCAAACGGGG CAACGACGAG

2201 CAGTCACATG CCAAGGCCAT CGTGCTGGTG CTCTTTGGTG CTCTTCTGGG CCTGAGCCTC TACGGAGCCA CCTTGGTGCA AGACGGCCTG GCCCTGACGG
     GTCAGTGTAC GGTTCCGGTA GCACGACCAC GAGAAACCAC GAGAAGACCC GGACTCGGAG ATGCCTCGGT GGAACCACGT TCTGCCGGAC CGGGACTGCC

2301 ATGTGGTGCC TCGGGGCACC AAGGAGCATG CCTTCCTGAG CGCCCAGCTC AGGTACTTCT CCCTGTACGA GGTGGCCCTG GTGACCCAGG GTGGCTTTGA
     TACACCACGG AGCCCCGTGG TTCCTCGTAC GGAAGGACTC GCGGGTCGAG TCCATGAAGA GGGACATGCT CCACCGGGAC CACTGGGTCC CACCGAAACT

2401 CTACGCCCAC TCCCAACGCG CCCTCTTTGA TCTGCACCAG CGCTTCAGTT CCCTCAAGGC GGTGCTGCCC CCACCGGCCA CCCAGGCACC CCGCACCTGG
     GATGCGGGTG AGGGTTGCGC GGGAGAAACT AGACGTGGTC GCGAAGTCAA GGGAGTTCCG CCACGACGGG GGTGGCCGGT GGGTCCGTGG GGCGTGGACC

2501 CTGCACTATT ACCGCAACTG GCTACAGGGA ATCCAGGCTG CCTTTGACCA GGACTGGGCT TCTGGGCGCA TCACCCGCCA CTCGTACCGC AATGGCTCTG
     GACGTGATAA TGGCGTTGAC CGATGTCCCT TAGGTCCGAC GGAAACTGGT CCTGACCCGA AGACCCGCGT AGTGGGCGGT GAGCATGGCG TTACCGAGAC

2601 AGGATGGGGC CCTGGCCTAC AAGCTGCTCA TCCAGACTGG AGACGCCCAG GAGCCTCTGG ATTTCAGCCA GGTTGGGAGA GGGCTGGAGG GGTCCACTAG
     TCCTACCCCG GGACCGGATG TTCGACGAGT AGGTCTGACC TCTGCGGGTC CTCGGAGACC TAAAGTCGGT CCAACCCTCT CCCGACCTCC CCAGGTGATC

2701 TACAGGGGCT GCAGGCCTCC TGGGCCCAGG CCTTCAGCCC TCTCTGCCTC TGCAGCTGAC CACAAGGAAG CTGGTGGACA GAGAGGGACT GATTCCACCC
     ATGTCCCCGA CGTCCGGAGG ACCCGGGTCC GGAAGTCGGG AGAGACGGAG ACGTCGACTG GTGTTCCTTC GACCACCTGT CTCTCCCTGA CTAAGGTGGG

2801 GAGCTCTTCT ACATGGGGCT GACCGTGTGG GTGAGCAGTG ACCCCCTGGG TCTGGCAGCC TCACAGGCCA ACTTCTACCC CCCACCTCCT GAATGGCTGC
     CTCGAGAAGA TGTACCCCGA CTGGCACACC CACTCGTCAC TGGGGGACCC AGACCGTCGG AGTGTCCGGT TGAAGATGGG GGGTGGAGGA CTTACCGACG

2901 ACGACAAATA CGACACCACG GGGGAGAACC TTCGCAGTGA GTCTTGGGGG GAGCTCGGCA AGAGCCTCAG CCTCGCCCAC ACAAGCCCTG AGCCTGAGGC
     TGCTGTTTAT GCTGTGGTGC CCCCTCTTGG AAGCGTCACT CAGAACCCCC CTCGAGCCGT TCTCGGAGTC GGAGCGGGTG TGTTCGGGAC TCGGACTCCG

3001 CCTGCCCACT CTGCCCCGTG CTCACCGCCC TGTCCCTCTC CCTCTTCTCC CTTCCCCTCC CCTCCACAGT CCCGCCAGCT CAGCCCTTGG AGTTTGCCCA
     GGACGGGTGA GACGGGGCAC GAGTGGCGGG ACAGGGAGAG GGAGAAGAGG GAAGGGGAGG GGAGGTGTCA GGGCGGTCGA GTCGGGAACC TCAAACGGGT
```

FIG. 10C

```
3101 GTTCCCCTTC CTGCTGCGTG GCCTCCAGAA GACTGCAGAC TTTGTGGAGG CCATCGAGGG GGCCCGGGCA GCATGCGCAG AGGCCGGCCA GGCTGGGGTG
     CAAGGGGAAG GACGACGCAC CGGAGGTCTT CTGACGTCTG AAACACCTCC GGTAGCTCCC CCGGGCCCGT CGTACGCGTC TCCGGCCGGT CCGACCCCAC

3201 CACGCCTACC CCAGCGGCTC CCCCTTCCTC TTCTGGGAAC AGTATCTGGG CCTGCGGCGC TGCTTCCTGC TGGCCGTCTG CATCCTGCTG GTGTGCACTT
     GTGCGGATGG GGTCGCCGAG GGGGAAGGAG AAGACCCTTG TCATAGACCC GGACGCCGCG ACGAAGGACG ACCGGCAGAC GTAGGACGAC CACACGTGAA

3301 TCCTCGTCTG TGCTCTGCTG CTCCTCAACC CCTGGACGGC TGGCCTCATA GTGAGTGCTT GCAGGAGTGG GGACAGAGAC ACCCCACCCT TCCCTGCCCA
     AGGAGCAGAC ACGAGACGAC GAGGAGTTGG GGACCTGCCG ACCGGAGTAT CACTCACGAA CGTCCTCACC CCTGTCTCTG TGGGGTGGGA AGGGACGGGT

3401 GCCTGTCATC CCTCCTGCCA GGAGCCCTCT GTGAGCCCTG TCTCCCTCAG GTGCTGGTCC TGGCGATGAT GACAGTGGAA CTCTTTGGTA TCATGGGTTT
     CGGACAGTAG GGAGGACGGT CCTCGGGAGA CACTCGGGAC AGAGGGAGTC CACGACCAGG ACCGCTACTA CTGTCACCTT GAGAAACCAT AGTACCCAAA

3501 CCTGGGCATC AAGCTGAGTG CCATCCCCGT GGTGATCCTT GTGGCCTCTG TAGGCATTGG CGTTGAGTTC ACAGTCCACG TGGCTCTGGT GAGCACGGGC
     GGACCCGTAG TTCGACTCAC GGTAGGGGCA CCACTAGGAA CACCGGAGAC ATCCGTAACC GCAACTCAAG TGTCAGGTGC ACCGAGACCA CTCGTGCCCG

3601 ACCCCGGGGA GGGACCAATC AGCTGATTCA GTATTCAACA CATATTGTTC AAGCCCCTAC TATGTGCTAG GTACTATTTA AGAATTTGGG CTGGGTGGAC
     TGGGGCCCCT CCCTGGTTAG TCGACTAAGT CATAAGTTGT GTATAACAAG TTCGGGGATG ATACACGATC CATGATAAAT TCTTAAACCC GACCCACCTG

3701 GTGGTGGCTC ATTCCTGTAA TCCCAGCACT TTGGGAGGCC GAGGCGGGTG GATCACCTGA GGTCGGGAGT TCGAAACCAG CCTGGCCAAC ATGGTGAAAC
     CACCACCGAG TAAGGACATT AGGGTCGTGA AACCCTCCGG CTCCGCCCAC CTAGTGGACT CCAGCCCTCA AGCTTTGGTC GGACCGGTTG TACCACTTTG

3801 CCTGTCTTTA CTAAAAATAC AAAAAATTAG CCAGGCGTGG TGGCACATGC CAGTAGTCCC AGCTACTTTG GAGGCTGAGG CAGAATTGCT TGAACCTGGG
     GGACAGAAAT GATTTTTATG TTTTTTAATC GGTCCGCACC ACCGTGTACG GTCATCAGGG TCGATGAAAC CTCCGACTCC GTCTTAACGA ACTTGGACCC

3901 AGGCGAAGGT TGCAGTGAGC TGAGATCGTG CCATTGCACT CCAGCCTGGG CAACAAGAGT GCAACTCTCC GTCTCAAAAA AAAAAAAAAA AAGGGCGGCC
     TCCGCTTCCA ACGTCACTCG ACTCTAGCAC GGTAACGTGA GGTCGGACCC GTTGTTCTCA CGTTGAGAGG CAGAGTTTTT TTTTTTTTTT TTCCCGCCGG

4001 GCGA
     CGCT
```

FIG. 10D

```
Clone 16.1  human patched 2
> length: 2082 bp

>  (SEQ ID NO:9)

  1 TTCCGGCATG ACTCGATCGC CGCCCCTCAG AGAGCTGCCC CCGAGTTACA CACCCCCAGC TCGAACCGCA GCACCCCAGA TCCTAGCTGG GAGCCTGAAG
    AAGGCCGTAC TGAGCTAGCG GCGGGGAGTC TCTCGACGGG GGCTCAATGT GTGGGGGTCG AGCTTGGCGT CGTGGGGTCT AGGATCGACC CTCGGACTTC

101 GCTCCACTCT GGCTTCGTGC TTACTTCCAG GGCCTGCTCT TCTCTCTGGG ATGCGGGATC CAGAGACATT GTGGCAAAGT GCTCTTTCTG GGACTGTTGG
    CGAGGTGAGA CCGAAGCACG AATGAAGGTC CCGGACGAGA AGAGAGACCC TACGCCCTAG GTCTCTGTAA CACCGTTTCA CGAGAAAGAC CCTGACAACC

201 CCTTTGGGGC CCTGGCATTA GGTCTCCGCA TGGCCATTAT TGAGACAAAC TTGGAACAGC TCTGGGTAGA AGTGGGCAGC CGGGTGAGCC AGGAGCTGCA
    GGAAACCCCG GGACCGTAAT CCAGAGGCGT ACCGGTAATA ACTCTGTTTG AACCTTGTCG AGACCCATCT TCACCCGTCG GCCCACTCGG TCCTCGACGT

301 TTACACCAAG GAGAAGCTGG GGGAGGAGGC TGCATACACC TCTCAGATGC TGATACAGAC CGCACGCCAG GAGGGAGAGA ACATCCTCAC ACCCGAAGCA
    AATGTGGTTC CTCTTCGACC CCCTCCTCCG ACGTATGTGG AGAGTCTACG ACTATGTCTG GCGTGCGGTC CTCCCTCTCT TGTAGGAGTG TGGGCTTCGT

401 CTTGGCCTCC ACCTCCAGGC AGCCCTCACT GCCAGTAAAG TCCAAGTATC ACTCTATGGG AAGTCCTGGG ATTTGAACAA AATCTGCTAC AAGTCAGGAG
    GAACCGGAGG TGGAGGTCCG TCGGGAGTGA CGGTCATTTC AGGTTCATAG TGAGATACCC TTCAGGACCC TAAACTTGTT TTAGACGATG TTCAGTCCTC

501 TTCCCCTTAT TGAAAATGGA ATGATTGAGT GGATGATTGA GAAGCTGTTT CCGTGCGTGA TCCTCACCCC CCTCGACTGC TTCTGGGAGG GAGCCAAACT
    AAGGGGAATA ACTTTTACCT TACTAACTCA CCTACTAACT CTTCGACAAA GGCACGCACT AGGAGTGGGG GGAGCTGACG AAGACCCTCC CTCGGTTTGA

601 CCAAGGGGGC TCCGCCTACC TGCCCGGCCG CCCGGATATC CAGTGGACCA ACCTGGATCC AGAGCAGCTG CTGGAGGAGC TGGGTCCCTT TGCCTCCCTT
    GGTTCCCCCG AGGCGGATGG ACGGGCCGGC GGGCCTATAG GTCACCTGGT TGGACCTAGG TCTCGTCGAC GACCTCCTCG ACCCAGGGAA ACGGAGGGAA

701 GAGGGCTTCC GGGAGCTGCT AGACAAGGCA CAGGTGGGCC AGGCCTACGT GGGGCGGCCC TGTCTGCACC CTGATGACCT CCACTGCCCA CCTAGTGCCC
    CTCCCGAAGG CCCTCGACGA TCTGTTCCGT GTCCACCCGG TCCGGATGCA CCCCGCCGGG ACAGACGTGG GACTACTGGA GGTGACGGGT GGATCACGGG

801 CCAACCATCA CAGCAGGCAG GCTCCCAATG TGGCTCACGA GCTGAGTGGG GGCTGCCATG GCTTCTCCCA CAAATTCATG CACTGGCAGG AGGAATTGCT
    GGTTGGTAGT GTCGTCCGTC CGAGGGTTAC ACCGAGTGCT CGACTCACCC CCGACGGTAC CGAAGAGGGT GTTTAAGTAC GTGACCGTCC TCCTTAACGA

901 GCTGGGAGGC ATGGCCAGAG ACCCCCAAGG AGAGCTGCTG AGGGCAGAGG CCCTGCAGAG CACCTTCTTG CTGATGAGTC CCCGCCAGCT GTACGAGCAT
    CGACCCTCCG TACCGGTCTC TGGGGGTTCC TCTCGACGAC TCCCGTCTCC GGGACGTCTC GTGGAAGAAC GACTACTCAG GGGCGGTCGA CATGCTCGTA
```

FIG. 11A

```
1001 TTCCGGGGTG ACTATCAGAC ACATGACATT GGCTGGAGTG AGGAGCAGGC CAGCACAGTG CTACAAGCCT GGCAGCGGCG CTTTGTGCAG CTGGCCCAGG
     AAGGCCCCAC TGATAGTCTG TGTACTGTAA CCGACCTCAC TCCTCGTCCG GTCGTGTCAC GATGTTCGGA CCGTCGCCGC GAAACACGTC GACCGGGTCC

1101 AGGCCCTGCC TGAGAACGCT TCCCAGCAGA TCCATGCCTT CTCCTCCACC ACCCTGGATA ACATCCTGCA TGCGTTCTCT GAAGTCAGTG CTGCCCGTGT
     TCCGGGACGG ACTCTTGCGA AGGGTCGTCT AGGTACGGAA GAGGAGGTGG TGGGACCTAT TGTAGGACGT ACGCAAGAGA CTTCAGTCAC GACGGGCACA

1201 GGTGGGAGGC TATCTGCTCA TGCTGGCCTA TGCCTGTGTG ACCATGCTGC GGTGGGACTG CGCCCAGTCC CAGGGTTCCG TGGGCCTTGC CGGGGTACTG
     CCACCCTCCG ATAGACGAGT ACGACCGGAT ACGGACACAC TGGTACGACG CCACCCTGAC GCGGGTCAGG GTCCCAAGGC ACCCGGAACG GCCCCATGAC

1301 CTGGTGGCCC TGGCGGTGGC CTCAGGCCTT GGGCTCTGTG CCCTGCTCGG CATCACCTTC AATGCTGCCA CTACCCAGGT GCTGCCCTTC TTGGCTCTGG
     GACCACCGGG ACCGCCACCG GAGTCCGGAA CCCGAGACAC GGGACGAGCC GTAGTGGAAG TTACGACGGT GATGGGTCCA CGACGGGAAG AACCGAGACC

1401 GAATCGGCGT GGATGACGTA TTCCTGCTGG CGCATGCCTT CACAGAGGCT CTGCCTGGCA CCCCTCTCCA GGAGCGCATG GGCGAGTGTC TGCAGCGCAC
     CTTAGCCGCA CCTACTGCAT AAGGACGACC GCGTACGGAA GTGTCTCCGA GACGGACCGT GGGGAGAGGT CCTCGCGTAC CCGCTCACAG ACGTCGCGTG

1501 GGGCACCAGT GTCGTACTCA CATCCATCAA CAACATGGCC GCCTTCCTCA TGGCTGCCCT CGTTCCCATC CCTGCGCTGC GAGCCTTCTC CTTACAGCCA
     CCCGTGGTCA CAGCATGAGT GTAGGTAGTT GTTGTACCGG CGGAAGGAGT ACCGACGGGA GCAAGGGTAG GGACGCGACG CTCGGAAGAG GAATGTCGGT

1601 TCCTCAGCCT GGACCTACGG CGGCGCCACT GCCAGCGCCT TGATGTGCTC TGCTGCTTCT CCAGTCCCTG CTCTGCTCAG GTGATTCAGA TCCTGCCCCA
     AGGAGTCGGA CCTGGATGCC GCCGCGGTGA CGGTCGCGGA ACTACACGAG ACGACGAAGA GGTCAGGGAC GAGACGAGTC CACTAAGTCT AGGACGGGGT

1701 GGAGCTGGGG GACGGGACAG TACCAGTGGG CATTGCCCAC CTCACTGCCA CAGTTCAAGC CTTTACCCAC TGTGAAGCCA GCAGCCAGCA TGTGGTCACC
     CCTCGACCCC CTGCCCTGTC ATGGTCACCC GTAACGGGTG GAGTGACGGT GTCAAGTTCG GAAATGGGTG ACACTTCGGT CGTCGGTCGT ACACCAGTGG

1801 ATCCTGCCTC CCCAAGCCCA CCTGGTGCCC CCACCTTCTG ACCCACTGGG CTCTGAGCTC TTCAGCCCTG GAGGGTCCAC ACGGGACCTT CTAGGCCAGG
     TAGGACGGAG GGGTTCGGGT GGACCACGGG GGTGGAAGAC TGGGTGACCC GAGACTCGAG AAGTCGGGAC CTCCCAGGTG TGCCCTGGAA GATCCGGTCC

1901 AGGAGGAGAC AAGGCAGAAG GCAGCCTGCA AGTCCCTGCC CTGTGCCCGC TGGAATCTTG CCCATTTCGC CCCGGAATTC CTGCAGCCCG GGGGATCCAC
     TCCTCCTCTG TTCCGTCTTC CGTCGGACGT TCAGGGACGG GACACGGGCG ACCTTAGAAC GGGTAAAGCG GGGCCTTAAG GACGTCGGGC CCCCTAGGTG

2001 TAGTTCTAGA GCGGCCGCCA CCGCGGTGGA GCTCCAGCTT TTGTTCCCTT TAGTGAGGGT TAATTGCGCG CTTGGGTATC TT
     ATCAAGATCT CGCCGGCGGT GGCGCCACCT CGAGGTCGAA AACAAGGGAA ATCACTCCCA ATTAACGCGC GAACCCATAG AA
```

FIG. 11B

Sequence Listing

<110> Genentech, Inc.

<120> Patched-2

<130> P1405PCT

<141> 1999-04-02

<150> US 09/060,939
<151> 1998-04-15

<160> 32

<210> 1
<211> 4030
<212> DNA
<213> Homo sapiens

<400> 1
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ataagataca ttgatgagtt tggacaaacc acaactagaa tgcagtgaaa 100

aaaatgcttt atttgtgaaa tttgtgatgc tattgcttta tttgtaacca 150

ttataagctg caataaacaa gttgggccat ggcggccaag cttctgcagg 200

tcgactctag aggatccccg ggaattccg gcatgactcg atcgccgccc 250

ctcagagagc tgcccccgag ttacacaccc ccagctcgaa ccgcagcacc 300

ccagatccta gctgggagcc tgaaggctcc actctggctt cgtgcttact 350

tccagggcct gctcttctct ctgggatgcg ggatccagag acattgtggc 400

aaagtgctct ttctgggact gttggccttt ggggccctgg cattaggtct 450

ccgcatggcc attattgaga caaacttgga acagctctgg gtagaagtgg 500

gcagccgggt gagccaggag ctgcattaca ccaaggagaa gctgggggag 550

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tcactgccag taaagtccaa gtatcactct atgggaagtc ctgggatttg 700

aacaaaatct gctacaagtc aggagttccc cttattgaaa atggaatgat 750

tgagtggatg attgagaagc tgtttccgtg cgtgatcctc acccccctcg 800

actgcttctg ggagggagcc aaactccaag gggctccgc ctacctgccc 850

ggccgcccgg atatccagtg gaccaacctg gatccagagc agctgctgga 900

ggagctgggt ccctttgcct cccttgaggg cttccgggag ctgctagaca 950
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caatgtggct cacgagctga gtgggggctg ccatggcttc tcccacaaat 1100
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acattggctg gagtgaggag caggccagca cagtgctaca agcctggcag 1300
cggcgctttg tgcagctggc ccaggaggcc ctgcctgaga acgcttccca 1350
gcagatccat gccttctcct ccaccaccct ggatgacatc ctgcatgcgt 1400
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<211> 1203
<212> PRT
<213> Homo sapiens

<400> 2

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Pro Pro Ala Arg Thr Ala Ala Pro Gln Ile Leu Ala Gly Ser Leu
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Lys Ala Pro Leu Trp Leu Arg Ala Tyr Phe Gln Gly Leu Leu Phe
35 40 45

Ser Leu Gly Cys Gly Ile Gln Arg His Cys Gly Lys Val Leu Phe
50 55 60

Leu Gly Leu Leu Ala Phe Gly Ala Leu Ala Leu Gly Leu Arg Met
65 70 75

Ala Ile Ile Glu Thr Asn Leu Glu Gln Leu Trp Val Glu Val Gly
80 85 90

Ser Arg Val Ser Gln Glu Leu His Tyr Thr Lys Glu Lys Leu Gly
95 100 105

Glu Glu Ala Ala Tyr Thr Ser Gln Met Leu Ile Gln Thr Ala Arg
110 115 120

Gln Glu Gly Glu Asn Ile Leu Thr Pro Glu Ala Leu Gly Leu His
125 130 135

Leu Gln Ala Ala Leu Thr Ala Ser Lys Val Gln Val Ser Leu Tyr
140 145 150

Gly Lys Ser Trp Asp Leu Asn Lys Ile Cys Tyr Lys Ser Gly Val
155 160 165

Pro Leu Ile Glu Asn Gly Met Ile Glu Trp Met Ile Glu Lys Leu
170 175 180

Phe Pro Cys Val Ile Leu Thr Pro Leu Asp Cys Phe Trp Glu Gly
185 190 195

Ala Lys Leu Gln Gly Gly Ser Ala Tyr Leu Pro Gly Arg Pro Asp
200 205 210

Ile Gln Trp Thr Asn Leu Asp Pro Glu Gln Leu Leu Glu Glu Leu
215 220 225

Gly Pro Phe Ala Ser Leu Glu Gly Phe Arg Glu Leu Leu Asp Lys
230 235 240

Ala Gln Val Gly Gln Ala Tyr Val Gly Arg Pro Cys Leu His Pro
245 250 255

Asp Asp Leu His Cys Pro Pro Ser Ala Pro Asn His His Ser Arg
260 265 270

Gln Ala Pro Asn Val Ala His Glu Leu Ser Gly Gly Cys His Gly
275 280 285

Phe Ser His Lys Phe Met His Trp Gln Glu Glu Leu Leu Leu Gly
290 295 300

Gly Met Ala Arg Asp Pro Gln Gly Glu Leu Leu Arg Ala Glu Ala
305 310 315

Leu Gln Ser Thr Phe Leu Leu Met Ser Pro Arg Gln Leu Tyr Glu
320 325 330

His Phe Arg Gly Asp Tyr Gln Thr His Asp Ile Gly Trp Ser Glu
335 340 345

Glu Gln Ala Ser Thr Val Leu Gln Ala Trp Gln Arg Arg Phe Val
350 355 360

Gln Leu Ala Gln Glu Ala Leu Pro Glu Asn Ala Ser Gln Gln Ile
365 370 375

His Ala Phe Ser Ser Thr Thr Leu Asp Asp Ile Leu His Ala Phe
380 385 390

Ser Glu Val Ser Ala Ala Arg Val Val Gly Gly Tyr Leu Leu Met
395 400 405

Leu Ala Tyr Ala Cys Val Thr Met Leu Arg Trp Asp Cys Ala Gln
410 415 420

Ser Gln Gly Ser Val Gly Leu Ala Gly Val Leu Leu Val Ala Leu
425 430 435

Ala Val Ala Ser Gly Leu Gly Leu Cys Ala Leu Leu Gly Ile Thr
440 445 450

Phe Asn Ala Ala Thr Thr Gln Val Leu Pro Phe Leu Ala Leu Gly
455 460 465

Ile Gly Val Asp Asp Val Phe Leu Leu Ala His Ala Phe Thr Glu
470 475 480

Ala Leu Pro Gly Thr Pro Leu Gln Glu Arg Met Gly Glu Cys Leu
485 490 495

Gln Arg Thr Gly Thr Ser Val Val Leu Thr Ser Ile Asn Asn Met
500 505 510

Ala Ala Phe Leu Met Ala Ala Leu Val Pro Ile Pro Ala Leu Arg
515 520 525

Ala Phe Ser Leu Gln Ala Ala Ile Val Val Gly Cys Thr Phe Val
530 535 540

Ala Val Met Leu Val Phe Pro Ala Ile Leu Ser Leu Asp Leu Arg
545 550 555

Arg Arg His Cys Gln Arg Leu Asp Val Leu Cys Cys Phe Ser Ser
560 565 570

Pro Cys Ser Ala Gln Val Ile Gln Ile Leu Pro Gln Glu Leu Gly
575 580 585

Asp Gly Thr Val Pro Val Gly Ile Ala His Leu Thr Ala Thr Val
590 595 600

Gln Ala Phe Thr His Cys Glu Ala Ser Ser Gln His Val Val Thr
605 610 615

Ile Leu Pro Pro Gln Ala His Leu Val Pro Pro Pro Ser Asp Pro
620 625 630

Leu Gly Ser Glu Leu Phe Ser Pro Gly Gly Ser Thr Arg Asp Leu
635 640 645

Leu Gly Gln Glu Glu Glu Thr Arg Gln Lys Ala Ala Cys Lys Ser
650 655 660

Leu Pro Cys Ala Arg Trp Asn Leu Ala His Phe Ala Arg Tyr Gln
665 670 675

Phe Ala Pro Leu Leu Leu Gln Ser His Ala Lys Ala Ile Val Leu
680 685 690

Val Leu Phe Gly Ala Leu Leu Gly Leu Ser Leu Tyr Gly Ala Thr
695 700 705

Leu Val Gln Asp Gly Leu Ala Leu Thr Asp Val Val Pro Arg Gly
710 715 720

Thr Lys Glu His Ala Phe Leu Ser Ala Gln Leu Arg Tyr Phe Ser
725 730 735

Leu Tyr Glu Val Ala Leu Val Thr Gln Gly Gly Phe Asp Tyr Ala
740 745 750

His Ser Gln Arg Ala Leu Phe Asp Leu His Gln Arg Phe Ser Ser
755 760 765

Leu Lys Ala Val Leu Pro Pro Pro Ala Thr Gln Ala Pro Arg Thr
770 775 780

Trp Leu His Tyr Tyr Arg Asn Trp Leu Gln Gly Ile Gln Ala Ala
785 790 795

Phe Asp Gln Asp Trp Ala Ser Gly Arg Ile Thr Arg His Ser Tyr
800 805 810

Arg Asn Gly Ser Glu Asp Gly Ala Leu Ala Tyr Lys Leu Leu Ile
815 820 825

Gln Thr Gly Asp Ala Gln Glu Pro Leu Asp Phe Ser Gln Leu Thr
830 835 840

Thr Arg Lys Leu Val Asp Arg Glu Gly Leu Ile Pro Pro Glu Leu
845 850 855

Phe Tyr Met Gly Leu Thr Val Trp Val Ser Ser Asp Pro Leu Gly
860 865 870

Leu Ala Ala Ser Gln Ala Asn Phe Tyr Pro Pro Pro Pro Glu Trp
875 880 885

Leu His Asp Lys Tyr Asp Thr Thr Gly Glu Asn Leu Arg Ile Pro
890 895 900

Pro Ala Gln Pro Leu Glu Phe Ala Gln Phe Pro Phe Leu Leu Arg
905 910 915

Gly Leu Gln Lys Thr Ala Asp Phe Val Glu Ala Ile Glu Gly Ala
920 925 930

Arg Ala Ala Cys Ala Glu Ala Gly Gln Ala Gly Val His Ala Tyr
935 940 945

Pro Ser Gly Ser Pro Phe Leu Phe Trp Glu Gln Tyr Leu Gly Leu
950 955 960

Arg Arg Cys Phe Leu Leu Ala Val Cys Ile Leu Leu Val Cys Thr
965 970 975

Phe Leu Val Cys Ala Leu Leu Leu Leu Asn Pro Trp Thr Ala Gly
980 985 990

Leu Ile Val Leu Val Leu Ala Met Met Thr Val Glu Leu Phe Gly
995 1000 1005

Ile Met Gly Phe Leu Gly Ile Lys Leu Ser Ala Ile Pro Val Val
1010 1015 1020

Ile Leu Val Ala Ser Val Gly Ile Gly Val Glu Phe Thr Val His
1025 1030 1035

Val Ala Leu Gly Phe Leu Thr Thr Gln Gly Ser Arg Asn Leu Arg
1040 1045 1050

Ala Ala His Ala Leu Glu His Thr Phe Ala Pro Val Thr Asp Gly
1055 1060 1065

Ala Ile Ser Thr Leu Leu Gly Leu Leu Met Leu Ala Gly Ser His
1070 1075 1080

Phe Asp Phe Ile Val Arg Tyr Phe Phe Ala Ala Leu Thr Val Leu
1085 1090 1095

Thr Leu Leu Gly Leu Leu His Gly Leu Val Leu Leu Pro Val Leu
1100 1105 1110

Leu Ser Ile Leu Gly Pro Pro Pro Glu Val Ile Gln Met Tyr Lys
1115 1120 1125

Glu Ser Pro Glu Ile Leu Ser Pro Pro Ala Pro Gln Gly Gly Gly
1130 1135 1140

Leu Arg Trp Gly Ala Ser Ser Ser Leu Pro Gln Ser Phe Ala Arg
1145 1150 1155

Val Thr Thr Ser Met Thr Val Ala Ile His Pro Pro Pro Leu Pro
1160 1165 1170

Gly Ala Tyr Ile His Pro Ala Pro Asp Glu Pro Pro Trp Ser Pro
1175 1180 1185

Ala Ala Thr Ser Ser Gly Asn Leu Ser Ser Arg Gly Pro Gly Pro
1190 1195 1200

Ala Thr Gly
1203

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<213> Homo sapiens

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<222> 20, 27, 135, 156, 210
<223> unknown base

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gtatctgggc ctgcggcgct gcttcctgct ggccgtctgc atcctgctgg 100

tgtgcacttt cctcgtctgt gctctgctgc tcctnaaccc ctggacggct 150

ggcctnatag tgctggtcct ggcgatgatg acagtggaac tctttggtat 200

catgggtttn ctgggcatca agctgagt 228

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<211> 76
<212> PRT
<213> Homo sapiens

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Glu Gln Tyr Ile Gly Leu Arg His Trp Leu Leu Leu Phe Ile Ser
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Val Val Leu Ala Cys Thr Phe Leu Val Cys Ala Val Phe Leu Leu
35 40 45

Asn Pro Trp Thr Ala Gly Ile Ile Val Met Val Leu Ala Leu Met
50 55 60

Thr Val Glu Leu Phe Gly Met Met Gly Leu Ile Gly Ile Lys Leu
65 70 75

Ser
76

<210> 5
<211> 125
<212> DNA
<213> Homo sapiens

<220>
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<222> 115
<223> unknown base

<400> 5
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gtatctgggc ctgcggcgct gcttcctgct ggccgtctgc atcctgctgg 100

tgtgcacttt cctcntctgt gctct 125

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<212> DNA
<213> Homo sapiens

<220>
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<223> unknown base

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<210> 7
<211> 1182

<212> PRT
<213> Mus musculus

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Pro Pro Ala Arg Ser Ser Ala Pro His Ile Leu Ala Gly Ser Leu
20 25 30

Gln Ala Pro Leu Trp Leu Arg Ala Tyr Phe Gln Gly Leu Leu Phe
35 40 45

Ser Leu Gly Cys Arg Ile Gln Lys His Cys Gly Lys Val Leu Phe
50 55 60

Leu Gly Leu Val Ala Phe Gly Ala Leu Ala Leu Gly Leu Arg Val
65 70 75

Ala Val Ile Glu Thr Asp Leu Glu Gln Leu Trp Val Glu Val Gly
80 85 90

Ser Arg Val Ser Gln Glu Leu His Tyr Thr Lys Glu Lys Leu Gly
95 100 105

Glu Glu Ala Ala Tyr Thr Ser Gln Met Leu Ile Gln Thr Ala His
110 115 120

Gln Glu Gly Gly Asn Val Leu Thr Pro Glu Ala Leu Asp Leu His
125 130 135

Leu Gln Ala Ala Leu Thr Ala Ser Lys Val Gln Val Ser Leu Tyr
140 145 150

Gly Lys Ser Trp Asp Leu Asn Lys Ile Cys Tyr Lys Ser Gly Val
155 160 165

Pro Leu Ile Glu Asn Gly Met Ile Glu Arg Met Ile Glu Lys Leu
170 175 180

Phe Pro Cys Val Ile Leu Thr Pro Leu Asp Cys Phe Trp Glu Gly
185 190 195

Ala Lys Leu Gln Gly Gly Ser Ala Tyr Leu Pro Gly Arg Pro Asp
200 205 210

Ile Gln Trp Thr Asn Leu Asp Pro Gln Gln Leu Leu Glu Glu Leu
215 220 225

Gly Pro Phe Ala Ser Leu Glu Gly Phe Arg Glu Leu Leu Asp Lys
230 235 240

Ala Gln Val Gly Gln Ala Tyr Val Gly Arg Pro Cys Leu Asp Pro
245 250 255

Asp Asp Pro His Cys Pro Pro Ser Ala Pro Asn Arg His Ser Arg

260 265 270

Gln Ala Pro Asn Val Ala Gln Glu Leu Ser Gly Gly Cys His Gly
275 280 285

Phe Ser His Lys Phe Met His Trp Gln Glu Glu Leu Leu Leu Gly
290 295 300

Gly Thr Ala Arg Asp Leu Gln Gly Gln Leu Leu Arg Ala Glu Ala
305 310 315

Leu Gln Ser Thr Phe Leu Leu Met Ser Pro Arg Gln Leu Tyr Glu
320 325 330

His Phe Arg Gly Asp Tyr Gln Thr His Asp Ile Gly Trp Ser Glu
335 340 345

Glu Gln Ala Ser Met Val Leu Gln Ala Trp Gln Arg Arg Phe Val
350 355 360

Gln Leu Ala Gln Glu Ala Leu Pro Ala Asn Ala Ser Gln Gln Ile
365 370 375

His Ala Phe Ser Ser Thr Thr Leu Asp Asp Ile Leu Arg Ala Phe
380 385 390

Ser Glu Val Ser Thr Thr Arg Val Val Gly Gly Tyr Leu Leu Met
395 400 405

Leu Ala Tyr Ala Cys Val Thr Met Leu Arg Trp Asp Cys Ala Gln
410 415 420

Ser Gln Gly Ala Val Gly Leu Ala Gly Val Leu Leu Val Ala Leu
425 430 435

Ala Val Ala Ser Gly Leu Gly Leu Cys Ala Leu Leu Gly Ile Thr
440 445 450

Phe Asn Ala Ala Thr Thr Gln Val Leu Pro Phe Leu Ala Leu Gly
455 460 465

Ile Gly Val Asp Asp Ile Phe Leu Leu Ala His Ala Phe Thr Lys
470 475 480

Ala Pro Pro Asp Thr Pro Leu Pro Glu Arg Met Gly Glu Cys Leu
485 490 495

Arg Ser Thr Gly Thr Ser Val Ala Leu Thr Ser Val Asn Asn Met
500 505 510

Val Ala Phe Phe Met Ala Ala Leu Val Pro Ile Pro Ala Leu Arg
515 520 525

Ala Phe Ser Leu Gln Ala Ala Ile Val Val Gly Cys Asn Phe Ala
530 535 540

Ala Val Met Leu Val Phe Pro Ala Ile Leu Ser Leu Asp Leu Arg
545 550 555

Arg Arg His Arg Gln Arg Leu Asp Val Leu Cys Cys Phe Ser Ser
560 565 570

Pro Cys Ser Ala Gln Val Ile Gln Met Leu Pro Gln Glu Leu Gly
575 580 585

Asp Arg Ala Val Pro Val Gly Ile Ala His Leu Thr Ala Thr Val
590 595 600

Gln Ala Phe Thr His Cys Glu Ala Ser Ser Gln His Val Val Thr
605 610 615

Ile Leu Pro Pro Gln Ala His Leu Leu Ser Pro Ala Ser Asp Pro
620 625 630

Leu Gly Ser Glu Leu Tyr Ser Pro Gly Gly Ser Thr Arg Asp Leu
635 640 645

Leu Ser Gln Glu Glu Gly Thr Gly Pro Gln Ala Ala Cys Arg Pro
650 655 660

Leu Leu Cys Ala His Trp Thr Leu Ala His Phe Ala Arg Tyr Gln
665 670 675

Phe Ala Pro Leu Leu Leu Gln Thr Arg Ala Lys Ala Leu Val Leu
680 685 690

Leu Phe Phe Gly Ala Leu Leu Gly Leu Ser Leu Tyr Gly Ala Thr
695 700 705

Leu Val Gln Asp Gly Leu Ala Leu Thr Asp Val Val Pro Arg Gly
710 715 720

Thr Lys Glu His Ala Phe Leu Ser Ala Gln Leu Arg Tyr Phe Ser
725 730 735

Leu Tyr Glu Val Ala Leu Val Thr Gln Gly Gly Phe Asp Tyr Ala
740 745 750

His Ser Gln Arg Ala Leu Phe Asp Leu His Gln Arg Phe Ser Ser
755 760 765

Leu Lys Ala Val Leu Pro Pro Pro Ala Thr Gln Ala Pro Arg Thr
770 775 780

Trp Leu His Tyr Tyr Arg Ser Trp Leu Gln Gly Ile Gln Ala Ala
785 790 795

Phe Asp Gln Asp Trp Ala Ser Gly Arg Ile Thr Cys His Ser Tyr
800 805 810

Arg Asn Gly Ser Glu Asp Gly Ala Leu Ala Tyr Lys Leu Leu Ile
815 820 825

Gln Thr Gly Asn Ala Gln Glu Pro Leu Asp Phe Ser Gln Leu Thr
830 835 840

Thr Arg Lys Leu Val Asp Lys Glu Gly Leu Ile Pro Pro Glu Leu
845 850 855

Phe Tyr Met Gly Leu Thr Val Trp Val Ser Ser Asp Pro Leu Gly
860 865 870

Leu Ala Ala Ser Gln Ala Asn Phe Tyr Pro Pro Pro Pro Glu Trp
875 880 885

Leu His Asp Lys Tyr Asp Thr Thr Gly Glu Asn Leu Arg Ile Pro
890 895 900

Ala Ala Gln Pro Leu Glu Phe Ala Gln Phe Pro Phe Leu Leu His
905 910 915

Gly Leu Gln Lys Thr Ala Asp Phe Val Glu Ala Ile Glu Gly Ala
920 925 930

Arg Ala Ala Cys Thr Glu Ala Gly Gln Ala Gly Val His Ala Tyr
935 940 945

Pro Ser Gly Ser Pro Phe Leu Phe Trp Glu Gln Tyr Leu Gly Leu
950 955 960

Arg Arg Cys Phe Leu Leu Ala Val Cys Ile Leu Leu Val Cys Thr
965 970 975

Phe Leu Val Cys Ala Leu Leu Leu Leu Ser Pro Trp Thr Ala Gly
980 985 990

Leu Ile Val Leu Val Leu Ala Met Met Thr Val Glu Leu Phe Gly
995 1000 1005

Ile Met Gly Phe Leu Gly Ile Lys Leu Ser Ala Ile Pro Val Val
1010 1015 1020

Ile Leu Val Ala Ser Ile Gly Ile Gly Val Glu Phe Thr Val His
1025 1030 1035

Val Ala Leu Gly Phe Leu Thr Ser His Gly Ser Arg Asn Leu Arg
1040 1045 1050

Ala Ala Ser Ala Leu Glu Gln Thr Phe Ala Pro Val Thr Asp Gly
1055 1060 1065

Ala Val Ser Thr Leu Leu Gly Leu Leu Met Leu Ala Gly Ser Asn
1070 1075 1080

Phe Asp Phe Ile Ile Arg Tyr Phe Phe Val Val Leu Thr Val Leu
1085 1090 1095

Thr Leu Leu Gly Leu Leu His Gly Leu Leu Leu Leu Pro Val Leu
1100 1105 1110

Leu Ser Ile Leu Gly Pro Pro Pro Gln Val Val Gln Val Tyr Lys
1115 1120 1125

Glu Ser Pro Gln Thr Leu Asn Ser Ala Ala Pro Gln Arg Gly Gly
1130 1135 1140

Leu Arg Trp Asp Arg Pro Pro Thr Leu Pro Gln Ser Phe Ala Arg
1145 1150 1155

Val Thr Thr Ser Met Thr Val Ala Leu His Pro Pro Pro Leu Pro
1160 1165 1170

Gly Ala Tyr Val His Pro Ala Ser Glu Glu Pro Thr
1175 1180 1182

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<211> 4004
<212> DNA
<213> Homo sapiens

<400> 8
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gcacttggcc tccacctcca ggcagccctc actgccagta aagtccaagt 150

atcactctat gggaagtcct gggatttgaa caaaatctgc tacaagtcag 200

gagttcccct tattgaaaat ggaatgattg agcggatgat tgagaagctg 250

tttccgtgcg tgatcctcac ccccctcgac tgcttctggg agggagccaa 300

actccaaggg ggctccgcct acctgccgct cccaatgtgg ctcacgagct 350

gagtgggggc tgccatggct tctcccacaa attcatgcac tggcaggagg 400

aattgctgct gggaggcatg gccagagacc cccaaggaga gctgctgagg 450

gcagaggccc tgcagagcac cttcttgctg atgagtcccc gccagctgta 500

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ggtatggaca aggacagggg ggtgccctga ggccattccc tcctcctgcc 650

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aggcgaaggt tgcagtgagc tgagatcgtg ccattgcact ccagcctggg 3950

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gcga 4004

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<212> DNA
<213> Homo sapiens

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ttccccttat tgaaaatgga atgattgagt ggatgattga gaagctgttt 550

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ccaagggggc tccgcctacc tgcccggccg cccggatatc cagtggacca 650

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<210> 10
<211> 1315
<212> PRT
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35 40 45

Arg Asn Leu Gln Arg Glu Ile Glu Ile Met Arg Gly Leu Arg His
50 55 60

Pro Asn Ile Val His Met Leu Asp Ser Phe Glu Thr Asp Lys Glu
65 70 75

Val Val Val Val Thr Asp Tyr Ala Glu Gly Glu Leu Phe Gln Ile
80 85 90

Leu Glu Asp Asp Gly Lys Leu Pro Glu Asp Gln Val Gln Ala Ile
95 100 105

Ala Ala Gln Leu Val Ser Ala Leu Tyr Tyr Leu His Ser His Arg
110 115 120

Ile Leu His Arg Asp Met Lys Pro Gln Asn Ile Leu Leu Ala Lys
125 130 135

Gly Gly Gly Ile Lys Leu Cys Asp Phe Gly Phe Ala Arg Ala Met
140 145 150

Ser Thr Asn Thr Met Val Leu Thr Ser Ile Lys Gly Thr Pro Leu
155 160 165

Tyr Met Ser Pro Glu Leu Val Glu Glu Arg Pro Tyr Asp His Thr
170 175 180

Ala Asp Leu Trp Ser Val Gly Cys Ile Leu Tyr Glu Leu Ala Val
185 190 195

Gly Thr Pro Pro Phe Tyr Ala Thr Ser Ile Phe Gln Leu Val Ser
200 205 210

Leu Ile Leu Lys Asp Pro Val Arg Trp Pro Ser Thr Ile Ser Pro
215 220 225

Cys Phe Lys Asn Phe Leu Gln Gly Leu Leu Thr Lys Asp Pro Arg
230 235 240

Gln Arg Leu Ser Trp Pro Asp Leu Leu Tyr His Pro Phe Ile Ala
245 250 255

Gly His Val Thr Ile Ile Thr Glu Pro Ala Gly Pro Asp Leu Gly
260 265 270

Thr Pro Phe Thr Ser Arg Leu Pro Pro Glu Leu Gln Val Leu Lys
275 280 285

Asp Glu Gln Ala His Arg Leu Ala Pro Lys Gly Asn Gln Ser Arg
290 295 300

Ile Leu Thr Gln Ala Tyr Lys Arg Met Ala Glu Glu Ala Met Gln
305 310 315

Lys Lys His Gln Asn Thr Gly Pro Ala Leu Glu Gln Glu Asp Lys
320 325 330

Thr Ser Lys Val Ala Pro Gly Thr Ala Pro Leu Pro Arg Leu Gly
335 340 345

Ala Thr Pro Gln Glu Ser Ser Leu Leu Ala Gly Ile Leu Ala Ser
350 355 360

Glu Leu Lys Ser Ser Trp Ala Lys Ser Gly Thr Gly Glu Val Pro
365 370 375

Ser Ala Pro Arg Glu Asn Arg Thr Thr Pro Asp Cys Glu Arg Ala
380 385 390

Phe Pro Glu Glu Arg Pro Glu Val Leu Gly Gln Arg Ser Thr Asp
395 400 405

Val Val Asp Leu Glu Asn Glu Glu Pro Asp Ser Asp Asn Glu Trp
410 415 420

Gln His Leu Leu Glu Thr Thr Glu Pro Val Pro Ile Gln Leu Lys
425 430 435

Ala Pro Leu Thr Leu Leu Cys Asn Pro Asp Phe Cys Gln Arg Ile
440 445 450

Gln Ser Gln Leu His Glu Ala Gly Gly Gln Ile Leu Lys Gly Ile
455 460 465

Leu Glu Gly Ala Ser His Ile Leu Pro Ala Phe Arg Val Leu Ser
470 475 480

Ser Leu Leu Ser Ser Cys Ser Asp Ser Val Ala Leu Tyr Ser Phe
485 490 495

Cys Arg Glu Ala Gly Leu Pro Gly Leu Leu Leu Ser Leu Leu Arg
500 505 510

His Ser Gln Glu Ser Asn Ser Leu Gln Gln Gln Ser Trp Tyr Gly
515 520 525

Thr Phe Leu Gln Asp Leu Met Ala Val Ile Gln Ala Tyr Phe Ala
530 535 540

Cys Thr Phe Asn Leu Glu Arg Ser Gln Thr Ser Asp Ser Leu Gln
545 550 555

Val Phe Gln Glu Ala Ala Asn Leu Phe Leu Asp Leu Leu Gly Lys
560 565 570

Leu Leu Ala Gln Pro Asp Asp Ser Glu Gln Thr Leu Arg Arg Asp
575 580 585

Ser Leu Met Cys Phe Thr Val Leu Cys Glu Ala Met Asp Gly Asn
590 595 600

Ser Arg Ala Ile Ser Lys Ala Phe Tyr Ser Ser Leu Leu Thr Thr
605 610 615

Gln Gln Val Val Leu Asp Gly Leu Leu His Gly Leu Thr Val Pro
620 625 630

Gln Leu Pro Val His Thr Pro Gln Gly Ala Pro Gln Val Ser Gln
635 640 645

Pro Leu Arg Glu Gln Ser Glu Asp Ile Pro Gly Ala Ile Ser Ser
650 655 660

Ala Leu Ala Ala Ile Cys Thr Ala Pro Val Gly Leu Pro Asp Cys
665 670 675

Trp Asp Ala Lys Glu Gln Val Cys Trp His Leu Ala Asn Gln Leu
680 685 690

Thr Glu Asp Ser Ser Gln Leu Arg Pro Ser Leu Ile Ser Gly Leu
695 700 705

Gln His Pro Ile Leu Cys Leu His Leu Leu Lys Val Leu Tyr Ser
710 715 720

Cys Cys Leu Val Ser Glu Gly Leu Cys Arg Leu Leu Gly Gln Glu
725 730 735

Pro Leu Ala Leu Glu Ser Leu Phe Met Leu Ile Gln Gly Lys Val
740 745 750

Lys Val Val Asp Trp Glu Glu Ser Thr Glu Val Thr Leu Tyr Phe
755 760 765

Leu Ser Leu Leu Val Phe Arg Leu Gln Asn Leu Pro Cys Gly Met
770 775 780

Glu Lys Leu Gly Ser Asp Val Ala Thr Leu Phe Thr His Ser His
785 790 795

Val Val Ser Leu Val Ser Ala Ala Ala Cys Leu Leu Gly Gln Leu
800 805 810

Gly Gln Gln Gly Val Thr Phe Asp Leu Gln Pro Met Glu Trp Met
815 820 825

Ala Ala Ala Thr His Ala Leu Ser Ala Pro Ala Glu Val Arg Leu
830 835 840

Thr Pro Pro Gly Ser Cys Gly Phe Tyr Asp Gly Leu Leu Ile Leu
845 850 855

Leu Leu Gln Leu Leu Thr Glu Gln Gly Lys Ala Ser Leu Ile Arg
860 865 870

Asp Met Ser Ser Ser Glu Met Trp Thr Val Leu Trp His Arg Phe
875 880 885

Ser Met Val Leu Arg Leu Pro Glu Glu Ala Ser Ala Gln Glu Gly
890 895 900

Glu Leu Ser Leu Ser Ser Pro Pro Ser Pro Glu Pro Asp Trp Thr
905 910 915

Leu Ile Ser Pro Gln Gly Met Ala Ala Leu Leu Ser Leu Ala Met
920 925 930

Ala Thr Phe Thr Gln Glu Pro Gln Leu Cys Leu Ser Cys Leu Ser
935 940 945

Gln His Gly Ser Ile Leu Met Ser Ile Leu Lys His Leu Leu Cys
950 955 960

Pro Ser Phe Leu Asn Gln Leu Arg Gln Ala Pro His Gly Ser Glu
965 970 975

Phe Leu Pro Val Val Val Leu Ser Val Cys Gln Leu Leu Cys Phe
980 985 990

Pro Phe Ala Leu Asp Met Asp Ala Asp Leu Leu Ile Val Val Leu
995 1000 1005

Ala Asp Leu Arg Asp Ser Glu Val Ala Ala His Leu Leu Gln Val
1010 1015 1020

Cys Cys Tyr His Leu Pro Leu Met Gln Val Glu Leu Pro Ile Ser
1025 1030 1035

Leu Leu Thr Arg Leu Ala Leu Met Asp Pro Thr Ser Leu Asn Gln
1040 1045 1050

Phe Val Asn Thr Val Ser Ala Ser Pro Arg Thr Ile Val Ser Phe
1055 1060 1065

Leu Ser Val Ala Leu Leu Ser Asp Gln Pro Leu Leu Thr Ser Asp
1070 1075 1080

Leu Leu Ser Leu Leu Ala His Thr Ala Arg Val Leu Ser Pro Ser
1085 1090 1095

His Leu Ser Phe Ile Gln Glu Leu Leu Ala Gly Ser Asp Glu Ser
1100 1105 1110

Tyr Arg Pro Leu Arg Ser Leu Leu Gly His Pro Glu Asn Ser Val
1115 1120 1125

Arg Ala His Thr Tyr Arg Leu Leu Gly His Leu Leu Gln His Ser
1130 1135 1140

Met Ala Leu Arg Gly Ala Leu Gln Ser Gln Ser Gly Leu Leu Ser
1145 1150 1155

Leu Leu Leu Leu Gly Leu Gly Asp Lys Asp Pro Val Val Arg Cys
1160 1165 1170

Ser Ala Ser Phe Ala Val Gly Asn Ala Ala Tyr Gln Ala Gly Pro
1175 1180 1185

Leu Gly Pro Ala Leu Ala Ala Ala Val Pro Ser Met Thr Gln Leu
1190 1195 1200

Leu Gly Asp Pro Gln Ala Gly Ile Arg Arg Asn Val Ala Ser Ala
1205 1210 1215

Leu Gly Asn Leu Gly Pro Glu Gly Leu Gly Glu Glu Leu Leu Gln
1220 1225 1230

Cys Glu Val Pro Gln Arg Leu Leu Glu Met Ala Cys Gly Asp Pro
1235 1240 1245

Gln Pro Asn Val Lys Glu Ala Ala Leu Ile Ala Leu Arg Ser Leu
1250 1255 1260

Gln Gln Glu Pro Gly Ile His Gln Val Leu Val Ser Leu Gly Ala
1265 1270 1275

Ser Glu Lys Leu Ser Leu Leu Ser Leu Gly Asn Gln Ser Leu Pro
1280 1285 1290

His Ser Ser Pro Arg Pro Ala Ser Ala Lys His Cys Arg Lys Leu
1295 1300 1305

Ile His Leu Leu Arg Pro Ala His Ser Met
1310 1315

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<212> DNA
<213> Artificial sequence

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<223> Artificial sequence 1-48

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<212> DNA
<213> Artificial sequence

<220>
<223> Artificial sequence 1-48

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<210> 13

<211> 396
<212> PRT
<213> Mus musculus

<400> 13

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Leu Ala Leu Ser Ala Gln Ser Cys Gly Pro Gly Arg Gly Pro Val
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Gly Arg Arg Arg Tyr Val Arg Lys Gln Leu Val Pro Leu Leu Tyr
35 40 45

Lys Gln Phe Val Pro Ser Met Pro Glu Arg Thr Leu Gly Ala Ser
50 55 60

Gly Pro Ala Glu Gly Arg Val Thr Arg Gly Ser Glu Arg Phe Arg
65 70 75

Asp Leu Val Pro Asn Tyr Asn Pro Asp Ile Ile Phe Lys Asp Glu
80 85 90

Glu Asn Ser Gly Ala Asp Arg Leu Met Thr Glu Arg Cys Lys Glu
95 100 105

Arg Val Asn Ala Leu Ala Ile Ala Val Met Asn Met Trp Pro Gly
110 115 120

Val Arg Leu Arg Val Thr Glu Gly Trp Asp Glu Asp Gly His His
125 130 135

Ala Gln Asp Ser Leu His Tyr Glu Gly Arg Ala Leu Asp Ile Thr
140 145 150

Thr Ser Asp Arg Asp Arg Asn Lys Tyr Gly Leu Leu Ala Arg Leu
155 160 165

Ala Val Glu Ala Gly Phe Asp Trp Val Tyr Tyr Glu Ser Arg Asn
170 175 180

His Ile His Val Ser Val Lys Ala Asp Asn Ser Leu Ala Val Arg
185 190 195

Ala Gly Gly Cys Phe Pro Gly Asn Ala Thr Val Arg Leu Arg Ser
200 205 210

Gly Glu Arg Lys Gly Leu Arg Glu Leu His Arg Gly Asp Trp Val
215 220 225

Leu Ala Ala Asp Ala Ala Gly Arg Val Val Pro Thr Pro Val Leu
230 235 240

Leu Phe Leu Asp Arg Asp Leu Gln Arg Arg Ala Ser Phe Val Ala
245 250 255

Val Glu Thr Glu Arg Pro Pro Arg Lys Leu Leu Leu Thr Pro Trp
260 265 270

His Leu Val Phe Ala Ala Arg Gly Pro Ala Pro Ala Pro Gly Asp
275 280 285

Phe Ala Pro Val Phe Ala Arg Arg Leu Arg Ala Gly Asp Ser Val
290 295 300

Leu Ala Pro Gly Gly Asp Ala Leu Gln Pro Ala Arg Val Ala Arg
305 310 315

Val Ala Arg Glu Glu Ala Val Gly Val Phe Ala Pro Leu Thr Ala
320 325 330

His Gly Thr Leu Leu Val Asn Asp Val Leu Ala Ser Cys Tyr Ala
335 340 345

Val Leu Glu Ser His Gln Trp Ala His Arg Ala Phe Ala Pro Leu
350 355 360

Arg Leu Leu His Ala Leu Gly Ala Leu Leu Pro Gly Gly Ala Val
365 370 375

Gln Pro Thr Gly Met His Trp Tyr Ser Arg Leu Leu Tyr Arg Leu
380 385 390

Ala Glu Glu Leu Met Gly
395 396

<210> 14
<211> 437
<212> PRT
<213> Mus musculus

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20 25 30

Phe Gly Lys Arg Arg His Pro Lys Lys Leu Thr Pro Leu Ala Tyr
35 40 45

Lys Gln Phe Ile Pro Asn Val Ala Glu Lys Thr Leu Gly Ala Ser
50 55 60

Gly Arg Tyr Glu Gly Lys Ile Thr Arg Asn Ser Glu Arg Phe Lys
65 70 75

Glu Leu Thr Pro Asn Tyr Asn Pro Asp Ile Ile Phe Lys Asp Glu
80 85 90

Glu Asn Thr Gly Ala Asp Arg Leu Met Thr Gln Arg Cys Lys Asp
95 100 105

Lys Leu Asn Ala Leu Ala Ile Ser Val Met Asn Gln Trp Pro Gly
110 115 120

Val Arg Leu Arg Val Thr Glu Gly Trp Asp Glu Asp Gly His His
125 130 135

Ser Glu Glu Ser Leu His Tyr Glu Gly Arg Ala Val Asp Ile Thr
140 145 150

Thr Ser Asp Arg Asp Arg Ser Lys Tyr Gly Met Leu Ala Arg Leu
155 160 165

Ala Val Glu Ala Gly Phe Asp Trp Val Tyr Tyr Glu Ser Lys Ala
170 175 180

His Ile His Cys Ser Val Lys Ala Glu Asn Ser Val Ala Ala Lys
185 190 195

Ser Gly Gly Cys Phe Pro Gly Ser Ala Thr Val His Leu Glu Gln
200 205 210

Gly Gly Thr Lys Leu Val Lys Asp Leu Arg Pro Gly Asp Arg Val
215 220 225

Leu Ala Ala Asp Asp Gln Gly Arg Leu Leu Tyr Ser Asp Phe Leu
230 235 240

Thr Phe Leu Asp Arg Asp Glu Gly Ala Lys Lys Val Phe Tyr Val
245 250 255

Ile Glu Thr Leu Glu Pro Arg Glu Arg Leu Leu Leu Thr Ala Ala
260 265 270

His Leu Leu Phe Val Ala Pro His Asn Asp Ser Gly Pro Thr Pro
275 280 285

Gly Pro Ser Ala Leu Phe Ala Ser Arg Val Arg Pro Gly Gln Arg
290 295 300

Val Tyr Val Val Ala Glu Arg Gly Gly Asp Arg Arg Leu Leu Pro
305 310 315

Ala Ala Val His Ser Val Thr Leu Arg Glu Glu Glu Ala Gly Ala
320 325 330

Tyr Ala Pro Leu Thr Ala His Gly Thr Ile Leu Ile Asn Arg Val
335 340 345

Leu Ala Ser Cys Tyr Ala Val Ile Glu Glu His Ser Trp Ala His
350 355 360

Arg Ala Phe Ala Pro Phe Arg Leu Ala His Ala Leu Leu Ala Ala
365 370 375

Leu Ala Pro Ala Arg Thr Asp Gly Gly Gly Gly Gly Ser Ile Pro
380 385 390

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Ala Ala Gln Ser Ala Thr Glu Ala Arg Gly Ala Glu Pro Thr Ala
                395                 400                 405

Gly Ile His Trp Tyr Ser Gln Leu Leu Tyr His Ile Gly Thr Trp
                410                 415                 420

Leu Leu Asp Ser Glu Thr Met His Pro Leu Gly Met Ala Val Lys
                425                 430                 435

Ala Ser
    437

<210> 15
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<212> PRT
<213> Artificial sequence

<220>
<223> Artificial sequence 1-803

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                 20                  25                  30

Arg Gly Ala Ala Leu Ser Gly Asn Val Thr Gly Pro Gly Pro Arg
                 35                  40                  45

Ser Ala Gly Gly Ser Ala Arg Arg Asn Ala Pro Val Thr Ser Pro
                 50                  55                  60

Pro Pro Pro Leu Leu Ser His Cys Gly Arg Ala Ala His Cys Glu
                 65                  70                  75

Pro Leu Arg Tyr Asn Val Cys Leu Gly Ser Ala Leu Pro Tyr Gly
                 80                  85                  90

Ala Thr Thr Thr Leu Leu Ala Gly Asp Ser Asp Ser Gln Glu Glu
                 95                 100                 105

Ala His Ser Lys Leu Val Leu Trp Ser Gly Leu Arg Asn Ala Pro
                110                 115                 120

Arg Cys Trp Ala Val Ile Gln Pro Leu Leu Cys Ala Val Tyr Met
                125                 130                 135

Pro Lys Cys Glu Asn Asp Arg Val Glu Leu Pro Ser Arg Thr Leu
                140                 145                 150

Cys Gln Ala Thr Arg Gly Pro Cys Ala Ile Val Glu Arg Glu Arg
                155                 160                 165

Gly Trp Pro Asp Phe Leu Arg Cys Thr Pro Asp His Phe Pro Glu
                170                 175                 180
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Gly Cys Pro Asn Glu Val Gln Asn Ile Lys Phe Asn Ser Ser Gly
                185                 190                 195

Gln Cys Glu Ala Pro Leu Val Arg Thr Asp Asn Pro Lys Ser Trp
                200                 205                 210

Tyr Glu Asp Val Glu Gly Cys Gly Ile Gln Cys Gln Asn Pro Leu
                215                 220                 225

Phe Thr Glu Ala Glu His Gln Asp Met His Ser Tyr Ile Ala Ala
                230                 235                 240

Phe Gly Ala Val Thr Gly Leu Cys Thr Leu Phe Thr Leu Ala Thr
                245                 250                 255

Phe Val Ala Asp Trp Arg Asn Ser Asn Arg Tyr Pro Ala Val Ile
                260                 265                 270

Leu Phe Tyr Val Asn Ala Cys Phe Phe Val Gly Ser Ile Gly Trp
                275                 280                 285

Leu Ala Gln Phe Met Asp Gly Ala Arg Arg Glu Ile Val Cys Arg
                290                 295                 300

Ala Asp Gly Thr Met Arg Phe Gly Glu Pro Thr Ser Ser Glu Thr
                305                 310                 315

Leu Ser Cys Val Ile Ile Phe Val Ile Val Tyr Tyr Ala Leu Met
                320                 325                 330

Ala Gly Val Val Trp Phe Val Val Leu Thr Tyr Ala Trp His Thr
                335                 340                 345

Ser Phe Lys Ala Leu Gly Thr Thr Tyr Gln Pro Leu Ser Gly Lys
                350                 355                 360

Thr Ser Tyr Phe His Leu Leu Thr Trp Ser Leu Pro Phe Val Leu
                365                 370                 375

Thr Val Ala Ile Leu Ala Val Ala Gln Val Asp Gly Asp Ser Val
                380                 385                 390

Ser Gly Ile Cys Phe Val Gly Tyr Lys Asn Tyr Arg Tyr Arg Ala
                395                 400                 405

Gly Phe Val Leu Ala Pro Ile Gly Leu Val Leu Ile Val Gly Gly
                410                 415                 420

Tyr Phe Leu Ile Arg Gly Val Met Thr Leu Phe Ser Ile Lys Ser
                425                 430                 435

Asn His Pro Gly Leu Leu Ser Glu Lys Ala Ala Ser Lys Ile Asn
                440                 445                 450

Glu Thr Met Leu Arg Leu Gly Ile Phe Gly Phe Leu Ala Phe Gly
                455                 460                 465
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Phe Val Leu Ile Thr Phe Ser Cys His Phe Tyr Asp Phe Phe Asn
470 475 480

Gln Ala Glu Trp Glu Arg Ser Phe Arg Asp Tyr Val Leu Cys Gln
485 490 495

Ala Asn Val Thr Ile Gly Leu Pro Thr Lys Lys Pro Ile Pro Asp
500 505 510

Cys Glu Ile Lys Asn Arg Pro Ser Leu Leu Val Glu Lys Ile Asn
515 520 525

Leu Phe Ala Met Phe Gly Thr Gly Ile Ala Met Ser Thr Trp Val
530 535 540

Trp Thr Lys Ala Thr Leu Leu Ile Trp Arg Arg Thr Trp Cys Arg
545 550 555

Leu Thr Gly His Ser Asp Asp Glu Pro Lys Arg Ile Lys Lys Ser
560 565 570

Lys Met Ile Ala Lys Ala Phe Ser Lys Arg Arg Glu Leu Leu Gln
575 580 585

Asn Pro Gly Gln Glu Leu Ser Phe Ser Met His Thr Val Ser His
590 595 600

Asp Gly Pro Val Ala Gly Leu Ala Phe Glu Leu Asn Glu Pro Ser
605 610 615

Ala Asp Val Ser Ser Ala Trp Ala Gln His Val Thr Lys Met Val
620 625 630

Ala Arg Arg Gly Ala Ile Leu Pro Gln Asp Val Ser Val Thr Pro
635 640 645

Val Ala Thr Pro Val Pro Pro Glu Glu Gln Ala Asn Leu Trp Leu
650 655 660

Val Glu Ala Glu Ile Ser Pro Glu Leu Glu Lys Arg Leu Gly Arg
665 670 675

Lys Lys Lys Arg Arg Lys Arg Lys Lys Glu Val Cys Pro Leu Gly
680 685 690

Pro Ala Pro Glu Leu His His Ser Ala Pro Val Pro Ala Thr Ser
695 700 705

Ala Val Pro Arg Leu Pro Gln Leu Pro Arg Gln Lys Cys Leu Val
710 715 720

Ala Ala Asn Ala Trp Gly Thr Gly Glu Pro Cys Arg Gln Gly Ala
725 730 735

Trp Thr Val Val Ser Asn Pro Phe Cys Pro Glu Pro Ser Pro His
740 745 750

Gln Asp Pro Phe Leu Pro Gly Ala Ser Ala Pro Arg Val Trp Ala
755 760 765

Gln Gly Arg Leu Gln Gly Leu Gly Ser Ile His Ser Arg Thr Asn
770 775 780

Leu Met Glu Ala Glu Leu Leu Asp Ala Asp Ser Asp Phe Glu Gln
785 790 795

Lys Leu Ile Ser Glu Glu Asp Leu
800 803

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35 40 45

Ser Ala Gly Gly Ser Ala Arg Arg Asn Ala Pro Val Thr Ser Pro
50 55 60

Pro Pro Pro Leu Leu Ser His Cys Gly Arg Ala Ala His Cys Glu
65 70 75

Pro Leu Arg Tyr Asn Val Cys Leu Gly Ser Ala Leu Pro Tyr Gly
80 85 90

Ala Thr Thr Thr Leu Leu Ala Gly Asp Ser Asp Ser Gln Glu Glu
95 100 105

Ala His Ser Lys Leu Val Leu Trp Ser Gly Leu Arg Asn Ala Pro
110 115 120

Arg Cys Trp Ala Val Ile Gln Pro Leu Leu Cys Ala Val Tyr Met
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Pro Lys Cys Glu Asn Asp Arg Val Glu Leu Pro Ser Arg Thr Leu
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Cys Gln Ala Thr Arg Gly Pro Cys Ala Ile Val Glu Arg Glu Arg
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Gly Trp Pro Asp Phe Leu Arg Cys Thr Pro Asp His Phe Pro Glu
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                245                 250                 255

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                260                 265                 270

Leu Phe Tyr Val Asn Ala Cys Phe Phe Val Gly Ser Ile Gly Trp
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Thr Ser Tyr Phe His Leu Leu Thr Trp Ser Leu Pro Phe Val Leu
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Thr Val Ala Ile Leu Ala Val Ala Gln Val Asp Gly Asp Ser Val
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Ser Gly Ile Cys Phe Val Gly Tyr Lys Asn Tyr Arg Tyr Arg Ala
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Cys Glu Ile Lys Asn Arg Pro Ser Leu Leu Val Glu Lys Ile Asn
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Leu Thr Gly His Ser Asp Asp Glu Pro Lys Arg Ile Lys Lys Ser
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Lys Met Ile Ala Lys Ala Phe Ser Lys Arg Arg Glu Leu Leu Gln
575 580 585

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Asp Gly Pro Val Ala Gly Leu Ala Phe Glu Leu Asn Glu Pro Ser
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Ala Arg Arg Gly Ala Ile Leu Pro Gln Asp Val Ser Val Thr Pro
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665 670 675

Lys Lys Lys Arg Arg Lys Arg Lys Lys Glu Val Cys Pro Leu Gly
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Gly Cys Pro Asn Glu Val Gln Asn Ile Lys Phe Asn Ser Ser Gly
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Gln Cys Glu Ala Pro Leu Val Arg Thr Asp Asn Pro Lys Ser Trp
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Leu Phe Tyr Val Asn Ala Cys Phe Phe Val Gly Ser Ile Gly Trp
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290 295 300

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Gly Phe Val Leu Ala Pro Ile Gly Leu Val Leu Ile Val Gly Gly
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Asn His Pro Gly Leu Leu Ser Glu Lys Ala Ala Ser Lys Ile Asn
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Cys Glu Ile Lys Asn Arg Pro Ser Leu Leu Val Glu Lys Ile Asn
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Trp Thr Lys Ala Thr Leu Leu Ile Trp Arg Arg Thr Trp Cys Arg
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Asn Pro Gly Gln Glu Leu Ser Phe Ser Met His Thr Val Ser His
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Lys Lys Lys Arg Arg Lys Arg Lys Lys Glu Val Cys Pro Leu Gly
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Pro Ala Pro Glu Leu His His Ser Ala Pro Val Pro Ala Thr Ser
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Ala Val Pro Arg Leu Pro Gln Leu Pro Arg Gln Lys Cys Leu Val
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Ala Ala Asn Ala Trp Gly Thr Gly Glu Pro Cys Arg Gln Gly Ala
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Trp Thr Val Val Ser Asn Pro Phe Cys Pro Glu Pro Ser Pro His
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Gln Asp Pro Phe Leu Pro Gly Ala Ser Ala Pro Arg Val Trp Ala
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50 55 60

Pro Ala Ala Arg Gly Cys Gly Pro Gly Arg Val Val Gly Ser Arg
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Arg Arg Pro Pro Arg Lys Leu Val Pro Leu Ala Tyr Lys Gln Phe
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95 100 105

Glu Gly Lys Ile Ala Arg Ser Ser Glu Arg Phe Lys Glu Leu Thr
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A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/12 C07K14/705 C12N1/19 C12N1/21 C12N5/10
C12N15/62 C07K19/00 C12N15/11 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N C07K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	TAKABATAKE T. ET AL.: "Hedgehog and patched gene expression in adult ocular tissues" FEBS LETTERS, vol. 410, 30 June 1997 (1997-06-30), pages 485-489, XP002101427 ISSN: 0014-5793 the whole document ---	1-15
A	MOTOYAMA J. ET AL.: "Ptch2, a second mouse Patched gene is co-expressed with Sonic hedgehog" NATURE GENETICS, vol. 18, February 1998 (1998-02), pages 104-106, XP002101696 ISSN: 1061-4036 cited in the application the whole document ---	1-15

-/--

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
19 July 1999	03/08/1999

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Mandl, B

2

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 97 45541 A (LELAND S STANFORD JUNIOR UNIVE ;UNIV CALIFORNIA (US)) 4 December 1997 (1997-12-04) the whole document ---	1-22, 26-28
E	WO 99 29854 A (ONTOGENY INC) 17 June 1999 (1999-06-17) the whole document ---	1-17, 19-21, 26-28
P,X	EP 0 879 888 A (SMITHKLINE BEECHAM PLC) 25 November 1998 (1998-11-25) the whole document ---	16-21,26
P,X	CARPENTER D. ET AL.: "Characterization of two patched receptors for the vertebrate hedgehog protein family" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 95, no. 23, 10 November 1998 (1998-11-10), pages 13630-13634 13634, XP002101428 ISSN: 0027-8424 the whole document ---	1-17
P,X	ZAPHIROPOULOS P. G. ET AL.: "PTCH2, a novel human patched gene undergoing alternative splicing and up-regulated in basal cell carcinomas." CANCER RESEACH, vol. 59, 15 February 1999 (1999-02-15), pages 787-792, XP002109560 the whole document -----	1-15

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 99/07417

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claims Nos.: 22-25
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 22-25

Present claims 19-25 relate to a compound defined by reference to a desirable characteristic or property, namely its antagonistic or agonistic effect on 'patched-2', respectively.

The claims cover all compounds having this characteristic or property, whereas the application provides support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT for only a very limited number of such compounds. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the compound by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, the search has been carried out for those parts of the claims which appear to be clear, supported and disclosed, namely those parts relating to antisense oligonucleotides which can be obtained from the specific sequence as it is presented by SEQ.ID.1 by using general methods known to the person skilled in the art. Consequently, claims 19 and 20 were searched partially and claim 21 was searched completely.
Moreover, there is an inconsistency between claim 21 referring to an 'antisense nucleotide' and page 9, line 1, referring to an 'antisense oligonucleotide'. Due to the fact that only the term 'antisense oligonucleotide' makes sense in the context of claim 21, claim 21 was read as referring to an 'antisense oligonucleotide'.

The applicant's attention is drawn to the fact that claims relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9745541 A	04-12-1997	AU 3227497 A	05-01-1998
WO 9929854 A	17-06-1999	NONE	
EP 0879888 A	25-11-1998	CA 2232808 A JP 11075874 A	23-11-1998 23-03-1999